

Supplementary Information

Selective and Cooperative Photocycloadditions within Multistranded Aromatic Sheets

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1. Materials and methods

1.1 Nuclear Magnetic Resonance

NMR spectra were recorded on 3 different NMR spectrometers: (1) an Avance II NMR spectrometer (Bruker Biospin, Wissembourg, France) with a vertical 7.05 T narrow-bore/ultrashield magnet operating at 300 MHz for ^1H spectra and 75 MHz for ^{13}C spectra by means of a 5mm BBFO $^1\text{H}/^{15}\text{N}$ - ^{31}P - ^{19}F probe with Z gradient capabilities; (2) a Avance III HD 400 NMR spectrometer (Bruker Biospin, Wissembourg, France) with a vertical 9.4 T narrow-bore/ultrashield magnet operating at 400 MHz for ^1H spectra and 100MHz for ^{13}C spectra by means of a 5mm Smartprobe BBFO $^1\text{H}/^{15}\text{N}$ - ^{31}P - ^{19}F probe with Z gradient capabilities; (3) an Avance NEO NMR spectrometer (Bruker Biospin, Wissembourg, France) with a vertical 16.45 T narrowbore/ultrashield magnet operating at 700 MHz for ^1H spectra by means of a 5mm TXI $^1\text{H}/^{13}\text{C}/^{15}\text{N}$ probe with Z gradient capabilities. Chemical shifts are reported in parts per million (ppm, δ) relative to the ^1H residual signal of the deuterated solvent used. ^1H NMR splitting patterns with observed first-order coupling are designated as singlet (s), doublet (d), triplet (t), or quartet (q). Coupling constants (J) are reported in hertz. Samples were not degassed otherwise it is specified. Data processing was performed with Topspin 4.0 software.

The assignment of NMR of the parallel aromatic sheets **1-7** and their respective photoproducts **1a-7a** and of remarkable resonances was performed using the following bi-dimensional NMR experiments.

TOCSY. Total Correlation Spectroscopy (TOCSY) experiments were recorded at 400 MHz. They were used to identify protons of oligomer that belong to the same spin system (e.g. aromatic ring) regardless of the exact topology with the following acquisition parameters: the acquisition was performed with $2048(t_2) \times 256(t_1)$ data points, relaxation delay of 2 s, and 32 scans per increment. Processing was done after a sine-bell multiplication in both dimensions and Fourier transformation in 1K x 1K real points.

HSQC. Heteronuclear Single-Quantum Correlation spectroscopy (HSQC) experiments were recorded at 400 MHz. They were used to observe correlations between nuclei of two different types which are separated by one bond with the following acquisition parameters: the acquisition was performed with $2048(t_2) \times 256(t_1)$ data points, relaxation delay of 2 s, and 64 scans per increment. Processing was done after a sine-bell multiplication in both dimensions and Fourier transformation in 1K x 1K real points.

HMBC. Heteronuclear Multiple Bond Correlation spectroscopy (HMBC) experiments were recorded at 400 MHz. They were used to detect heteronuclear correlations over longer ranges of about 2–4 bonds with the following acquisition parameters: the acquisition was performed with $2048(t_2) \times 256(t_1)$ data points, relaxation delay of 2 s, and 64 scans per increment. Processing was done after a sine-bell multiplication in both dimensions and Fourier transformation in 1K x 1K real points.

1.2 Photochemistry

Electronic absorption spectra were measured on a Cary 5000 UV-vis-NIR spectrophotometer. Steady-state emission spectra were recorded on a Horiba Jobin-Yvon Fluorlog-3 spectrofluorometer fitted with a Hamamatsu R928P PMT detector and exciting with a 450W Xe-lamp across a double monochromator and were corrected for instrumental response.

Photoirradiation experiments were carried out on solution samples in NMR tubes. Respective compounds were placed in NMR tubes, degassed and filled with argon. Subsequently, argon purged deuterated solvents were used to dissolve the compounds. The solutions were then subjected to irradiation by EXFO Lite (Model No. E3000-01) portable device having light source of 320 - 390 nm with 50 W mercury lamp. NMR were checked at different time intervals to follow the photoproduct formation. The experiments were repeated several times to establish reproducibility. Thermal reversibility experiments were performed in NMR tubes. The anthracene derivatives were dissolved in the appropriate deuterated solvent ($C_2D_2Cl_4$ or $CDCl_3$). The tube was heated to 393K or 333K for $C_2D_2Cl_4$ or $CDCl_3$, respectively. The kinetic reversibility was followed by 1H NMR.

1.3 Molecular mechanics

Molecular Models calculation were done using MacroModel version 8.6 (Schrödinger Inc.) with the Merck Molecular Force Field static (MMFFs) as implemented in this software. Energy minimized structures were obtained using 500 steps of Truncated Newton Conjugate Gradient (TNCG), chloroform as implicit solvent and the extended Cutoff option.

1.4 Crystallography

The diffraction data for compounds **8**, **10** and **13**-*sym* were collected at the IECB X-ray facility (CNRS UMS 3033 – INSERM US001, University of Bordeaux) with a Rigaku FRX rotating anode (2.9 kW) diffractometer using $CuK\alpha$ wavelength with a partial chi goniometer (AFC11). The X-ray source is equipped with high flux Osmic Varimax mirrors and a Dectris Pilatus 200K detector. Data were processed with the Rigaku Oxford Diffraction CrysAlisPro software (version 1.171.40.69a).^[1]

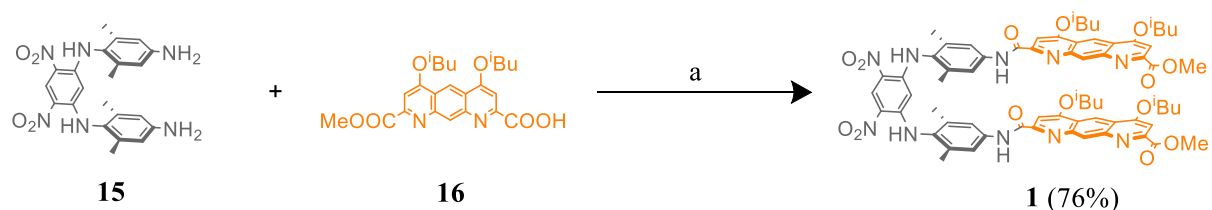
Data for compound **14**-*asym* were collected on a MetalJet (Excillum) X-ray source at the $GaK\alpha$ wavelength with a STOE Stadivari goniometer and a Dectris Eiger 1M detector. Data were processed with X-area version 1.87.

All structures were solved with Shelxt^[2] and refined by full-matrix least-squares method on F2 with Shelxl-2014^[2] within Olex2.^[3] Only non-H atoms of the backbones and side chains observable in the electron density maps were refined with anisotropic displacement parameters. H-atoms were refined in the riding-model approximation, with $U_{iso}(H)=1.2U_{eq}(CH, CH_2, NH)$. DFIX, AFIX, RIGU and SIMU restraints were applied to model geometry of the molecules and thermal motion parameters.

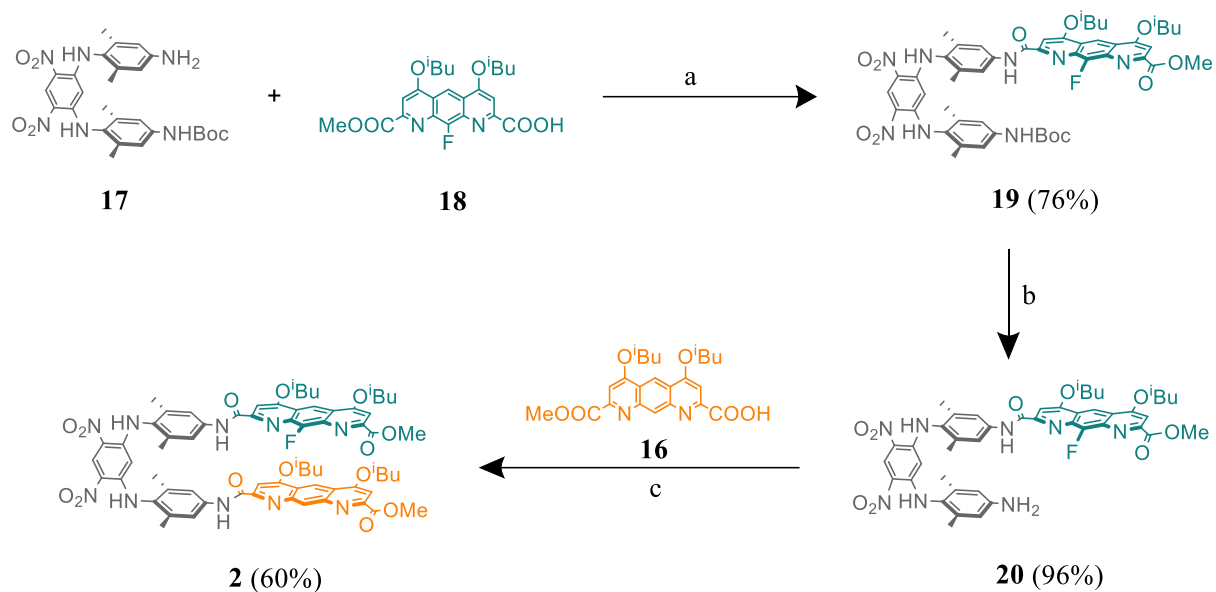
2. Methods for chemical synthesis

All reactions were carried out under a dry inert atmosphere, it is specified otherwise. Commercial reagents were purchased from Sigma Aldrich, TCI Chemicals or Alfa-Aesar and were used without further purification. Tetrahydrofuran (THF) and dichloromethane (CH_2Cl_2) were dried over alumina columns (MBRAUN SPS-800 solvent purification system); chloroform (CHCl_3) and diisopropylethylamine (DIPEA) were distilled over P_2O_5 and calcium hydride (CaH_2) respectively prior to use. Reactions were monitored by thin layer chromatography (TLC) on Merck silica gel 60-F254 plates and observed under UV light. Column chromatography purifications were carried out on Merck GEDURAN Si60 (40-63 μm). Preparative recycling GPC (gel permeation chromatography) were performed on JAIGEL 20*600 mm columns (Japan Analytical Industry) at a flow rate of 7 mL min^{-1} with a mobile phase composed of 1% (vol/vol) ethanol and 0.5% (vol/vol) Et_3N in chloroform. Monitoring was carried out by a UV detector at 254 nm, 280 nm, 300 nm and 360 nm. ESI mass spectra were obtained from the Mass Spectrometry Laboratory at the European Institute of Chemistry and Biology (UMS 3033 & US01 - IECB), Pessac, France.

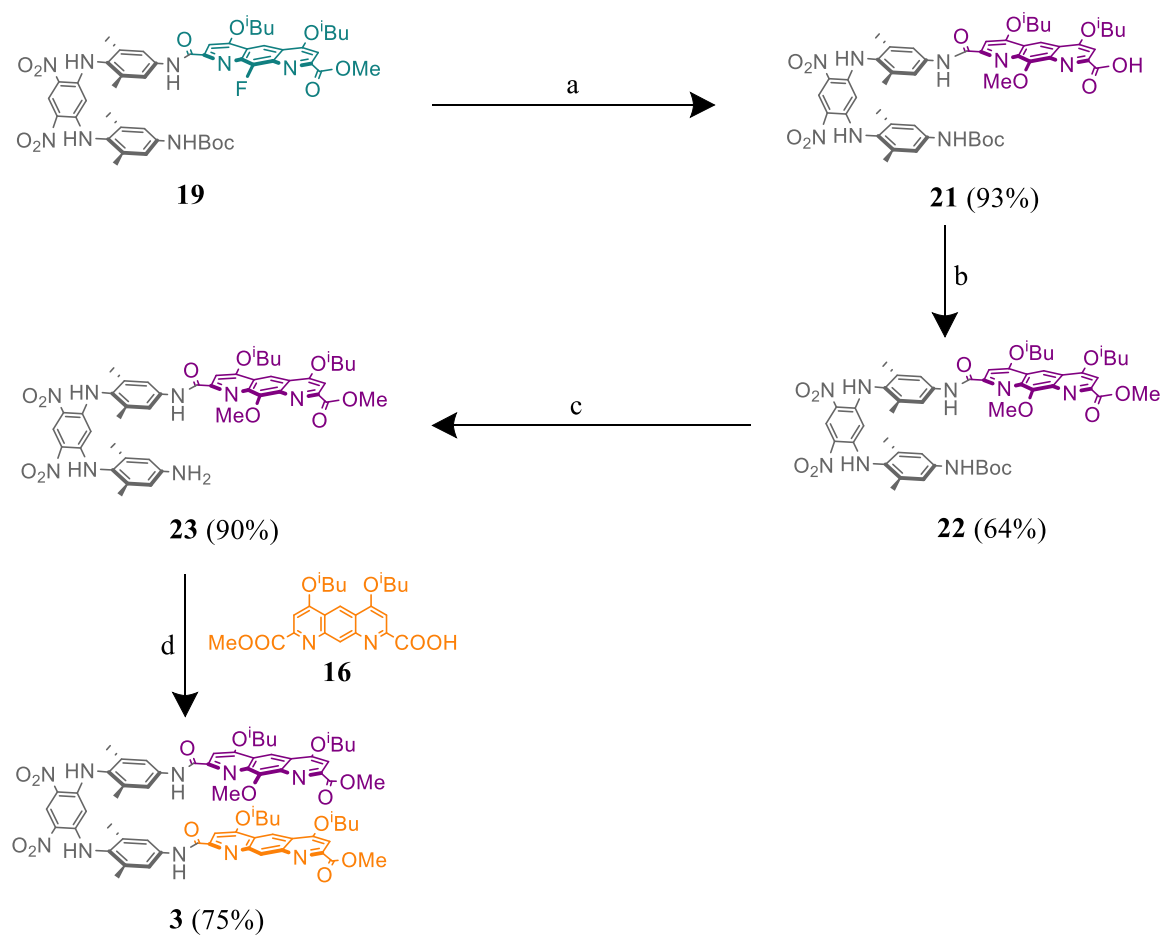
2.1 Synthesis of oligomers



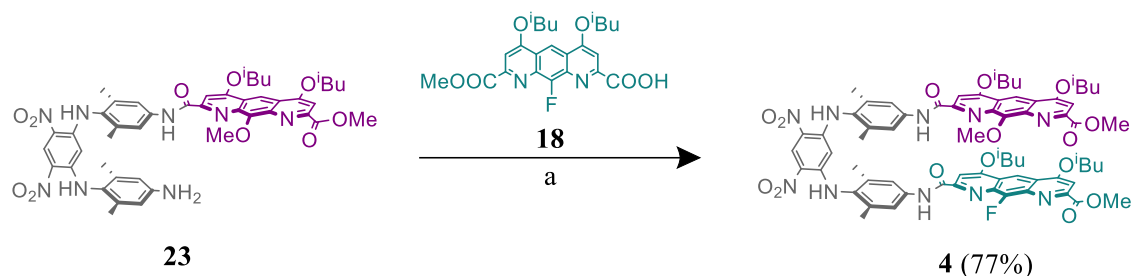
Scheme S1. Synthesis of **1**. (a) PyBOP, DIPEA, CHCl_3 , 16 hours, room temperature.^[4]



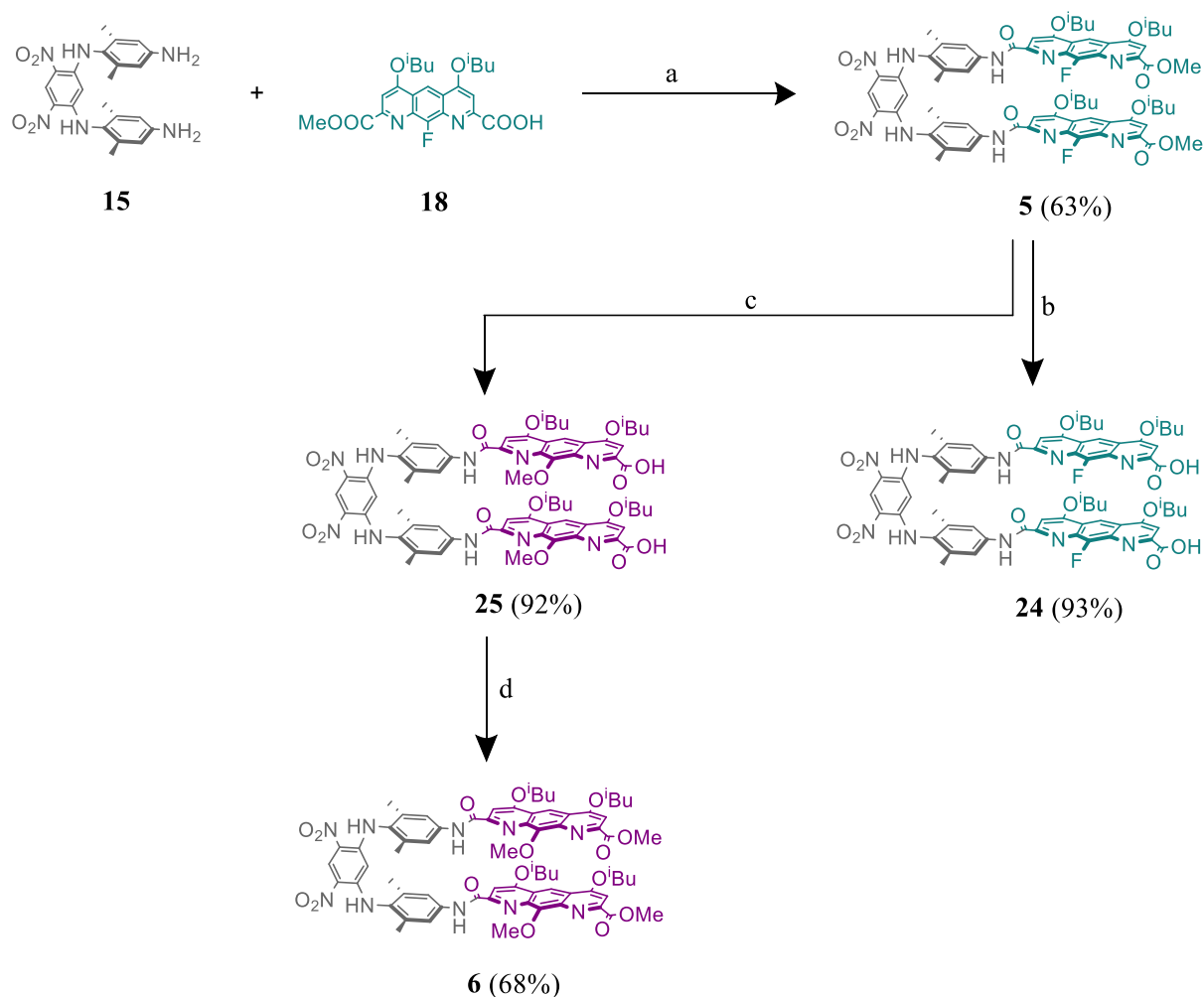
Scheme S2. Synthesis of **2**. (a) PyBOP, DIPEA, CHCl_3 , 16 hours, room temperature. (b) TFA, CHCl_3 , 4 hours, room temperature. (c) PyBOP, DIPEA, CHCl_3 , 16 hours, room temperature.



Scheme S3. Synthesis of **3**. (a) (i) NaOH, MeOH, THF, 12 hours, room temperature. (ii) Citric acid. (b) (i) 1-Chloro-*N,N*,2-trimethyl-1-propenylamine, CHCl₃, 2 hours, room temperature. (ii) MeOH, DIPEA, CHCl₃, overnight, room temperature. (c) TFA, DCM, 3 hours, room temperature. (d) PyBOP, DIPEA, CHCl₃, 16 hours, room temperature.

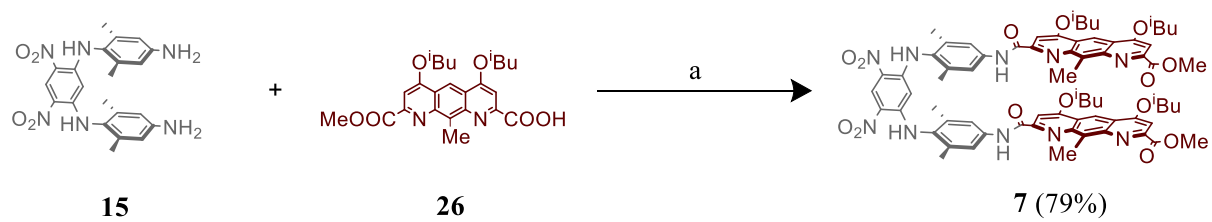


Scheme S4. Synthesis of **4**. (a) PyBOP, DIPEA, CHCl₃, 16 hours, room temperature.

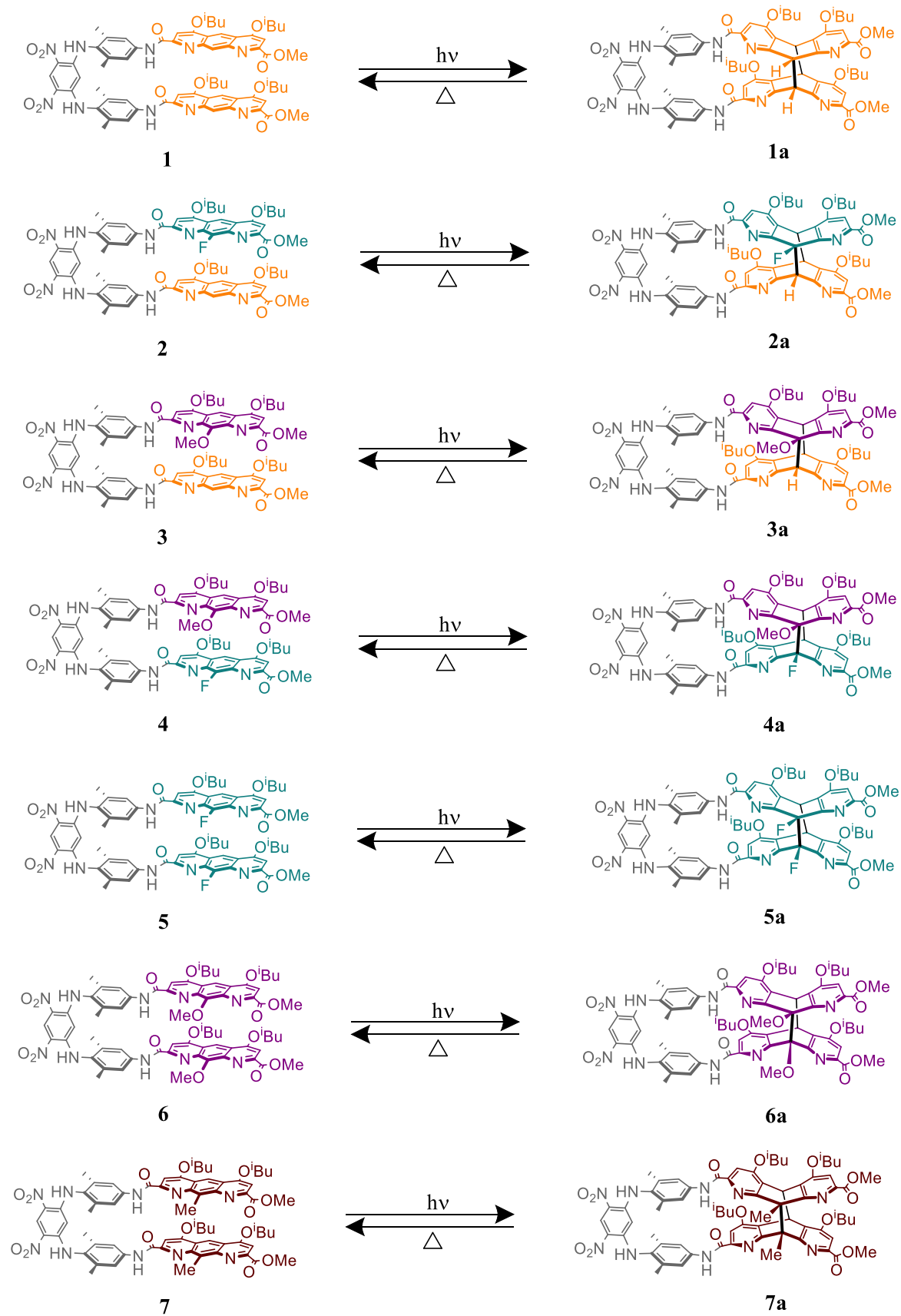


Scheme S5. Synthesis of **5** and **6**. (a) PyBOP, DIPEA, CHCl₃, 16 hours, room temperature. (b) (i) LiI, ethyl acetate, 3 hours, 351 K. (ii) Citric acid. (c) (i) NaOH, MeOH, THF, 12 hours, room temperature.

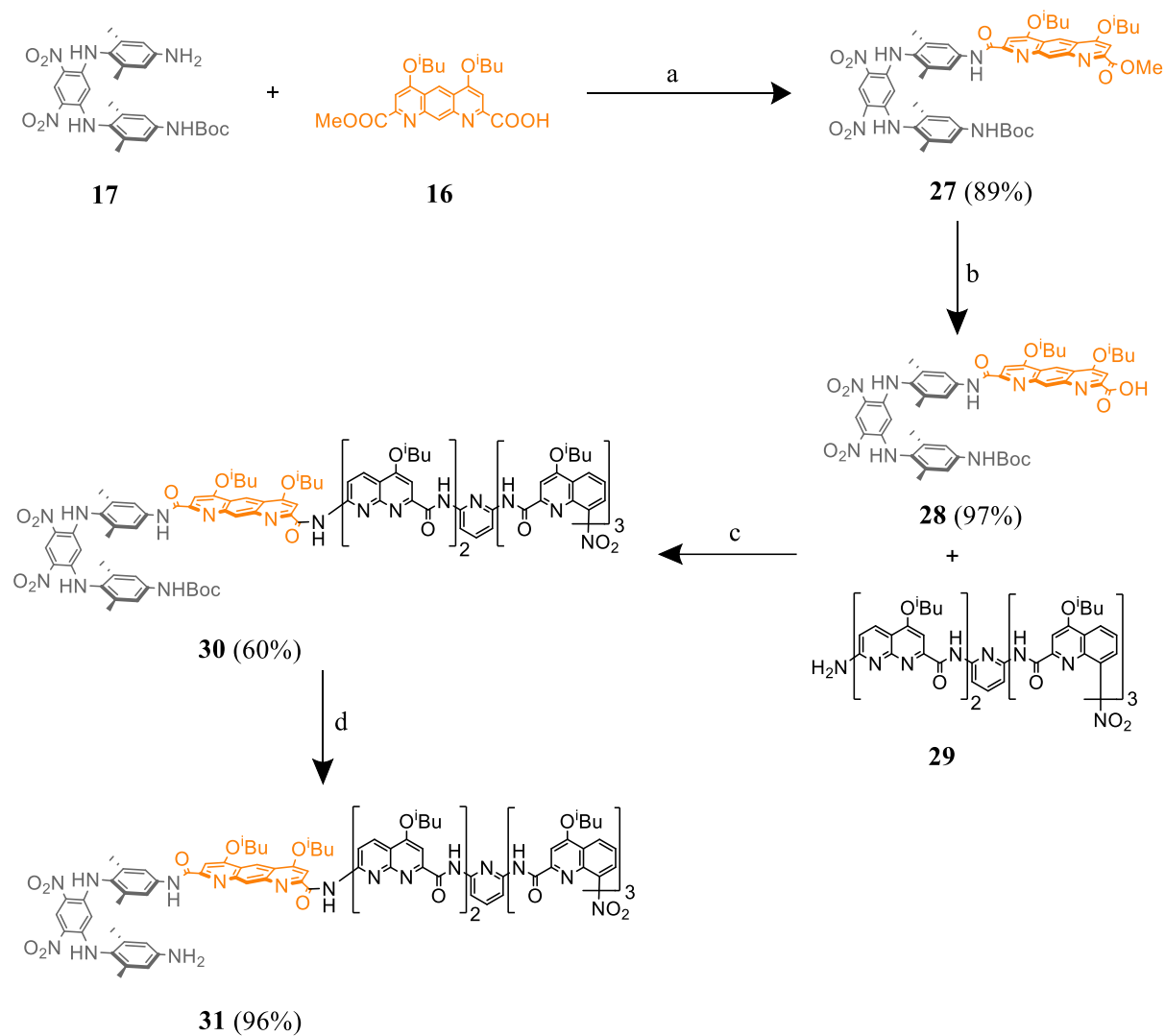
(ii) Citric acid. (d) $(\text{COCl})_2$, CHCl_3 , 2 hours, room temperature. (ii) MeOH , DIPEA , CHCl_3 , overnight, room temperature.



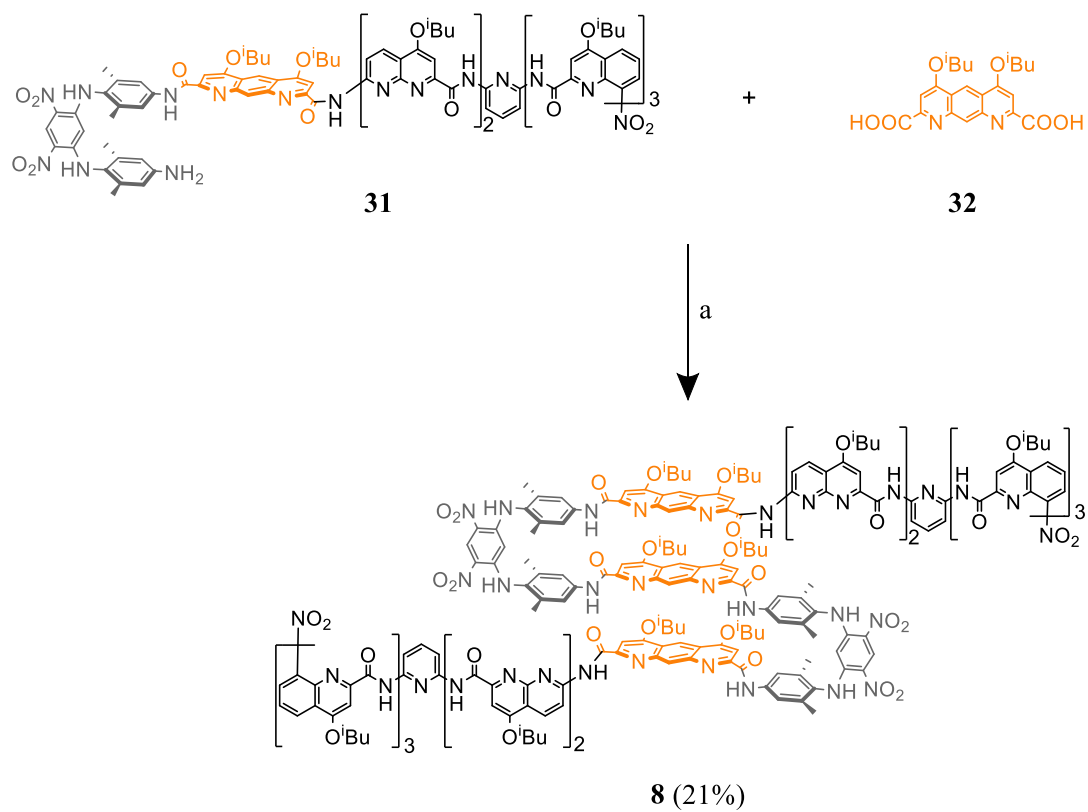
Scheme S6. Synthesis of 7. (a) PyBOP , DIPEA , CHCl_3 , 16 hours, room temperature.



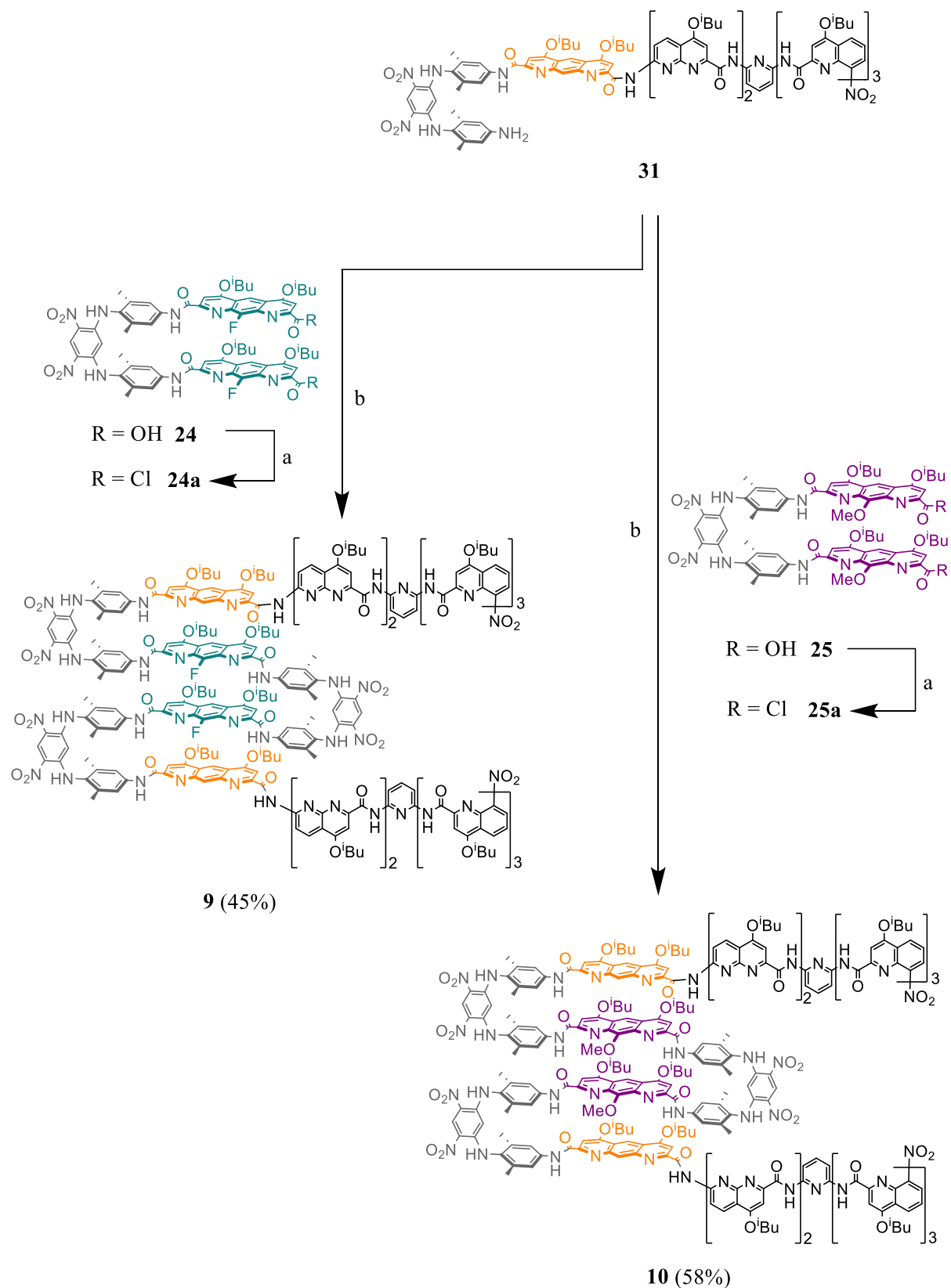
Scheme S7. Photoirradiation of **1-7** with a light source of 320 - 390 nm followed by the thermolysis at 393 K for 15 hours in tetrachloroethane- d_2 .



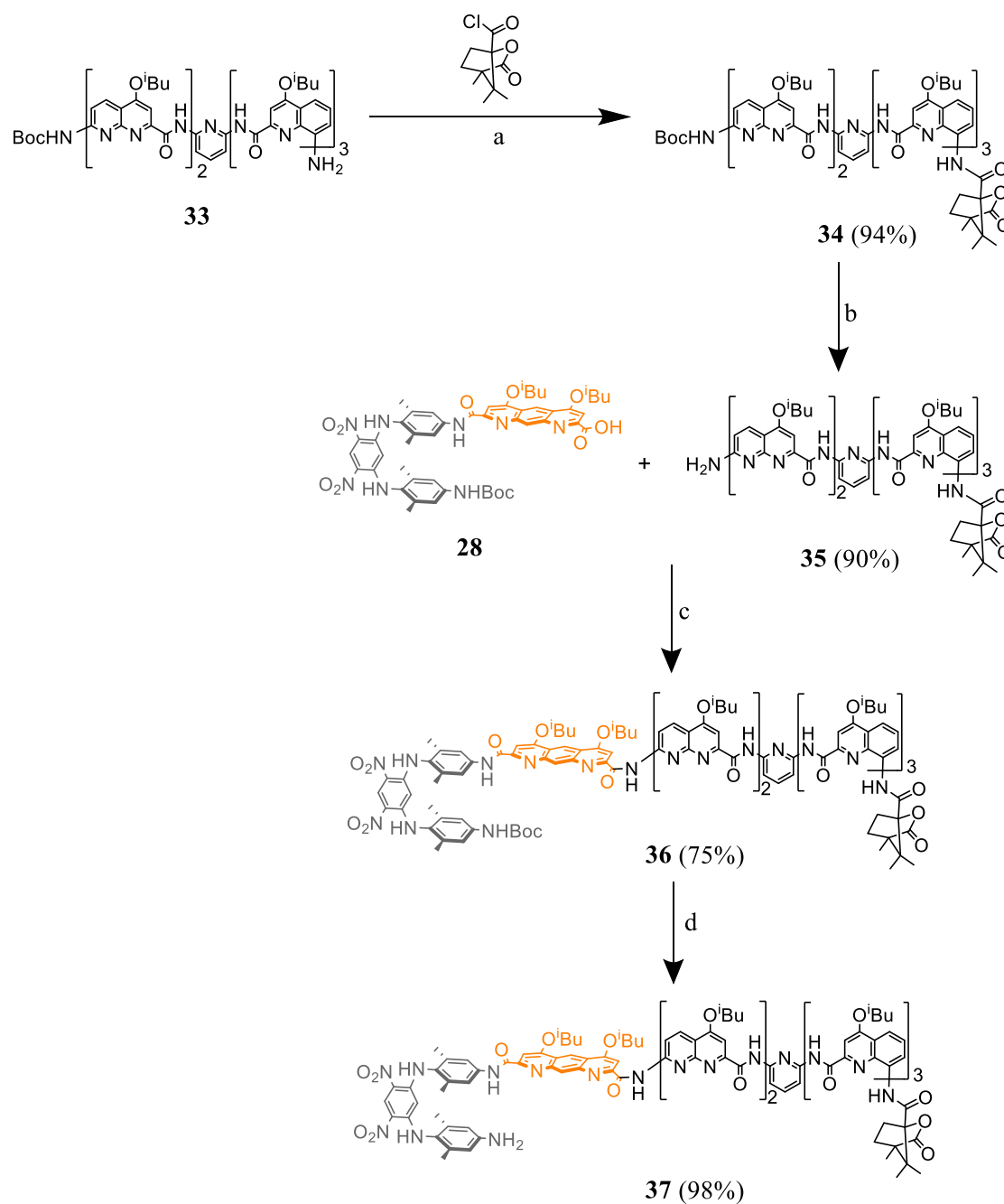
Scheme S8. Synthesis of **31**. (a) PyBOP, DIPEA, CHCl_3 , 16 hours, room temperature. (b) (i) NaOH, MeOH, THF, 12 hours, room temperature. (ii) Citric acid. (c) PyBOP, DIPEA, CHCl_3 , 16 hours, room temperature. (d) HCl (4M in dioxane), room temperature.



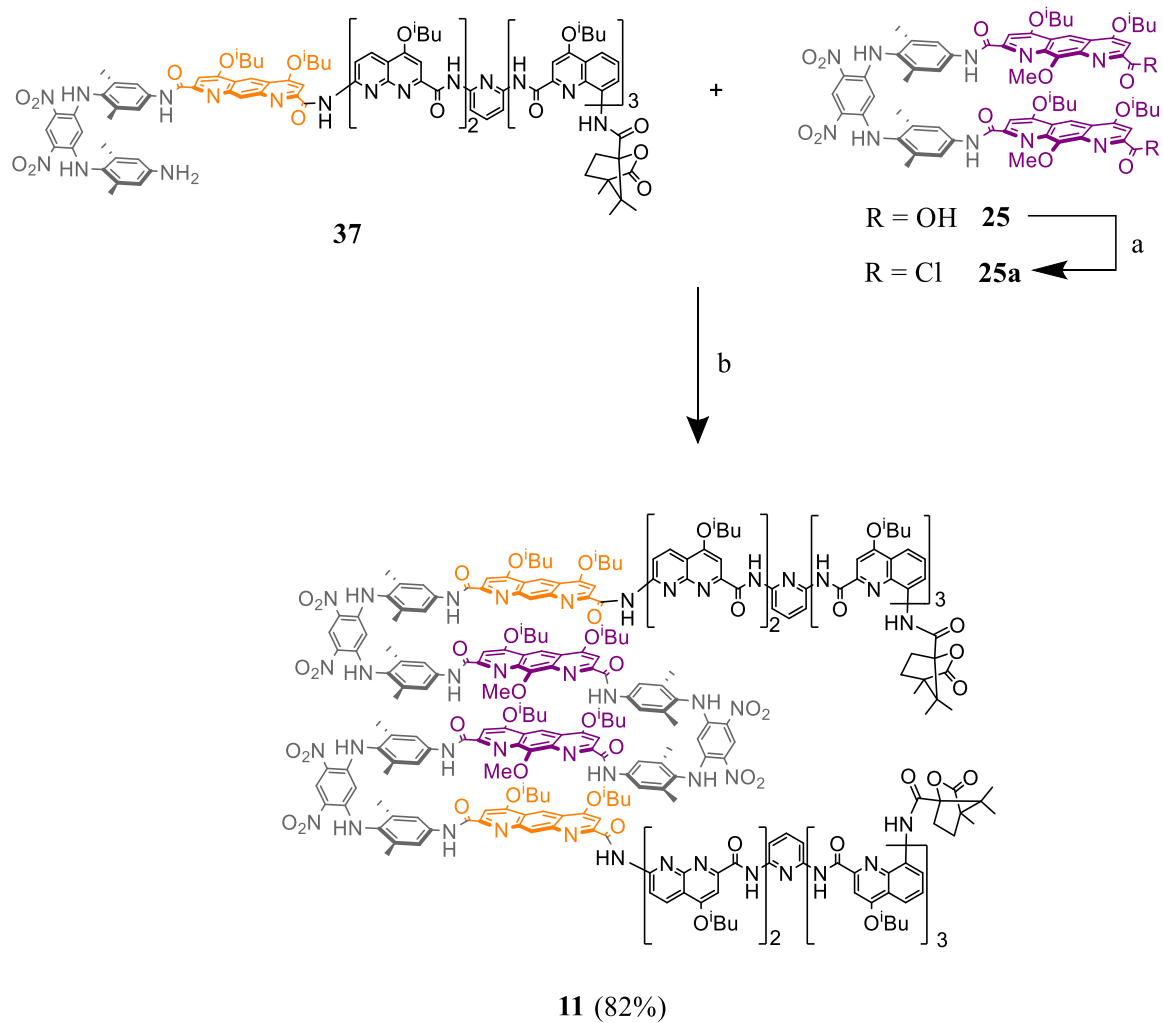
Scheme S9. Synthesis of **8**. (a) PyBOP, DIPEA, CHCl_3 , 2 days, 318 K.



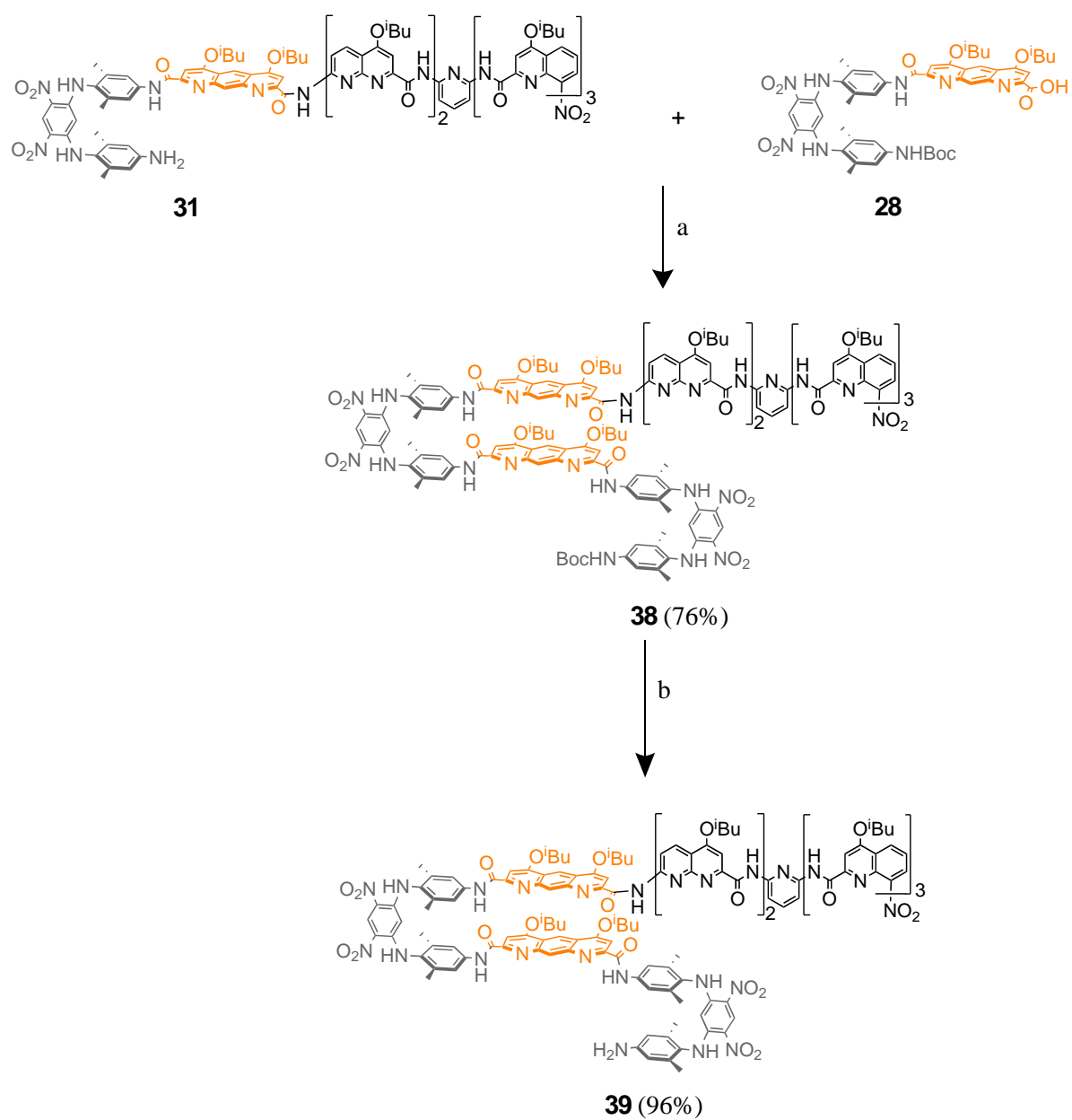
Scheme S10. Synthesis of **9** and **10**. (a) $(\text{COCl})_2$, CHCl_3 , 3 hours, room temperature. (b) DIPEA, CHCl_3 , 16 hours, room temperature.



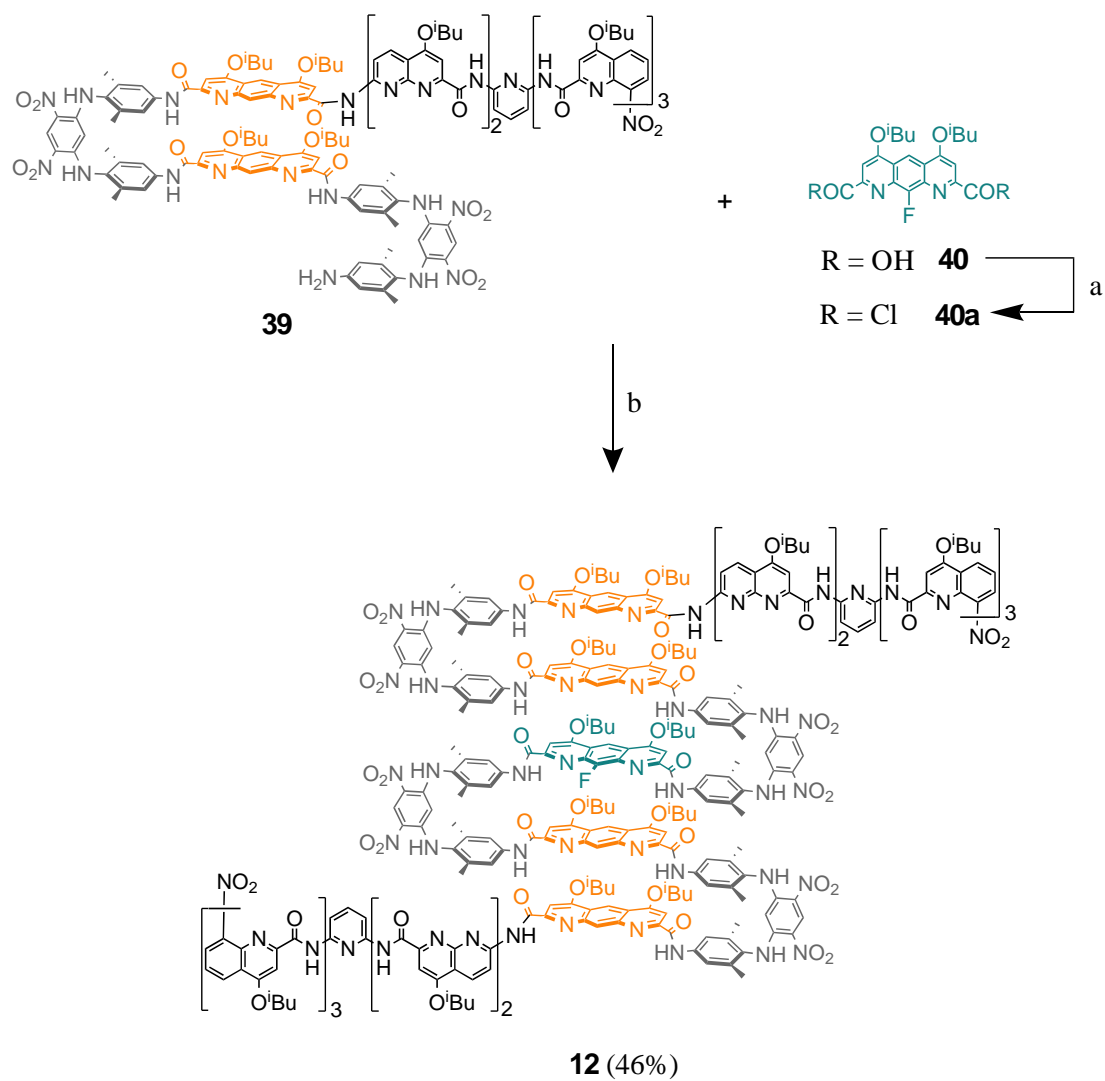
Scheme S11. Synthesis of **37**. (a) DIPEA, CHCl₃, 12 hours, room temperature. (b) TFA, CHCl₃, 4 hours, room temperature. (c) PyBOP, DIPEA, CHCl₃, 36 hours, room temperature. (d) HCl (4M in dioxane), room temperature.



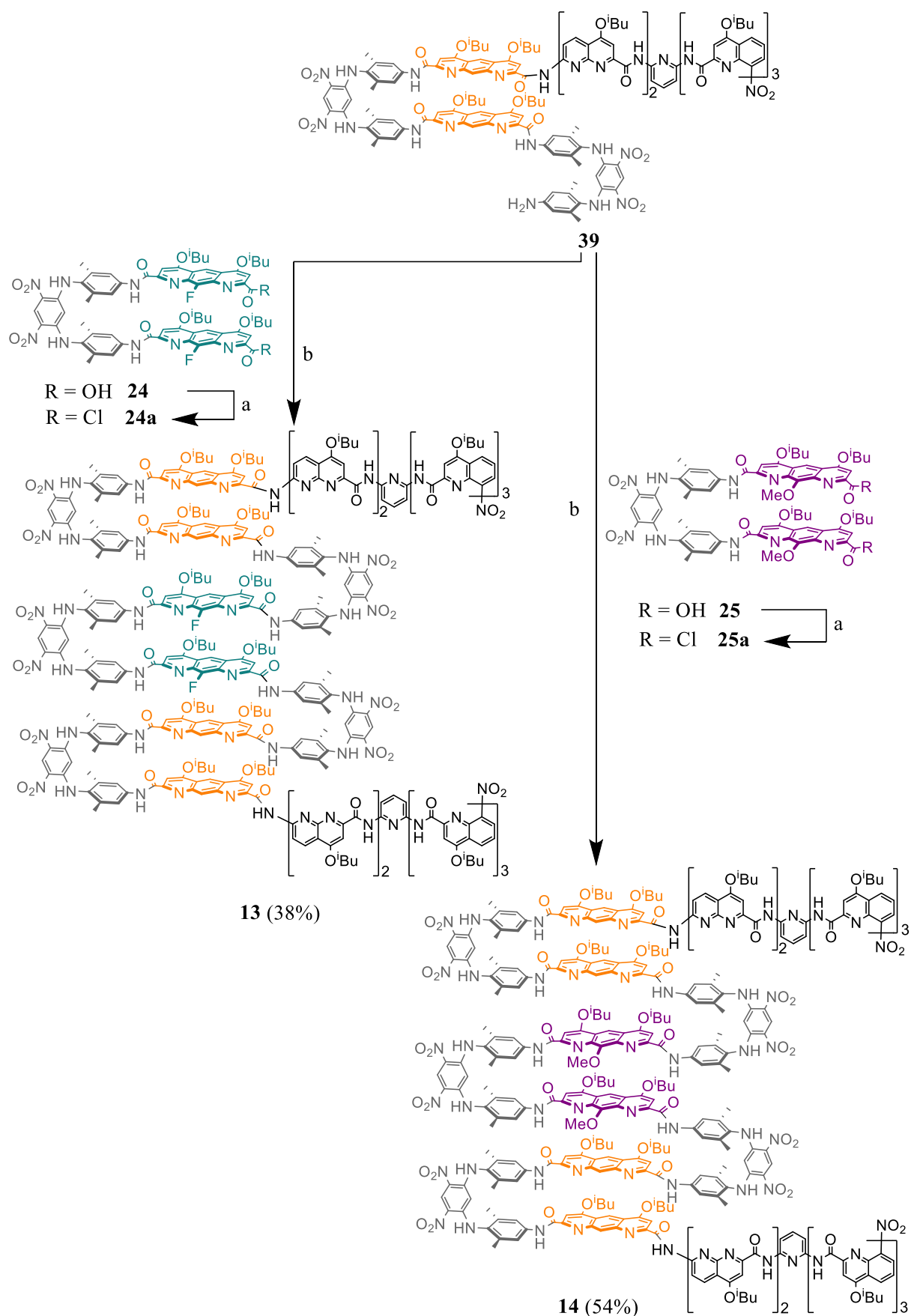
Scheme S12. Synthesis of **31**. (a) $(\text{COCl})_2$, CHCl_3 , 3 hours, room temperature. (b) DIPEA, CHCl_3 , 16 hours, room temperature.



Scheme S13. Synthesis of **39**. (a) PyBOP, DIPEA, CHCl₃, 16 hours, room temperature. (b) HCl (4M in dioxane), room temperature.

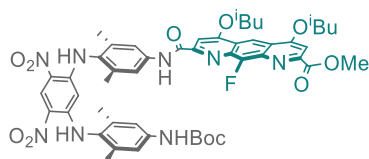


Scheme S14. Synthesis of **12**. (a) $(\text{COCl})_2$, CHCl_3 , 3 hours, room temperature. (b) DIPEA, CHCl_3 , 16 hours, room temperature.



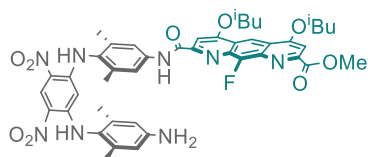
Scheme S15. Synthesis of **13** and **14**. (a) $(\text{COCl})_2$, CHCl_3 , 3 hours, room temperature. (b) DIPEA, CHCl_3 , 16 hours, room temperature.

2.3 Synthetic procedures



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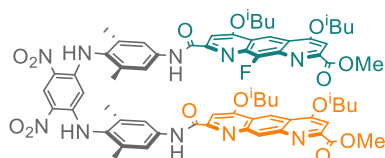
Tetramer 19. Monoamine turn **17**^[51] (0.097 g, 0.18 mmol), 9-fluorodiazanthracene **18**^[61] (0.089 g, 0.2 mmol), and PyBOP (0.187 g, 0.36 mmol) were placed in a 5 mL round bottom flask then dissolved in freshly distilled CHCl₃ (2 mL) and DIPEA (63 μ L, 0.36 mmol) was added under nitrogen atmosphere. The reaction mixture was allowed to stir at room temperature for 16 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane. The solution was then washed successively with 5% NH₄Cl, water and brine. Finally, the organic layer was dried over Na₂SO₄, filtered and evaporated. The residue was purified by recycling GPC and product **19** was obtained as a yellow solid (0.132 g, 76% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 10.19 (s, 1H), 9.35 (s, 1H), 9.16 (s, 1H), 9.12 (s, 1H), 9.06 (d, J = 2.2, 1H), 7.82 (s, 1H), 7.58 (s, 1H), 7.47 (s, 2H), 7.04 (s, 2H), 6.72 (s, 1H), 4.90 (s, 1H), 4.25 (d, J = 6.4 Hz, 2H), 4.19 (d, J = 6.4 Hz, 2H), 4.13 (s, 3H), 2.41 (m, 2H), 2.09 (s, 6H), 2.01 (s, 6H), 1.23 (d, J = 6.7 Hz, 12H), 1.15 (s, 9H). ¹³C NMR (75 MHz, CDCl₃, 298 K, δ ppm): 165.9, 163.8, 163.7, 163.3, 163.2, 162.1, 155.5, 153.3, 153.2, 152.7, 151.9, 151.6, 151.6, 148.4, 148.1, 138.0, 137.0, 136.6, 136.5, 136.3, 136.2, 135.2, 135.1, 130.9, 129.4, 129.2, 124.9, 124.9, 122.4, 122.1, 120.6, 118.9, 111.2, 111.1, 99.7, 97.5, 93.9, 80.0, 77.2, 75.6, 75.5, 53.6, 28.3, 28.3, 28.0, 19.2, 19.1, 18.1, 17.9. ¹⁹F NMR (282 MHz, CDCl₃, 298 K, δ ppm): -128.1. HRMS (ESI): m/z calcd for C₅₀H₅₆FN₈O₁₁ [M+H]⁺ 963.4047, Found 963.4115.



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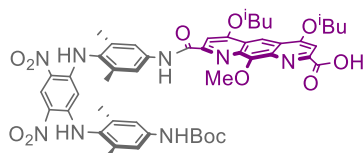
Tetramer amine 20. In a round-bottom flask, compound **19** (0.075 g, 0.078 mmol) was dissolved in 2 mL chloroform and subsequently, 2 mL TFA was added dropwise. The mixture was allowed to stir for 3 hours at room temperature. A saturated aqueous solution of NaHCO₃ was added to quench the excess acid. Then, the solution was extracted with CH₂Cl₂, washed with water, dried over Na₂SO₄ and evaporated. Product **19** was obtained without any further purification as a yellow solid (65 mg, 96% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 10.44 (s, 1H), 9.31 (s, 1H), 9.15 – 8.98 (m, 2H), 8.91 (s, 1H), 7.80 (s, 1H), 7.58 (s, 1H), 7.46 (s, 2H), 6.29 (s, 2H), 4.75 (s, 1H), 4.23 (d, J = 6.4 Hz, 2H), 4.19 (d, J = 6.4 Hz, 2H), 4.14 (s, 3H), 3.94 (s, 2H), 2.39 (m, 2H), 2.07 (s, 6H), 1.90 (s, 6H), 1.22 (d, J =

6.7 Hz, 12H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 165.5, 163.5, 163.53, 163.1, 163.0, 161.9, 154.6, 153.5, 153.5, 151.9, 151.4, 151.3, 148.8, 147.7, 146.2, 136.8, 136.8, 136.8, 136.6, 135.97, 135.9, 135.0, 134.9, 131.1, 128.9, 125.0, 124.8, 124.6, 122.2, 121.8, 120.6, 114.5, 111.2, 111.1, 99.5, 97.4, 94.3, 77.2, 77.0, 76.8, 76.5, 75.4, 75.3, 53.4, 29.5, 28.1, 28.0, 18.9, 17.7, 17.6, 17.5. HRMS (ESI): m/z calcd for $\text{C}_{45}\text{H}_{48}\text{FN}_8\text{O}_9$ $[\text{M}+\text{H}]^+$ 863.3528, Found 863.3587.



2

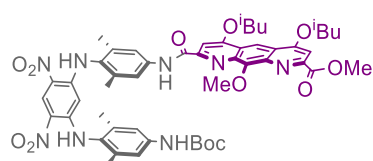
Sequence 2. Tetramer amine **20** (0.040 g, 0.046 mmol), monoacid **16**^[4] (0.02 g, 0.046 mmol) and PyBOP (0.120 g, 0.23 mmol) were placed in a 5 mL round-bottom flask filled with argon. Freshly distilled CHCl_3 (1 mL) and DIPEA (16 μL , 0.1 mmol) were then successively added. The solution was stirred at room temperature for 16 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane, washed with 5% NH_4Cl , distilled water and brine. The organic layer was dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude product was purified by recycling GPC and oligomer **2** was obtained as a yellow solid (0.095 g, 60% yield). ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 10.08 (s, 1H), 10.04 (s, 1H), 9.35 (s, 1H), 9.02 (d, $J = 2.2$ Hz, 2H), 8.58 (d, $J = 0.9$ Hz, 1H), 8.54 (d, $J = 0.9$ Hz, 1H), 8.23 (d, $J = 2.0$ Hz, 1H), 7.59 (d, $J = 4.1$ Hz, 4H), 7.40 (s, 1H), 7.36 (s, 1H), 7.31 (s, 1H), 7.18 (s, 1H), 4.76 (s, 1H), 4.09 (m, 10H), 3.82 (d, $J = 6.7$ Hz, 2H), 3.71 (d, $J = 6.7$ Hz, 2H), 2.33 (m, 2H), 2.12 (m, 14H), 1.20 (dd, $J = 6.7, 1.1$ Hz, 12H), 1.12 – 0.99 (m, 12H). ^{19}F NMR (282 MHz, CDCl_3 , 298 K, δ ppm): -127.2. ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 166.0, 165.9, 162.9, 162.8, 162.6, 161.7, 161.4, 153.2, 152.8, 151.0, 150.8, 148.4, 148.3, 147.1, 145.9, 137.6, 137.4, 137.3, 130.6, 130.6, 129.4, 125.0, 121.6, 121.4, 121.2, 121.1, 119.6, 119.5, 116.1, 110.1, 99.3, 98.8, 96.9, 96.5, 94.5, 75.4, 75.3, 75.1, 53.5, 53.5, 28.5, 28.3, 28.2, 19.3, 19.1, 19.1, 18.1. HRMS (ESI): m/z calcd for $\text{C}_{68}\text{H}_{72}\text{FN}_{10}\text{O}_{14}$ $[\text{M}+\text{H}]^+$ 1271.5214, Found 1271.5286.



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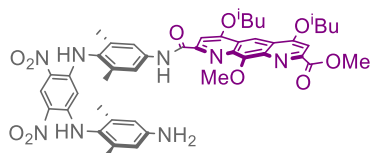
Tetramer acid 21. In a round bottom flask, compound **19** (0.096 g, 0.1 mmol) was dissolved in THF (4 mL). Sodium hydroxide (0.020 g, 0.5 mmol) dissolved in 1 mL methanol was then added dropwise. The reaction mixture was allowed to proceed at room temperature for 16 hours. Solvents were removed

under reduced pressure and the residue was suspended in water and acidified by 5% citric acid aqueous solution. The solid precipitate obtained was filtered and washed with water several times. Finally, dried over reduced pressure to get compound **21** as light brown solid which was used without further purification (0.89 g, 93% yield). $^1\text{H NMR}$ (300 MHz, CDCl_3 , 298 K, δ ppm): 10.21 (s, 1H), 9.36 (s, 1H), 9.18 (s, 1H), 9.14 (s, 1H), 9.06 (s, 1H), 7.81 (s, 1H), 7.65 (s, 1H), 7.44 (s, 2H), 7.04 (s, 2H), 6.75 (s, 1H), 4.92 (s, 1H), 4.57 (s, 3H), 4.23 (dd, $J = 6.4, 3.7$ Hz, 4H), 2.52 – 2.31 (m, 2H), 2.10 (s, 6H), 2.02 (s, 6H), 1.25 – 1.19 (m, 12H), 1.17 (s, 9H). $^{13}\text{C NMR}$ (75 MHz, CDCl_3 , 298 K, δ ppm): 165.9, 164.5, 163.9, 162.3, 152.9, 152.8, 149.7, 148.5, 148.2, 140.2, 138.1, 137.3, 136.7, 136.7, 131.1, 129.6, 129.3, 125.1, 125.0, 123.2, 122.3, 120.5, 119.0, 111.7, 97.6, 94.0, 80.2, 77.4, 76.3, 75.8, 64.7, 28.4, 28.4, 28.2, 19.3, 19.2, 18.2, 18.1. HRMS (ESI): m/z calcd for $\text{C}_{50}\text{H}_{57}\text{N}_8\text{O}_{12}$ $[\text{M}+\text{H}]^+$ 961.4096, Found 961.4161.



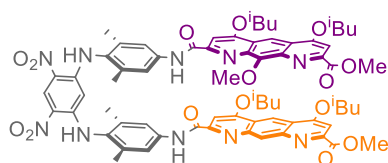
22

Tetramer 22. Tetramer acid **21** (0.085 g, 0.088 mmol) was dissolved in anhydrous CHCl_3 (1 mL), then 1-Chloro-N,N,2-trimethyl-1-propenylamine (60 μL , 0.47 mmol) was added and the reaction was allowed to stir at room temperature for 2 hours under argon. The solvent and excess reagent were removed under reduced pressure and the residue was dried under high vacuum for at least 3 hours to yield the corresponding acid chloride as a yellow solid. The residue was dissolved in 1 mL of dry chloroform and 1 mL freshly dried methanol was added dropwise. Finally, distilled DIPEA (80 μL , 0.47 mmol) was added. The reaction was allowed to stir at room temperature for 12 hours. Solvents were removed under reduced pressure and the residue was dissolved in CH_2Cl_2 before being washed with a saturated solution of NH_4Cl , water and brine. The organic layer was dried over Na_2SO_4 , filtered and evaporated. The crude product was purified by column chromatography using silica gel as stationary phase and $\text{DCM}:\text{MeOH}$ (99:1) as eluent to obtain **22** as yellow powder (55 mg, 64% yield). $^1\text{H NMR}$ (300 MHz, CDCl_3 , 298 K, δ ppm): 10.28 (s, 1H), 9.34 (s, 1H), 9.14 (s, 1H), 9.10 (s, 1H), 8.97 (s, 1H), 7.75 (s, 1H), 7.51 (s, 1H), 7.42 (s, 2H), 7.05 (s, 2H), 6.90 (s, 1H), 4.88 (s, 1H), 4.69 (s, 3H), 4.18 (dd, $J = 16.6, 6.3$ Hz, 4H), 4.09 (s, 3H), 2.38 (m, 2H), 2.08 (s, 6H), 2.01 (s, 6H), 1.21 (d, $J = 6.7$ Hz, 12H), 1.11 (s, 9H). $^{13}\text{C NMR}$ (75 MHz, CDCl_3 , 298 K, δ ppm): 166.4, 164.2, 163.6, 162.8, 153.5, 152.9, 152.0, 149.9, 148.5, 148.2, 140.6, 139.6, 138.2, 137.1, 136.9, 136.6, 131.0, 129.5, 129.2, 125.1, 125.0, 122.8, 122.6, 120.7, 119.0, 110.1, 99.3, 97.1, 94.1, 80.0, 75.5, 75.4, 64.7, 53.4, 28.4, 28.4, 28.1, 19.3, 19.3, 18.2, 18.1. HRMS (ESI): m/z calcd for $\text{C}_{51}\text{H}_{59}\text{N}_8\text{O}_{12}$ $[\text{M}+\text{H}]^+$ 975.4247, Found 975.4317.



23

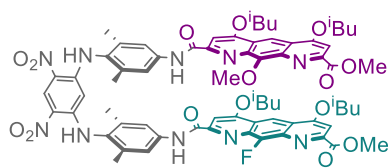
Tetramer amine 11. In a round-bottom flask, compound **22** (0.055 g, 0.056 mmol) was dissolved in 2 mL chloroform and subsequently, 2 mL TFA was added dropwise. The mixture was allowed to stir for 3 hours at room temperature. A saturated aqueous solution of NaHCO₃ was added to quench the excess acid. Then, the solution was extracted with CH₂Cl₂, washed with water, dried over Na₂SO₄ and evaporated. Product **23** was obtained without any further purification as a yellow solid (43 mg, 90% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 10.52 (s, 1H), 9.31 (s, 1H), 9.07 (s, 1H), 9.01 (s, 1H), 8.92 (s, 1H), 7.74 (s, 1H), 7.51 (s, 1H), 7.45 (s, 2H), 6.29 (s, 2H), 4.77 (s, 1H), 4.67 (s, 3H), 4.21 (d, *J* = 6.4 Hz, 2H), 4.16 (d, *J* = 6.3 Hz, 2H), 4.10 (s, 3H), 2.38 (m, 2H), 2.07 (s, 6H), 1.90 (s, 6H), 1.29 – 1.17 (m, 12H). ¹³C NMR (75 MHz, CDCl₃, 298 K, δ ppm): 166.4, 164.3, 163.7, 162.8, 153.5, 152.6, 150.1, 149.1, 148.1, 146.6, 140.7, 139.8, 137.2, 137.1, 131.3, 129.3, 125.4, 125.1, 124.9, 122.8, 122.6, 120.6, 114.8, 110.6, 99.3, 97.2, 94.6, 75.6, 75.4, 64.7, 53.4, 28.4, 28.4, 19.3, 18.1, 17.9. HRMS (ESI): *m/z* calcd for C₄₆H₅₁N₈O₁₀ [M+H]⁺ 875.3723, Found 875.3784.



3

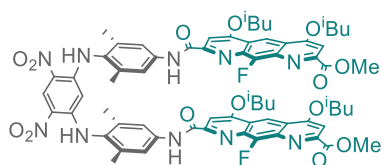
Sequence 3. Tetramer amine **23** (0.025 g, 0.028 mmol), monoacid **16**^[4] (0.02 g, 0.046 mmol) and PyBOP (0.075 g, 0.143 mmol) were placed in a 5 mL round-bottom flask filled with argon. Freshly distilled CHCl₃ (1 mL) and DIPEA (10 μL, 0.06 mmol) were then successively added. The solution was stirred at room temperature for 16 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane, washed with 5% NH₄Cl, distilled water and brine. The organic layer was dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was purified by recycling GPC and sequence **3** was obtained as a yellow solid (0.028 g, 75% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 10.29 (s, 1H), 10.05 (s, 1H), 9.34 (s, 1H), 9.02 (s, 2H), 8.77 (s, 1H), 8.55 (s, 1H), 8.20 (s, 1H), 7.57 (s, 4H), 7.37 (s, 1H), 7.35 (s, 1H), 7.21 (s, 1H), 6.94 (s, 1H), 4.77 (s, 4H), 4.09 (m, 10H), 3.63 (d, *J* = 6.7 Hz, 2H), 3.46 (d, *J* = 6.7 Hz, 2H), 2.34 (m, 2H), 2.11 (d, *J* = 2.1 Hz, 12H), 2.07 – 1.92 (m, 2H), 1.21 (dd, *J* = 6.7, 5.0 Hz, 12H), 0.99 (dd, *J* = 9.6, 6.7 Hz, 12H). ¹³C NMR (75 MHz, CDCl₃, 298 K, δ ppm): 166.3, 165.9, 163.0, 163.0, 162.9, 162.8, 161.9, 161.7, 153.1, 152.9, 151.3, 150.9, 149.2, 148.4, 147.0, 146.0, 139.9, 138.7, 137.7, 137.6, 137.3, 137.2, 130.5, 130.3, 129.4, 129.3, 125.0, 121.9, 121.7, 121.4, 121.2, 119.3, 119.3, 116.4, 108.8, 98.9, 98.8,

96.4, 96.2, 94.5, 77.4, 75.3, 75.2, 75.0, 74.7, 64.5, 53.4, 53.3, 28.5, 28.5, 28.2, 28.2, 19.3, 19.3, 19.1, 19.0, 18.1, 18.1. HRMS (ESI): m/z calcd for $C_{69}H_{75}N_{10}O_{15}$ $[M+H]^+$ 1283.5408, Found 1283.5504.



4

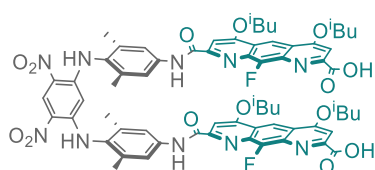
Sequence 4. Tetramer amine **23** (0.015 g, 0.017 mmol), monoacid **18**^[6] (0.015 g, 0.034 mmol) and PyBOP (0.044 g, 0.085 mmol) were placed in a 5 mL round-bottom flask filled with argon. Freshly distilled $CHCl_3$ (1 mL) and DIPEA (6 μ L, 0.034 mmol) were then successively added. The solution was stirred at room temperature for 16 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane, washed with 5% NH_4Cl , distilled water and brine. The organic layer was dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude product was purified by recycling GPC and sequencer **4** was obtained as a yellow solid (0.017 g, 77% yield). 1H NMR (300 MHz, $CDCl_3$, 298 K, δ ppm): 10.22 (s, 1H), 10.07 (s, 1H), 9.03 (s, 2H), 8.46 (d, J = 2.1 Hz, 1H), 8.12 (s, 1H), 7.58 (s, 2H), 7.56 (s, 2H), 7.38 (s, 2H), 7.35 (d, J = 1.0 Hz, 2H), 7.05 (s, 1H), 4.78 (m, 4H), 4.15 – 3.99 (m, 10H), 3.80 (d, J = 6.7 Hz, 2H), 3.67 (d, J = 6.7 Hz, 2H), 2.32 (m, 2H), 2.12 (m, 14H), 1.20 (t, J = 6.8 Hz, 12H), 1.05 (dd, J = 6.7, 5.3 Hz, 12H). ^{13}C NMR (75 MHz, $CDCl_3$, 298 K, δ ppm): 166.3, 165.7, 162.9, 162.7, 161.9, 161.5, 153.1, 152.9, 151.3, 150.7, 149.0, 148.4, 139.9, 138.6, 137.7, 137.5, 137.3, 137.2, 130.6, 130.4, 129.4, 125.0, 121.8, 121.5, 119.6, 119.4, 110.4, 110.3, 108.4, 99.2, 98.9, 97.0, 96.1, 94.5, 75.4, 75.3, 75.2, 74.8, 64.5, 53.4, 53.2, 28.5, 28.5, 28.3, 28.2, 19.3, 19.3, 19.1, 18.1, 18.1. HRMS (ESI): m/z calcd for $C_{69}H_{74}FN_{10}O_{15}$ $[M+H]^+$ 1301.5314, Found 1301.5408.



5

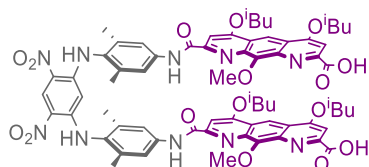
Sequence 5. Diamino turn **15**^[5] (0.1 g, 0.23 mmol), 9-fluorodiazaanthracene **18**^[6] (0.214 g, 0.48 mmol), and PyBOP (0.598 g, 1.15 mmol) were placed in a 5 mL round bottom flask then dissolved in freshly distilled $CHCl_3$ (2 mL) and DIPEA (0.2 mL, 1.15 mmol) was added under nitrogen atmosphere. The reaction mixture was allowed to stir at room temperature for 16 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane. The solution was then washed successively with 5% NH_4Cl , water and brine. Finally, the organic layer was dried over Na_2SO_4 , filtered, and evaporated. The residue was purified by recycling GPC and product **5** was obtained as a yellow

solid (0.187 g, 63% yield). ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 10.02 (s, 2H), 9.35 (s, 1H), 9.02 (s, 2H), 8.20 (d, $J = 2.0$ Hz, 2H), 7.58 (s, 4H), 7.42 (s, 2H), 7.37 (s, 2H), 4.76 (s, 1H), 4.15 – 4.01 (m, 10H), 3.86 (d, $J = 6.7$ Hz, 4H), 2.42 – 2.24 (m, 2H), 2.12 (m, 14H), 1.18 (d, $J = 6.7$ Hz, 12H), 1.08 (d, $J = 6.7$ Hz, 12H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 165.8, 162.6, 162.5, 162.5, 162.4, 161.4, 154.9, 152.8, 152.8, 151.3, 150.8, 150.7, 148.3, 137.2, 137.1, 135.7, 135.6, 134.4, 134.3, 130.6, 129.2, 124.9, 121.5, 121.3, 119.8, 109.9, 109.8, 99.2, 96.9, 94.5, 77.3, 75.3, 75.2, 53.4, 28.4, 28.1, 19.1, 19.0, 17.9. ^{19}F NMR (282 MHz, CDCl_3 , 298 K, δ ppm): -127.2. HRMS (ESI): m/z calcd for $\text{C}_{68}\text{H}_{71}\text{F}_2\text{N}_{10}\text{O}_{14}$ $[\text{M}+\text{H}]^+$ 1289.5114, Found 1289.5202.



24

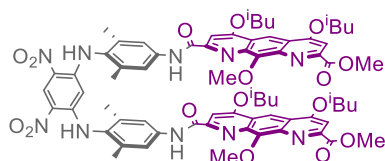
Pentamer diacid 24. In a round bottom flask, compound **5** (0.03 g, 0.023 mmol) was taken along with lithium iodide (0.025 g, 0.184 mmol) and freshly dried ethyl acetate (2 mL) was added under nitrogen atmosphere. The mixture was refluxed at 351 K for 3 hours before it cooled down to room temperature. Solvent was removed under reduced pressure, followed by diethyl ether was added to obtained precipitate. The precipitate was filtered, and 5% aqueous citric acid solution was added. Finally, the precipitate was washed with water (three times) and cold methanol followed by dried under vacuum to obtain yellow powder which was used without further purification (0.027 g, 93% yield). ^1H NMR (300 MHz, $\text{DMSO}-d_6$, 298 K, δ ppm): 10.14 (s, 2H), 9.43 (s, 2H), 9.06 (s, 1H), 8.20 (s, 2H), 7.65 (s, 4H), 7.26 (s, 2H), 7.10 (s, 2H), 4.63 (s, 1H), 4.11 (d, $J = 6.3$ Hz, 4H), 3.76 (s, 4H), 2.33 – 2.16 (m, 4H), 2.03 (s, 12H), 1.14 (d, $J = 6.6$ Hz, 12H), 1.04 (d, $J = 6.6$ Hz, 12H). ^{19}F NMR (282 MHz, CDCl_3 , 298 K, δ ppm): -129.7. HRMS (ESI): m/z calcd for $\text{C}_{66}\text{H}_{65}\text{F}_2\text{N}_{10}\text{O}_{14}$ $[\text{M}-\text{H}]^-$ 1259.4655, Found 1259.4611.



25

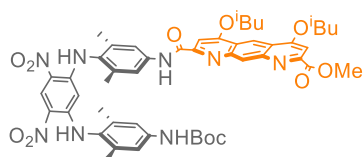
Pentamer diacid 25. In a round bottom flask, compound **5** (0.187 g, 0.145 mmol) was dissolved in THF (4 mL). Sodium hydroxide (0.035 g, 0.87 mmol) dissolved in 1 mL methanol was then added dropwise. The reaction mixture was allowed to proceed at room temperature for 16 hours. Solvents were removed under reduced pressure and the residue was suspended in water and acidified by 5% citric acid aqueous solution. The solid precipitate obtained was filtered, washed with water and diethyl ether several times and finally, dried over reduced pressure to get compound **25** as light brown solid which was used without

further purification (0.172 g, 92% yield). ^1H NMR (300 MHz, DMSO- d_6 , 298 K, δ ppm): 10.19 (s, 2H), 9.43 (s, 2H), 9.06 (s, 1H), 8.37 (s, 2H), 7.56 (s, 4H), 7.42 (s, 2H), 6.77 (s, 2H), 4.61 (m, 7H), 4.17 (d, $J = 6.2$ Hz, 4H), 3.16 (s, 4H), 2.38 – 2.19 (m, 4H), 2.03 (s, 12H), 1.16 (d, $J = 6.6$ Hz, 12H), 0.80 (d, $J = 6.6$ Hz, 12H). ^{13}C NMR (75 MHz, DMSO- d_6 , 298 K, δ ppm): 166.3, 162.4, 162.1, 160.8, 152.6, 150.5, 150.4, 147.4, 139.4, 138.0, 136.8, 136.6, 131.2, 124.0, 121.3, 121.0, 118.3, 108.2, 99.0, 95.3, 93.9, 74.6, 73.6, 64.9, 63.9, 27.8, 27.4, 18.8, 18.2, 17.6, 15.2. HRMS (ESI): m/z calcd for $\text{C}_{68}\text{H}_{73}\text{N}_{10}\text{O}_{16}$ $[\text{M}+\text{H}]^+$ 1285.5201, Found 1285.5295.



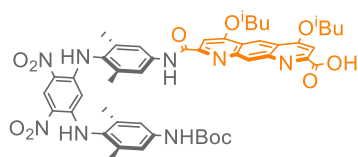
6

Sequence 6. Diacid **25** (0.025 g, 0.02 mmol) was dissolved in anhydrous CHCl_3 (1 mL), then oxalyl chloride (17 μL , 0.2 mmol) was added and the reaction was allowed to stir at room temperature for 3 hours under nitrogen atmosphere. The solvent and excess oxalyl chloride were removed under reduced pressure and the residue was dried under high vacuum for at least 2 hours to yield the corresponding acid chloride as a yellow solid. The residue was dissolved in 1 mL of dry chloroform and 1 mL freshly dried methanol was added dropwise. Finally, distilled DIPEA (14 μL , 0.08 mmol) was added. The reaction was allowed to stir at room temperature for 12 hours. Solvents were removed under reduced pressure and the residue was dissolved in CH_2Cl_2 before being washed with a saturated solution of NH_4Cl , water and brine. The organic layer was dried over Na_2SO_4 , filtered, and evaporated. The crude product was purified by recycling GPC and the sequence **6** was obtained as a yellow powder (19 mg, 68% yield). ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 10.20 (s, 2H), 9.35 (s, 1H), 9.04 (s, 2H), 8.40 (s, 2H), 7.53 (s, 4H), 7.40 (s, 2H), 7.01 (s, 2H), 4.79 (s, 1H), 4.76 (s, 6H), 4.11 (d, $J = 6.4$ Hz, 4H), 4.05 (s, 6H), 3.42 (d, $J = 6.6$ Hz, 4H), 2.35 (m, 2H), 2.12 (s, 12H), 1.95 (m, 2H), 1.21 (d, $J = 6.7$ Hz, 12H), 0.93 (d, $J = 6.7$ Hz, 12H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 166.1, 163.0, 162.8, 161.8, 152.7, 151.1, 149.0, 148.3, 139.7, 138.4, 137.7, 137.1, 130.2, 129.3, 124.9, 121.9, 121.8, 119.1, 108.7, 98.8, 96.1, 94.3, 77.3, 75.1, 74.6, 64.4, 53.2, 28.4, 28.0, 19.2, 18.9, 18.0. HRMS (ESI): m/z calcd for $\text{C}_{70}\text{H}_{77}\text{N}_{10}\text{O}_{16}$ $[\text{M}+\text{H}]^+$ 1313.5514, Found 1313.5614.



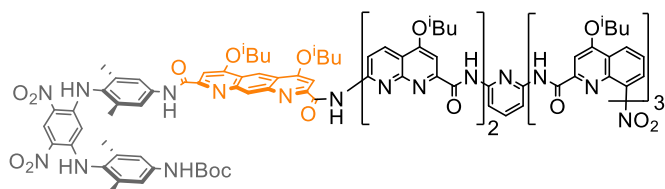
27

Tetramer 27. Monoamine turn **17**^[5] (0.2 g, 0.37 mmol), diazaanthracene monoacid **16**^[4] (0.19 g, 0.444 mmol), and PyBOP (0.481 g, 0.925 mmol) were placed in a 5 mL round bottom flask then dissolved in freshly distilled CHCl₃ (2 mL) and DIPEA (130 μL, 0.74 mmol) was added under nitrogen atmosphere. The reaction mixture was allowed to stir at room temperature for 48 hours. The mixture was dissolved in dichloromethane. The solution was then washed successively with 5% NH₄Cl, water and brine. Finally, the organic layer was dried over Na₂SO₄, filtered and evaporated. The crude product was subjected to column chromatography with silica gel as stationary phase and methanol/DCM (1:99) as eluent to obtain **27** as a yellow solid (0.313 g, 89% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 10.18 (s, 1H), 9.35 (s, 1H), 9.31 (d, *J* = 0.8 Hz, 1H), 9.15 (s, 1H), 9.12 (s, 1H), 9.07 (d, *J* = 0.9 Hz, 1H), 7.78 (s, 1H), 7.54 (s, 1H), 7.48 (s, 2H), 7.03 (s, 2H), 6.68 (s, 1H), 4.91 (s, 1H), 4.23 (d, *J* = 6.4 Hz, 2H), 4.18 (d, *J* = 6.4 Hz, 2H), 4.14 (s, 3H), 2.40 (m, 2H), 2.09 (s, 6H), 2.01 (s, 6H), 1.23 (d, *J* = 6.7 Hz, 12H), 1.14 (s, 9H). ¹³C NMR (75 MHz, CDCl₃, 298 K, δ ppm): 166.1, 163.9, 163.4, 162.3, 153.4, 152.7, 151.7, 148.4, 148.1, 147.7, 146.7, 138.0, 136.9, 136.7, 136.5, 130.7, 129.6, 129.4, 129.1, 124.9, 124.9, 122.0, 121.8, 120.3, 118.9, 117.2, 99.1, 96.9, 93.9, 80.0, 77.5, 77.3, 77.1, 76.6, 75.4, 75.3, 53.5, 28.3, 28.23, 28.0, 19.4, 19.2, 19.2, 18.0, 17.9. HRMS (ESI): *m/z* calcd for C₅₀H₅₇N₈O₁₁ [M+H]⁺ 945.4141, Found 945.4211.



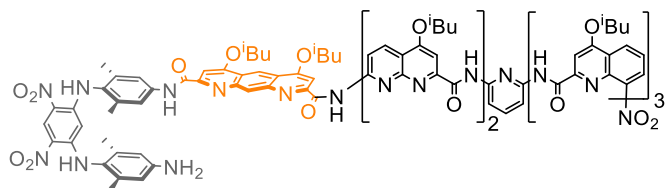
28

Tetramer acid 28. In a round bottom flask, compound **27** (0.2 g, 0.21 mmol) was dissolved in THF (3 mL). Sodium hydroxide (0.042 g, 1.05 mmol) dissolved in 1 mL methanol was then added dropwise. The reaction mixture was allowed to proceed at room temperature for 16 hours. Solvents were removed under reduced pressure and the residue was suspended in water and acidified by 5% citric acid aqueous solution. The solid precipitate obtained was filtered, washed with water and diethyl ether several times and finally, dried over reduced pressure to get compound **28** as brown solid which was used without further purification (0.190 g, 97% yield). ¹H NMR (300 MHz, DMSO-d₆, 298 K, δ ppm): 10.22 (s, 1H), 9.35 (d, *J* = 2.8 Hz, 2H), 9.17 (s, 1H), 9.13 (s, 1H), 9.03 (s, 1H), 7.81 (s, 1H), 7.65 (s, 1H), 7.50 (s, 2H), 7.05 (s, 2H), 6.75 (s, 1H), 4.93 (s, 1H), 4.23 (dd, *J* = 6.4, 4.9 Hz, 4H), 2.54 – 2.33 (m, 2H), 2.10 (s, 6H), 2.02 (s, 6H), 1.23 (dd, *J* = 6.7, 2.1 Hz, 12H), 1.16 (s, 9H). ¹³C NMR (75 MHz, DMSO-d₆, 298 K, δ ppm): 164.1, 163.3, 162.1, 154.5, 152.9, 151.8, 148.5, 148.2, 147.3, 138.2, 137.1, 136.8, 136.7, 131.0, 129.6, 129.3, 126.0, 125.1, 125.0, 122.5, 121.3, 120.4, 119.0, 118.4, 98.0, 97.6, 94.0, 80.1, 76.5, 74.6, 28.4, 28.4, 28.1, 19.3, 19.2, 18.2, 18.1. HRMS (ESI): *m/z* calcd for C₄₉H₅₅N₈O₁₁ [M+H]⁺ 931.3985, Found 931.4059.



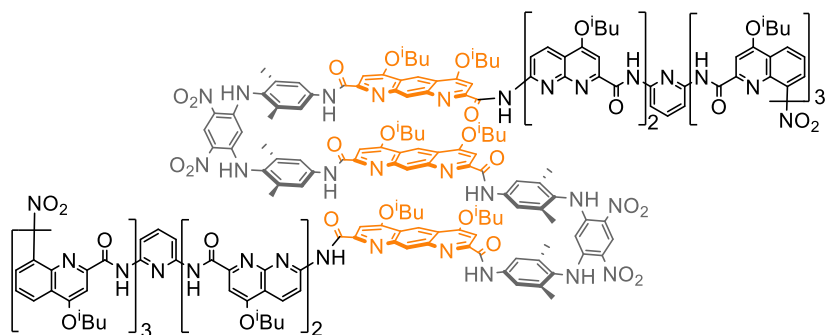
30

Oligomer 30. Monoacid **28** (0.072 g, 0.077 mmol), amino hexamer **29**^[7] (0.095 g, 0.07 mmol) and PyBOP (0.182 g, 0.35 mmol) were placed in a 5 mL round-bottom flask filled with argon. Freshly distilled CHCl_3 (1 mL) and DIPEA (60 μL , 0.35 mmol) were then successively added. The solution was stirred at room temperature for 16 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane, washed with 5% NH_4Cl , distilled water and brine. The organic layer was dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude product was purified by recycling GPC and oligomer **30** was obtained as a yellow solid (0.095 g, 60% yield). ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 11.90 (s, 1H), 11.85 (s, 1H), 11.61 (s, 1H), 11.59 (s, 1H), 9.99 (s, 1H), 9.76 (s, 1H), 9.60 (s, 1H), 9.30 (s, 1H), 8.99 (s, 1H), 8.98 (s, 1H), 8.92 (s, 1H), 8.90 – 8.76 (m, 4H), 8.63 (d, $J = 0.9$ Hz, 1H), 8.46 (dd, $J = 7.7, 1.3$ Hz, 1H), 8.36 (ddd, $J = 7.1, 5.6, 1.5$ Hz, 2H), 7.97 (d, $J = 7.8$ Hz, 1H), 7.87 (s, 1H), 7.86 (s, 1H), 7.84 (s, 1H), 7.79 (s, 1H), 7.71 (s, 1H), 7.32 (s, 1H), 7.31 – 7.28 (m, 1H), 7.24 – 7.16 (m, 2H), 7.11 (d, $J = 7.9$ Hz, 1H), 7.05 (s, 1H), 6.99 (dd, $J = 8.3, 1.3$ Hz, 1H), 6.79 (s, 1H), 6.76 – 6.66 (m, 2H), 6.50 (s, 1H), 6.41 – 6.32 (m, 2H), 6.23 (s, 1H), 4.43 – 4.14 (m, 7H), 3.94 (m, 4H), 3.70 (d, $J = 7.9$ Hz, 1H), 3.30 (d, $J = 8.6$ Hz, 2H), 2.40 (m, 7H), 1.71 (s, 4H), 1.46 (s, 4H), 1.33 (d, $J = 6.8$, 6H), 1.30 – 1.13 (m, 36H), 0.99 (s, 9H), 0.75 (d, $J = 6.6$ Hz, 3H), 0.51 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 164.6, 163.9, 163.6, 163.4, 162.8, 162.2, 161.8, 161.3, 160.9, 160.1, 154.7, 154.6, 154.1, 153.9, 153.7, 152.6, 152.6, 151.8, 151.7, 151.2, 149.2, 148.2, 148.0, 146.9, 145.7, 145.3, 144.6, 138.9, 138.8, 138.6, 138.0, 137.6, 136.5, 136.3, 136.2, 135.9, 134.6, 134.5, 134.4, 134.4, 129.6, 129.4, 129.1, 128.1, 126.4, 126.1, 125.7, 124.8, 124.8, 124.0, 123.4, 122.7, 121.5, 120.9, 120.7, 119.1, 118.9, 118.8, 118.8, 118.7, 118.6, 117.7, 117.0, 116.2, 115.6, 115.0, 115.0, 114.5, 114.4, 109.0, 108.4, 101.3, 99.3, 98.8, 98.1, 96.9, 96.1, 95.7, 93.8, 79.8, 77.5, 77.3, 77.1, 76.6, 76.0, 75.8, 75.4, 75.2, 74.9, 29.7, 28.4, 28.4, 28.3, 28.2, 28.2, 27.9, 27.8, 19.5, 19.3, 19.3, 19.2, 18.4, 18.2, 17.9, 17.7, 17.0. HRMS (ESI⁺): calcd. for $\text{C}_{122}\text{H}_{127}\text{N}_{23}\text{O}_{22}$ [$\text{M} + 2\text{H}$]²⁺ 1133.4774, found 1133.4861.



31

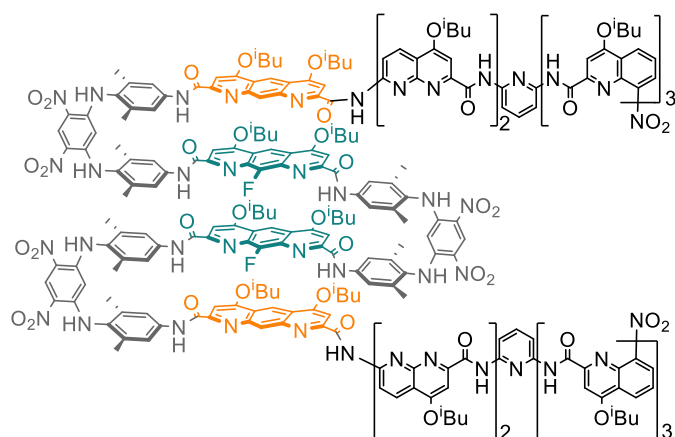
Oligomer 31. In a round-bottom flask, compound **30** (0.052 g, 0.023 mmol) was dissolved in HCl in dioxane (4M, 2mL) and chloroform (0.3 mL). The mixture was allowed to stir for 7 hours at room temperature. The reaction was quenched with saturated solution of NaHCO₃, and the solution was extracted three times with CH₂Cl₂, washed with water, dried over Na₂SO₄ and evaporated. Compound **31** was obtained as yellow solid which was used without any further purification (0.048 g, 96% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 11.92 (s, 1H), 11.82 (s, 1H), 11.65 (s, 1H), 11.60 (s, 1H), 10.04 (s, 1H), 9.75 (s, 1H), 9.69 (s, 1H), 9.24 (s, 1H), 8.95 – 8.87 (m, 2H), 8.87 – 8.77 (m, 4H), 8.75 (s, 1H), 8.62 (s, 1H), 8.47 (d, *J* = 7.6 Hz, 1H), 8.42 (d, *J* = 7.7 Hz, 1H), 8.37 (dd, *J* = 8.0, 1.7 Hz, 1H), 7.93 (s, 1H), 7.90 (s, 1H), 7.87 (s, 1H), 7.85 (s, 1H), 7.83 (s, 1H), 7.69 (s, 1H), 7.37 (s, 1H), 7.21 (ddd, *J* = 11.4, 5.6, 2.8 Hz, 4H), 7.04 (s, 1H), 6.97 (d, *J* = 8.3 Hz, 1H), 6.77 (t, *J* = 8.0 Hz, 1H), 6.45 (s, 1H), 6.37 (t, *J* = 8.0 Hz, 1H), 6.08 (s, 1H), 5.97 – 5.85 (m, 2H), 4.44 – 4.14 (m, 6H), 4.07 – 3.60 (m, 8H), 3.37 – 3.17 (m, 4H), 2.40 (m, 4H), 1.99 (s, 3H), 1.57 (d, *J* = 8.2 Hz, 4H), 1.44 (s, 4H), 1.33 (dd, *J* = 6.7, 3.8 Hz, 4H), 1.29 – 1.08 (m, 36H), 0.77 (d, *J* = 6.5 Hz, 3H), 0.55 (d, *J* = 6.7 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃, 298 K, δ ppm): 165.6, 164.7, 164.0, 163.7, 163.4, 162.9, 162.2, 161.9, 161.5, 160.9, 160.1, 154.6, 154.2, 154.0, 153.7, 152.6, 152.2, 151.9, 151.3, 149.3, 148.8, 148.2, 148.0, 146.9, 146.1, 145.7, 145.4, 144.7, 139.1, 139.0, 138.9, 138.0, 136.8, 136.4, 136.2, 134.7, 134.6, 134.3, 130.0, 129.8, 129.3, 128.8, 128.2, 126.5, 126.2, 125.8, 124.8, 124.1, 123.5, 1222.8, 121.5, 120.9, 120.7, 118.6, 118.3, 117.8, 117.1, 116.5, 115.6, 115.1, 114.5, 109.2, 108.5, 101.4, 99.3, 98.8, 98.0, 96.8, 96.3, 95.9, 94.2, 77.4, 76.1, 75.9, 75.5, 74.9, 30.2, 29.8, 28.5, 28.4, 28.3, 28.2, 27.9, 19.8, 19.4, 19.2, 18.6, 18.3, 14.3. HRMS (ES⁺): *m/z* calcd for C₁₁₇H₁₁₉N₂₃O₂₀ [M+2H]²⁺ 1083.4512 Found 1083.4587.



8

Oligomer 8. 1,8-diaza-4,5-diisobutoxy-2,7-anthracene dicarboxylate **32** (4.54 mg, 0.011 mmol), oligomer **31** (50 mg, 0.023 mmol) and PyBOP (172 mg, 0.33 mmol) were placed in a 5 mL round-bottom flask filled with argon. Freshly distilled CHCl₃ (1 mL) and DIPEA (11 μL, 0.066 mmol) were then successively added. The solution was stirred at 318 K for two days. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane, washed with 5% NH₄Cl, distilled water and brine. The organic layer was dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The crude product was purified by recycling GPC and oligomer **8** was obtained as a yellow

solid (11 mg, 21% yield). ^1H NMR (700 MHz, CDCl_3 , 298 K, δ ppm): 11.54 (s, 2H), 10.97 (s, 2H), 10.90 (s, 2H), 10.00 (s, 2H), 9.89 (s, 2H), 9.85 (s, 2H), 9.27 (s, 2H), 9.05 (d, $J = 11.8$ Hz, 4H), 8.89 (d, $J = 8.3$ Hz, 2H), 8.85 (s, 2H), 8.74 (d, $J = 8.3$ Hz, 2H), 8.64 (s, 1H), 8.58 (d, $J = 7.5$ Hz, 4H), 8.47 – 8.39 (m, 3H), 8.22 (d, $J = 6.9$ Hz, 2H), 8.19 (d, $J = 6.9$ Hz, 2H), 8.04 (s, 2H), 7.95 (d, $J = 7.1$ Hz, 2H), 7.74 (s, 2H), 7.57 (d, $J = 6.9$ Hz, 2H), 7.34 (s, 2H), 7.18 (s, 2H), 7.15 (t, $J = 7.1$ Hz, 2H), 7.09 – 7.01 (m, 5H), 6.91 – 6.85 (m, 5H), 6.83 (s, 2H), 6.74 (d, $J = 7.6$ Hz, 2H), 6.39 (s, 1H), 6.27 (d, $J = 5.3$ Hz, 3H), 6.23 (s, 2H), 6.06 (t, $J = 7.3$ Hz, 2H), 5.80 (s, 2H), 5.64 (s, 2H), 4.28 (t, $J = 5.5$ Hz, 3H), 4.23 (s, 3H), 4.18 (t, $J = 6.2$ Hz, 3H), 4.15 (t, $J = 5.7$ Hz, 2H), 4.10 (q, $J = 5.8, 5.3$ Hz, 4H), 4.05 (t, $J = 6.5$ Hz, 3H), 3.99 (t, $J = 6.8$ Hz, 2H), 3.92 (t, $J = 6.5$ Hz, 2H), 3.85 (m, 4H), 3.77 (t, $J = 6.1$ Hz, 3H), 3.66 (t, $J = 5.6$ Hz, 2H), 3.48 (t, $J = 7.2$ Hz, 2H), 3.25 (d, $J = 5.5$ Hz, 2H), 3.17 (q, $J = 3.2$ Hz, 2H), 2.78 – 2.71 (m, 2H), 2.43 (m, 7H), 2.37 – 2.27 (m, 7H), 2.27 – 2.16 (m, 6H), 1.97 (s, 6H), 1.81 (d, $J = 8.3$ Hz, 7H), 1.40 (d, $J = 7.0$ Hz, 8H), 1.34 (m, 16H), 1.32 – 1.23 (m, 24H), 1.19 (m, 20H), 1.16 – 1.04 (m, 20H), 0.91 – 0.78 (m, 6H), 0.47 (d, $J = 7.0$ Hz, 6H), 0.25 (d, $J = 6.5$ Hz, 6H). HRMS (ES⁺): m/z calcd for $\text{C}_{256}\text{H}_{257}\text{N}_{48}\text{O}_{44}$ $[\text{M}+3\text{H}]^{3+}$ 1569.6466 Found 1569.6572.

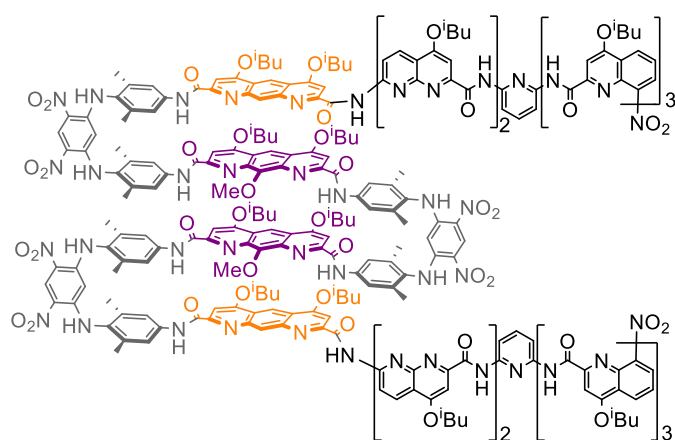


9

Oligomer 9. Dicarboxylate **24** (10.1 mg, 8 μmol) was suspended in anhydrous CHCl_3 (1 mL), then oxalyl chloride (7 μL , 80 μmol) was added and the reaction was allowed to stir at room temperature for 2 hours. The solvent and excess oxalyl chloride were removed under reduced pressure and the residue was dried under high vacuum for 3 hours to yield the corresponding acid chloride **24a** as a yellow solid. The solution of amine **31** (38 mg, 17 μmol) and distilled DIPEA (7 μL , 40 μmol) in anhydrous CHCl_3 (1 mL) was added dropwise via a syringe to solution of the freshly prepared **24a** dissolved in anhydrous CHCl_3 (0.5 mL). The reaction was allowed to proceed at room temperature for 16 hours. After evaporation of the solvents, the crude product was purified by recycling GPC. Oligomer **9** was obtained as a yellow product (20 mg, 45% yield). ^1H NMR (700 MHz, Acetone- d_6 , 258 K, δ ppm; only peaks are given due to the presence of two conformers): 11.61, 11.48, 11.32, 11.18, 11.11, 10.47, 10.28, 10.05, 9.93, 9.81, 9.73, 9.51, 9.47, 9.46, 9.45, 9.38, 9.29, 9.26, 9.21, 9.20, 9.11, 9.06, 9.03, 8.98, 8.95, 8.94, 8.91, 8.90, 8.85, 8.84, 8.76, 8.75, 8.75, 8.74, 8.73, 8.60, 8.59, 8.57, 8.53, 8.52, 8.47, 8.35, 8.29, 8.28,

8.27, 8.23, 8.22, 8.21, 8.19, 8.18, 8.01, 7.93, 7.92, 7.90, 7.89, 7.89, 7.88, 7.87, 7.75, 7.74, 7.72, 7.69, 7.68, 7.63, 7.62, 7.56, 7.43, 7.36, 7.35, 7.34, 7.31, 7.27, 7.26, 7.25, 7.24, 7.16, 7.14, 7.13, 7.11, 7.11, 7.09, 7.07, 7.07, 7.07, 7.01, 7.01, 6.98, 6.96, 6.94, 6.92, 6.90, 6.89, 6.88, 6.88, 6.87, 6.82, 6.79, 6.69, 6.66, 6.65, 6.63, 6.62, 6.61, 6.57, 6.56, 6.55, 6.54, 6.32, 6.22, 6.20, 6.19, 6.18, 6.17, 6.16, 6.15, 6.11, 6.03, 5.91, 5.76, 5.69, 5.69, 5.66, 5.56, 5.34, 4.64, 4.51, 4.49, 4.47, 4.47, 4.42, 4.41, 4.40, 4.38, 4.37, 4.35, 4.34, 4.33, 4.32, 4.32, 4.31, 4.30, 4.29, 4.25, 4.24, 4.23, 4.23, 4.22, 4.16, 4.16, 4.13, 4.10, 4.07, 4.06, 4.05, 3.98, 3.98, 3.97, 3.95, 3.94, 3.93, 3.92, 3.91, 3.89, 3.88, 3.82, 3.70, 3.66, 3.61, 3.59, 3.56, 3.50, 3.49, 3.48, 3.47, 3.37, 3.34, 3.33, 3.32, 3.32, 3.31, 3.16, 3.14, 3.12, 3.11, 2.86, 2.84, 2.79, 2.79, 2.78, 2.61, 2.60, 2.59, 2.57, 2.50, 2.49, 2.48, 2.47, 2.46, 2.45, 2.44, 2.43, 2.42, 2.41, 2.40, 2.39, 2.35, 2.34, 2.33, 2.32, 2.32, 2.31, 2.30, 2.27, 2.26, 2.25, 2.24, 2.23, 2.22, 2.21, 2.20, 2.17, 2.17, 2.16, 2.15, 2.09, 2.08, 2.07, 2.07, 2.06, 2.06, 2.05, 2.05, 2.05, 2.04, 2.03, 2.01, 1.98, 1.97, 1.95, 1.95, 1.83, 1.69, 1.58, 1.56, 1.54, 1.53, 1.51, 1.51, 1.44, 1.43, 1.43, 1.42, 1.41, 1.38, 1.37, 1.36, 1.35, 1.33, 1.32, 1.31, 1.29, 1.28, 1.27, 1.26, 1.26, 1.26, 1.25, 1.25, 1.24, 1.23, 1.22, 1.22, 1.21, 1.20, 1.20, 1.19, 1.19, 1.18, 1.17, 1.15, 1.14, 1.13, 1.12, 1.12, 1.11, 1.11, 1.10, 1.10, 1.09, 1.04, 1.03, 1.03, 1.00, 0.93, 0.89, 0.88, 0.87, 0.86, 0.85, 0.85, 0.83, 0.83, 0.73, 0.69, 0.68, 0.68, 0.67, 0.63, 0.62, 0.56, 0.55, 0.19, 0.14, 0.11, 0.11, 0.08, 0.07, 0.04, 0.03. ^{19}F NMR (376 MHz, Acetone- d_6 , 258 K, δ ppm): -130.3, -131.8.

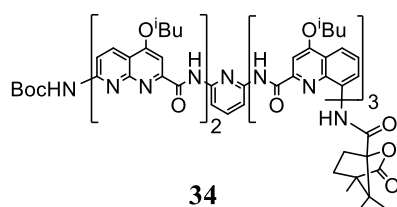
HRMS (ES $^+$): m/z calcd for $\text{C}_{300}\text{H}_{299}\text{F}_2\text{N}_{56}\text{O}_{52}$ [$\text{M}+3\text{H}$] $^{3+}$: 1852.7509 found 1852.7653.



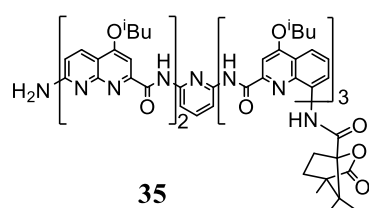
10

Oligomer 10. Dicarboxylate **25** (10.1 mg, 7.8 μmol) was suspended in anhydrous CHCl_3 (1 mL), then oxalyl chloride (7 μL , 78 μmol) was added and the reaction was allowed to stir at room temperature for 2 hours. The solvent and excess oxalyl chloride were removed under reduced pressure and the residue was dried under high vacuum for 3 hours to yield the corresponding acid chloride **25a** as a yellow solid. The solution of amine **31** (37 mg, 17 μmol) and distilled DIPEA (7 μL , 40 μmol) in anhydrous CHCl_3 (1 mL) was added dropwise via a syringe to solution of the freshly prepared **25a** dissolved in anhydrous CHCl_3 (0.5 mL). The reaction was allowed to proceed at room temperature for 16 hours. After evaporation of the solvents, the crude product was purified by recycling GPC. Oligomer **10** was obtained as a yellow product (26 mg, 58% yield). ^1H NMR (700 MHz, CDCl_3 , 238 K, δ ppm): 11.76 (s, 2H),

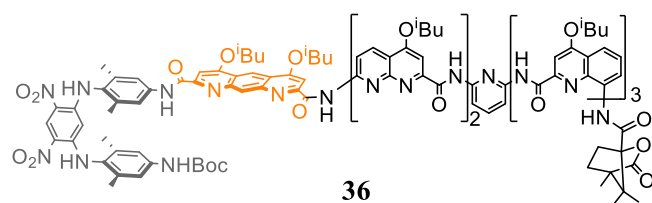
11.02 (s, 2H), 10.55 (s, 2H), 10.50 (s, 2H), 10.37 (s, 2H), 10.21 (s, 2H), 9.88 (s, 2H), 9.46 – 9.29 (m, 5H), 9.14 (d, $J = 7.9$ Hz, 3H), 9.08 – 8.91 (m, 7H), 8.85 (d, $J = 7.9$ Hz, 3H), 8.57 (d, $J = 14.5$ Hz, 5H), 8.46 (d, $J = 7.7$ Hz, 3H), 8.44 – 8.36 (m, 5H), 8.31 (dd, $J = 10.0, 6.6$ Hz, 5H), 8.06 (s, 2H), 7.92 (s, 3H), 7.75 (s, 2H), 7.45 (d, $J = 4.9$ Hz, 5H), 7.31 (d, $J = 6.5$ Hz, 4H), 7.18 (d, $J = 12.7$ Hz, 6H), 7.12 (d, $J = 13.7$ Hz, 3H), 7.11 – 7.00 (m, 8H), 6.92 (d, $J = 6.9$ Hz, 5H), 6.87 (s, 3H), 6.78 (s, 3H), 6.67 (d, $J = 13.7$ Hz, 5H), 6.49 (s, 2H), 6.39 (s, 3H), 6.18 (t, $J = 6.8$ Hz, 3H), 6.11 (t, $J = 6.8$ Hz, 3H), 5.92 (s, 2H), 5.66 (s, 3H), 4.99 (s, 6H), 4.40 (s, 3H), 4.35 (s, 2H), 4.22 (s, 6H), 4.16 (s, 3H), 4.07 (s, 6H), 3.97 (d, $J = 17.8$ Hz, 4H), 3.92 – 3.71 (m, 8H), 3.57 (s, 6H), 2.78 (d, $J = 10.2$ Hz, 3H), 2.64 (s, 3H), 2.48 (s, 3H), 2.43 – 2.34 (m, 4H), 2.34 – 2.16 (m, 8H), 2.07 (s, 8H), 1.94 (s, 6H), 1.43 (dd, $J = 24.7, 6.9$ Hz, 9H), 1.36 – 1.00 (m, 66H), 0.90 – 0.67 (m, 9H), 0.63 – 0.48 (m, 6H), 0.41 (d, $J = 7.2$ Hz, 6H), -0.05 (d, $J = 6.2$ Hz, 6H), -0.12 (d, $J = 6.4$ Hz, 6H). HRMS (ES⁺): m/z calcd for C₃₀₂H₃₀₅N₅₆O₅₄ [M+3H]³⁺: 1860.7642 found 1860.7791.



Heptamer 34. Amino oligomer **33**^[7] (57 mg, 0.04 mmol) was dissolved in dry chloroform (2 mL) containing DIPEA (0.04 mL, 0.226 mmol), then commercially available 1S-(-)-camphanic chloride (43 mg, 0.2 mmol) was added as a solid to the solution. The reaction mixture was stirred at room temperature overnight and the solvents were removed under reduced pressure. The residue was purified by recycling GPC to obtain **34** as a pale-yellow solid (60 mg, 94%). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 12.31 (s, 1H), 12.18 (s, 1H), 10.98 (s, 1H), 10.13 (s, 1H), 9.90 (s, 1H), 9.83 (s, 1H), 9.78 (s, 1H), 9.45 (dd, $J = 7.7, 1.3$ Hz, 1H), 8.82 – 8.68 (m, 2H), 8.65 (s, 1H), 8.61 (dd, $J = 7.7, 1.3$ Hz, 1H), 8.58 (s, 2H), 8.16 (dd, $J = 7.6, 1.1$ Hz, 1H), 8.04 (dd, $J = 8.4, 1.3$ Hz, 1H), 7.98 (dd, $J = 8.4, 1.3$ Hz, 1H), 7.88 (dd, $J = 7.7, 1.3$ Hz, 1H), 7.82 (t, $J = 7.8$ Hz, 1H), 7.77 (d, $J = 3.0$ Hz, 2H), 7.75 – 7.65 (m, 2H), 7.29 (s, 1H), 7.23 (d, $J = 2.9$ Hz, 1H), 6.95 (s, 1H), 6.89 (dd, $J = 8.4, 1.3$ Hz, 1H), 6.58 (t, $J = 8.0$ Hz, 1H), 4.68 (dd, $J = 9.0, 5.8$ Hz, 1H), 4.33 – 4.01 (m, 7H), 3.96 (d, $J = 6.4$ Hz, 2H), 2.36 (m, 6H), 2.10 – 1.94 (m, 1H), 1.86 (q, $J = 6.8$ Hz, 2H), 1.77 (s, 10H), 1.35 – 1.12 (m, 22H), 0.79 (t, $J = 7.6$ Hz, 10H), 0.52 (d, $J = 6.7$ Hz, 3H), 0.44 (s, 3H). ¹³C NMR (75 MHz, CDCl₃, 298 K, δ ppm): 176.3, 164.9, 164.6, 164.2, 164.1, 163.9, 163.8, 163.5, 163.2, 162.0, 161.8, 160.5, 156.7, 155.1, 154.5, 153.9, 153.6, 153.4, 152.3, 151.5, 151.1, 149.6, 148.7, 147.9, 140.0, 138.9, 138.5, 137.7, 134.9, 134.4, 134.3, 133.4, 132.7, 127.5, 126.5, 126.0, 122.9, 122.3, 122.2, 117.7, 117.0, 116.8, 116.7, 116.5, 116.2, 115.0, 114.9, 114.8, 114.2, 109.6, 108.7, 102.2, 99.5, 98.5, 98.4, 98.3, 91.8, 81.6, 77.6, 77.4, 77.2, 76.7, 75.9, 75.8, 75.5, 75.4, 75.3, 54.9, 54.2, 29.7, 28.9, 28.8, 28.4, 28.3, 28.3, 19.6, 19.5, 19.4, 19.4, 18.8, 16.4, 16.4, 9.8. HRMS (ES⁺): m/z calcd for C₈₈H₉₆N₁₅O₁₅ [M+H]⁺ 1602.7205 Found 1602.7341.

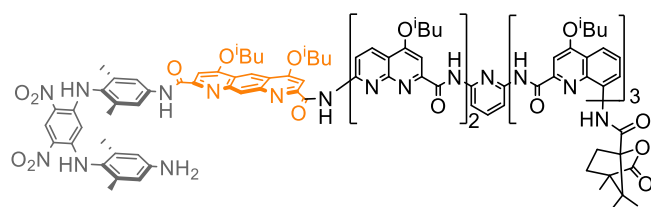


Heptamer amine 35. In a round-bottom flask, heptamer **34** (60 mg, 0.037 mmol) was dissolved in 2 mL chloroform and subsequently, 2 mL TFA was added dropwise. The mixture was allowed to stir for 3 hours at room temperature. A saturated aqueous solution of NaHCO_3 was added to quench the excess acid. Then, the solution was extracted with CH_2Cl_2 , washed with water, dried over Na_2SO_4 , and evaporated. Heptamer amine **35** was obtained without any further purification as a yellow solid (50 mg, 90% yield). ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 12.34 (s, 1H), 12.18 (s, 1H), 11.35 (s, 1H), 10.03 (s, 1H), 9.81 (s, 1H), 9.75 (s, 1H), 9.13 (d, $J = 7.6$ Hz, 1H), 8.74 (s, 2H), 8.59 (d, $J = 7.5$ Hz, 1H), 8.43 (s, 1H), 8.27 (d, $J = 9.1$ Hz, 1H), 8.20 (d, $J = 7.0$ Hz, 1H), 8.05 – 7.93 (m, 2H), 7.88 (t, $J = 7.3$ Hz, 1H), 7.80 (dd, $J = 9.6, 7.8$ Hz, 3H), 7.66 (t, $J = 8.0$ Hz, 1H), 7.54 (s, 1H), 7.28 (d, $J = 3.1$ Hz, 2H), 7.24 (d, $J = 8.0$ Hz, 2H), 6.99 (d, $J = 8.4$ Hz, 1H), 6.91 (s, 1H), 6.85 (d, $J = 9.0$ Hz, 1H), 6.67 (t, $J = 8.0$ Hz, 1H), 6.21 (s, 2H), 4.53 (s, 1H), 4.34 – 4.13 (m, 4H), 4.13 – 3.81 (m, 4H), 2.36 (dddd, $J = 35.6, 29.1, 13.3, 6.7$ Hz, 4H), 1.77 (dd, $J = 14.0, 5.5$ Hz, 2H), 1.39 – 1.01 (m, 24H), 0.76 (dd, $J = 13.5, 8.5$ Hz, 9H), 0.57 – 0.37 (m, 6H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 176.4, 164.8, 164.6, 164.0, 163.9, 163.7, 163.5, 163.1, 162.4, 161.8, 161.5, 160.6, 156.4, 154.7, 154.1, 153.6, 151.5, 151.1, 150.8, 149.7, 148.8, 147.8, 140.0, 138.8, 138.3, 137.7, 134.8, 134.3, 132.7, 132.4, 127.5, 126.5, 126.1, 122.7, 122.2, 117.3, 116.9, 116.8, 116.7, 116.5, 116.2, 114.8, 114.5, 113.5, 111.0, 110.0, 108.9, 101.5, 99.5, 98.6, 98.2, 96.6, 91.9, 91.6, 77.4, 75.9, 75.5, 75.4, 75.3, 75.2, 54.8, 54.2, 29.8, 29.3, 28.8, 28.4, 28.4, 28.2, 28.1, 19.6, 19.5, 19.5, 19.4, 19.3, 18.6, 16.4, 16.4, 9.8. HRMS (ESI): m/z calcd for $\text{C}_{83}\text{H}_{88}\text{N}_{15}\text{O}_{13}$ $[\text{M}+\text{H}]^+$ 1502.6681, Found 1502.6788.



Oligomer 36. Amino heptamer **35** (50 mg, 0.033 mmol), tetramer acid **28** (37 g, 0.04 mmol), and PyBOP (86 mg, 0.165 mmol) were placed in a 5 mL round bottom flask then dissolved in freshly distilled CHCl_3 (1 mL) and DIPEA (30 μL , 0.165 mmol) was added under nitrogen atmosphere. The reaction mixture was allowed to stir at room temperature for 36 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane. The solution was then washed successively with 5% NH_4Cl , water and brine. Finally, the organic layer was dried over Na_2SO_4 , filtered, and evaporated. The

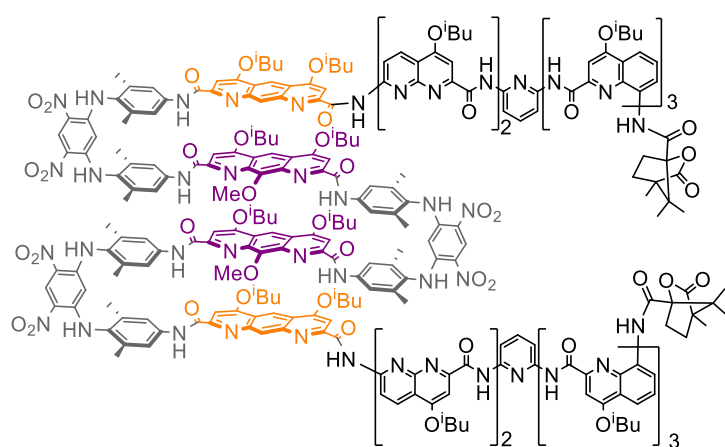
residue was purified by recycling GPC and product **36** was obtained as a yellow solid (60 mg, 75% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 11.94 (s, 2H), 11.85 (s, 1H), 11.76 (s, 1H), 9.79 (s, 1H), 9.75 (s, 1H), 9.61 (d, *J* = 5.2 Hz, 2H), 9.30 (s, 1H), 9.06 – 8.97 (m, 2H), 8.92 (s, 1H), 8.88 – 8.74 (m, 4H), 8.62 (s, 1H), 8.47 (dd, *J* = 7.6, 1.3 Hz, 1H), 8.22 (d, *J* = 7.5 Hz, 1H), 7.92 (d, *J* = 7.5 Hz, 1H), 7.83 (s, 2H), 7.81 – 7.72 (m, 3H), 7.68 (s, 1H), 7.56 (dd, *J* = 7.7, 1.3 Hz, 1H), 7.31 (d, *J* = 10.0 Hz, 2H), 7.24 – 7.15 (m, 2H), 7.08 – 6.97 (m, 2H), 6.86 (dd, *J* = 8.4, 1.3 Hz, 1H), 6.79 (s, 1H), 6.73 – 6.61 (m, 2H), 6.47 (t, *J* = 8.0 Hz, 1H), 6.42 (s, 1H), 6.30 (s, 1H), 6.24 (s, 1H), 4.75 (s, 1H), 4.41 – 4.12 (m, 8H), 4.01 (m, 2H), 3.88 (t, *J* = 8.0 Hz, 1H), 3.78 (t, *J* = 7.5 Hz, 1H), 3.67 (t, *J* = 8.0 Hz, 1H), 3.62 – 3.50 (m, 1H), 3.38 (t, *J* = 8.9 Hz, 1H), 2.57 – 2.23 (m, 6H), 1.96 (d, *J* = 14.4 Hz, 7H), 1.71 (s, 4H), 1.46 (s, 4H), 1.39 – 1.10 (m, 36H), 0.99 (s, 10H), 0.77 (d, *J* = 6.5 Hz, 3H), 0.60 (s, 6H), 0.52 (d, *J* = 6.7 Hz, 3H), 0.21 (s, 3H). ¹³C NMR (75 MHz, CDCl₃, 298 K, δ ppm): 176.1, 164.7, 164.5, 164.1, 164.0, 163.9, 163.8, 163.4, 163.4, 163.2, 162.1, 161.8, 161.7, 160.9, 159.6, 154.8, 154.7, 154.6, 154.1, 153.8, 152.7, 152.1, 151.7, 151.3, 151.2, 149.1, 148.3, 148.1, 147.0, 145.6, 145.4, 138.8, 138.8, 137.7, 137.4, 137.1, 136.6, 136.4, 136.2, 135.9, 134.6, 134.5, 134.4, 134.2, 132.4, 129.7, 129.4, 129.2, 129.0, 126.1, 125.8, 124.9, 124.8, 122.7, 121.9, 121.4, 121.0, 120.5, 119.4, 119.1, 118.8, 116.8, 116.6, 116.3, 116.2, 115.8, 115.2, 115.1, 114.9, 114.4, 108.9, 108.7, 101.1, 99.3, 98.9, 98.1, 97.3, 96.1, 96.0, 93.9, 91.5, 79.9, 77.4, 76.1, 76.0, 75.5, 75.3, 74.9, 54.6, 54.0, 46.4, 46.4, 45.9, 29.8, 29.5, 29.0, 28.7, 28.5, 28.5, 28.3, 28.3, 28.0, 27.9, 27.0, 26.6, 26.5, 19.6, 19.5, 19.5, 19.4, 19.2, 18.6, 18.3, 18.1, 17.8, 17.1, 16.2, 16.2, 9.7. HRMS (ESI): *m/z* calcd for C₁₃₂H₁₄₁N₂₃O₂₃ [M+2H]²⁺ 1208.5297, Found 1208.5390.



37

Oligomer 37. In a round-bottom flask, oligomer **36** (60 mg, 0.025 mmol) was dissolved in HCl in dioxane (4M, 2mL) and chloroform (0.5 mL). The mixture was allowed to stir for 4 hours at room temperature. The reaction was quenched with saturated solution of NaHCO₃, and the solution was extracted three times with CH₂Cl₂, washed with water, dried over Na₂SO₄, and evaporated. Compound **37** was obtained as yellow solid which was used without any further purification (57 mg, 98% yield). ¹H NMR (300 MHz, CDCl₃, 298 K, δ ppm): 11.96 (s, 1H), 11.93 (s, 1H), 11.82 (s, 2H), 9.85 (s, 1H), 9.73 (s, 1H), 9.70 (s, 1H), 9.60 (s, 1H), 9.23 (s, 1H), 8.99 – 8.88 (m, 2H), 8.88 – 8.71 (m, 5H), 8.61 (s, 1H), 8.48 (d, *J* = 7.5 Hz, 1H), 8.30 (d, *J* = 7.5 Hz, 1H), 7.96 – 7.85 (m, 2H), 7.84 – 7.77 (m, 3H), 7.75 (d, *J* = 5.3 Hz, 2H), 7.66 (s, 1H), 7.56 (d, *J* = 7.6 Hz, 1H), 7.38 (s, 1H), 7.36 – 7.28 (m, 3H), 7.21 (d, *J* = 8.2 Hz, 1H), 7.04 (t, *J* = 8.0 Hz, 1H), 6.99 (s, 1H), 6.84 (d, *J* = 8.3 Hz, 1H), 6.71 (t, *J* = 8.0 Hz, 1H), 6.47 (t, *J* = 8.0 Hz, 1H), 6.25 (s, 1H), 6.08 (s, 1H), 5.95 (s, 1H), 5.90 (s, 1H), 4.52 (s, 1H), 4.26 (m, 8H), 4.01 (m, 2H), 3.86 (t, *J* = 8.0 Hz, 1H), 3.73 (d, *J* = 6.8 Hz, 1H), 3.67 (d, *J* = 7.7 Hz, 1H), 3.51 (t, *J* = 7.4

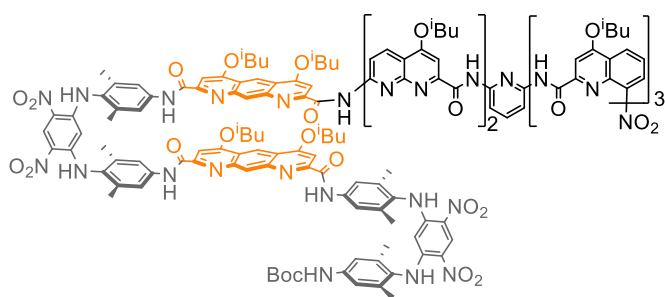
Hz, 1H), 3.37 (t, $J = 9.0$ Hz, 2H), 2.43 (m, 6H), 2.00 (s, 3H), 1.70 (s, 4H), 1.43 (s, 5H), 1.39 – 1.06 (m, 36H), 0.79 (d, $J = 6.6$ Hz, 4H), 0.58 (d, $J = 12.6$ Hz, 9H), 0.21 (s, 3H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 176.1, 164.8, 164.5, 164.1, 163.9, 163.8, 163.6, 163.3, 163.3, 162.0, 161.8, 160.9, 159.6, 154.8, 154.6, 154.0, 153.8, 152.6, 152.4, 151.9, 151.3, 151.3, 149.1, 148.8, 148.4, 148.0, 147.0, 146.2, 145.6, 145.4, 139.3, 138.8, 137.4, 137.1, 136.9, 136.8, 136.4, 136.2, 134.6, 134.4, 134.2, 132.4, 130.0, 129.3, 128.6, 126.2, 126.1, 125.8, 124.9, 124.8, 122.8, 121.9, 121.4, 120.9, 120.5, 118.6, 118.4, 116.8, 116.6, 116.4, 115.4, 115.2, 115.1, 115.0, 114.6, 114.6, 114.3, 109.1, 108.8, 101.1, 99.2, 98.8, 98.1, 97.2, 96.2, 96.1, 94.2, 91.5, 77.6, 77.4, 77.2, 76.7, 76.1, 76.0, 75.5, 75.4, 74.9, 54.6, 54.0, 29.8, 29.4, 28.9, 28.7, 28.5, 28.4, 28.4, 28.3, 27.9, 19.5, 19.5, 19.5, 19.4, 19.4, 19.4, 19.2, 18.7, 18.3, 17.7, 17.0, 16.2, 16.2, 9.7, 1.2. HRMS (ESI): m/z calcd for $\text{C}_{254}\text{H}_{264}\text{N}_{46}\text{O}_{42}$ $[\text{2M}+\text{2H}]^{2+}$ 2315.9996, Found 2316.0141.



11

Oligomer 11. Dicarboxylate **25** (6.43 mg, 5 μmol) was suspended in anhydrous CHCl_3 (1 mL), then oxalyl chloride (4.3 μL , 50 μmol) was added and the reaction was allowed to stir at room temperature for 2 hours. The solvent and excess oxalyl chloride were removed under reduced pressure and the residue was dried under high vacuum for 3 hours to yield the corresponding acid chloride **25a** as a yellow solid. The solution of amine **37** (37 mg, 17 μmol) and distilled DIPEA (7 μL , 40 μmol) in anhydrous CHCl_3 (1 mL) was added dropwise via a syringe to solution of the freshly prepared **25a** dissolved in anhydrous CHCl_3 (0.5 mL). The reaction was allowed to proceed at room temperature for 16 hours. After evaporation of the solvents, the crude product was purified by recycling GPC. Oligomer **11** was obtained as a yellow product (24 mg, 82% yield). ^1H NMR (700 MHz, CDCl_3 , 238 K, δ ppm): 11.68 (s, 2H), 11.38 (s, 2H), 10.83 (s, 2H), 10.26 (s, 2H), 10.19 (s, 2H), 10.05 (s, 2H), 9.64 (s, 2H), 9.33 (m, 4H), 9.13 (s, 2H), 9.06 (d, $J = 8.1$ Hz, 2H), 8.96 (s, 2H), 8.90 – 8.74 (m, 3H), 8.66 (s, 2H), 8.48 (m, 4H), 8.33 (d, $J = 6.7$ Hz, 2H), 8.18 (d, $J = 7.1$ Hz, 2H), 8.10 (s, 2H), 7.86 (s, 2H), 7.76 (d, $J = 7.5$ Hz, 3H), 7.47 (d, $J = 6.1$ Hz, 5H), 7.21 – 7.05 (m, 7H), 7.00 (s, 2H), 6.95 – 6.78 (m, 7H), 6.71 (s, 2H), 6.51 (s, 2H), 6.45 – 6.18 (m, 5H), 6.18 – 5.98 (m, 4H), 5.93 (s, 2H), 4.88 (s, 4H), 4.44 (s, 2H), 4.24 (d, $J = 6.1$ Hz, 4H), 4.18 (m, 3H), 4.11 – 3.84 (m, 13H), 3.80 (t, $J = 6.3$ Hz, 2H), 3.69 – 3.56 (m, 4H), 3.49 (s, 2H), 3.06 (s, 2H),

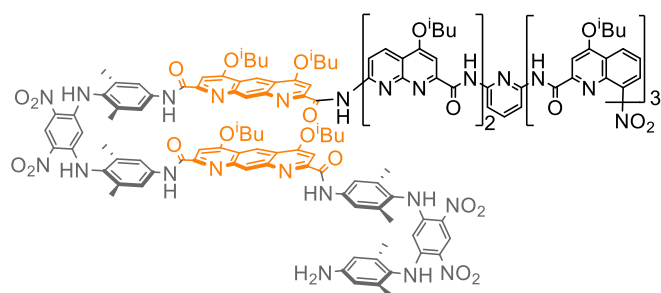
2.87 (s, 2H), 2.65 (m, 2H), 2.40 (m, 4H), 2.26 (m, 8H), 2.06 (s, 10H), 1.96 (s, 6H), 1.49 – 0.99 (m, 128H), 0.99 – 0.76 (m, 12H), 0.61 (s, 6H), 0.46 (d, $J = 7.1$ Hz, 6H), 0.39 (d, $J = 17.1$ Hz, 12H), 0.18 (s, 6H), 0.13 – 0.05 (m, 6H), 0.01 (s, 6H). ^{13}C NMR (75 MHz, CDCl_3 , 238 K, δ ppm): 175.6, 164.3, 163.9, 163.9, 163.6, 163.5, 163.3, 163.2, 163.1, 163.1, 162.4, 162.3, 162.3, 162.1, 161.7, 161.7, 161.6, 160.5, 160.1, 159.9, 159.5, 154.7, 154.2, 153.9, 153.7, 153.5, 151.5, 151.4, 151.2, 150.6, 150.6, 150.3, 149.2, 148.6, 148.6, 148.3, 148.1, 147.4, 144.4, 143.7, 138.6, 138.5, 138.3, 138.2, 137.3, 137.2, 137.0, 136.9, 136.9, 136.9, 136.8, 136.6, 136.4, 136.1, 134.9, 134.2, 134.1, 133.6, 132.3, 130.7, 130.1, 129.4, 129.2, 129.1, 127.4, 126.4, 125.3, 125.1, 125.0, 124.7, 122.5, 122.0, 120.7, 120.6, 120.5, 120.3, 119.7, 119.5, 119.5, 119.4, 119.2, 118.2, 117.4, 117.4, 116.9, 116.4, 116.2, 115.9, 115.0, 114.4, 114.2, 109.2, 108.8, 108.4, 100.6, 99.0, 99.0, 98.3, 97.8, 95.8, 94.9, 94.4, 94.3, 91.2, 77.6, 77.4, 77.2, 76.7, 76.0, 75.3, 75.0, 74.1, 65.7, 54.3, 53.8, 29.8, 28.7, 28.5, 28.4, 28.4, 28.3, 28.3, 28.2, 27.6, 27.4, 19.7, 19.6, 19.5, 19.4, 19.4, 19.3, 19.2, 19.1, 18.4, 18.2, 17.9, 17.9, 17.7, 17.0, 16.7, 16.0, 16.0, 9.44, 1.2. HRMS (ES⁺): m/z calcd for $\text{C}_{322}\text{H}_{333}\text{N}_{56}\text{O}_{56}$ $[\text{M}+3\text{H}]^{3+}$: 1960.8338 found 1960.8502.



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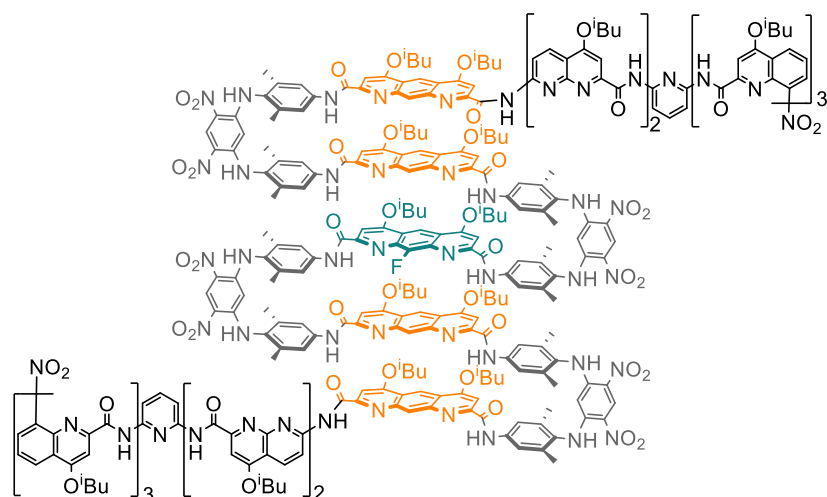
Oligomer 38. Amino oligomer **31** (61 mg, 0.028 mmol), tetramer acid **28** (31 mg, 0.034 mmol), and PyBOP (73 mg, 0.14 mmol) were placed in a 5 mL round bottom flask then dissolved in freshly distilled CHCl_3 (1 mL) and DIPEA (15 μL , 0.084 mmol) was added under nitrogen atmosphere. The reaction mixture was allowed to stir at room temperature for 16 hours. The solvent was removed under reduced pressure and the residue was dissolved in dichloromethane. The solution was then washed successively with 5% NH_4Cl , water and brine. Finally, the organic layer was dried over Na_2SO_4 , filtered and evaporated. The residue was purified by recycling GPC and product **38** was obtained as a yellow solid (65 mg, 76% yield). ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 11.79 (s, 1H), 11.33 (s, 1H), 11.22 (s, 1H), 10.61 (s, 1H), 10.10 (s, 1H), 9.99 (s, 1H), 9.48 (s, 1H), 9.39 (s, 1H), 9.27 (d, $J = 8.4$ Hz, 2H), 9.06 (s, 1H), 8.99 (s, 1H), 8.84 (t, $J = 10.8$ Hz, 4H), 8.73 (s, 1H), 8.55 (s, 1H), 8.38 (d, $J = 7.6$ Hz, 1H), 8.32 (d, $J = 8.1$ Hz, 1H), 8.18 (s, 1H), 8.07 (s, 1H), 7.90 (d, $J = 11.4$ Hz, 3H), 7.80 (s, 1H), 7.58 (d, $J = 12.9$ Hz, 2H), 7.36 (d, $J = 21.0$ Hz, 3H), 7.22 – 7.07 (m, 2H), 7.02 (d, $J = 8.2$ Hz, 2H), 6.91 (s, 3H), 6.58 (d, $J = 18.7$ Hz, 2H), 6.41 (s, 3H), 6.26 (s, 1H), 6.06 (s, 1H), 4.77 (s, 1H), 4.41 (s, 1H), 4.39 – 3.73 (m, 14H), 3.56 (s, 3H), 2.98 (s, 1H), 2.66 (m, 1H), 2.60 – 2.11 (m, 9H), 2.11 – 1.88 (m, 12H), 1.81 (s, 6H),

1.48 – 1.01 (m, 58H), 0.90 (s, 9H), 0.63 (d, $J = 6.4$ Hz, 6H), 0.42 (s, 3H). HRMS (ESI): m/z calcd for $C_{166}H_{171}N_{31}O_{30}$ $[M+2H]^{2+}$ 1539.6415, Found 1539.6097.



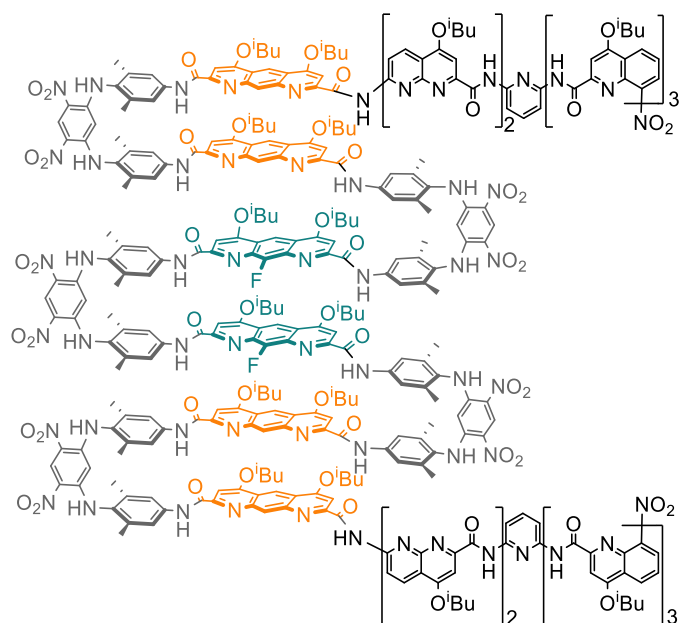
39

Oligomer 39. In a round-bottom flask, oligomer **38** (65 mg, 0.021 mmol) was dissolved in HCl in dioxane (4M, 2mL) and chloroform (0.8 mL). The mixture was allowed to stir for 5 hours at room temperature. The reaction was quenched with saturated solution of $NaHCO_3$, and the solution was extracted three times with CH_2Cl_2 , washed with water, dried over Na_2SO_4 , and evaporated. Compound **39** was obtained as yellow solid which was used without any further purification (60 mg, 96% yield). 1H NMR (300 MHz, $CDCl_3$, 298 K, δ ppm): 11.78 (s, 1H), 11.33 (s, 2H), 10.63 (s, 1H), 10.08 (s, 1H), 9.96 (s, 2H), 9.48 (s, 1H), 9.40 (s, 1H), 9.33 (s, 1H), 9.28 (s, 1H), 9.09 (s, 1H), 8.97 (s, 1H), 8.93 – 8.66 (m, 7H), 8.52 (s, 1H), 8.38 (d, $J = 7.6$ Hz, 1H), 8.31 (dd, $J = 7.8, 1.9$ Hz, 1H), 8.18 (s, 1H), 8.00 (d, $J = 19.2$ Hz, 3H), 7.88 (s, 1H), 7.82 (s, 1H), 7.48 (d, $J = 38.0$ Hz, 5H), 7.28 (d, $J = 3.2$ Hz, 1H), 7.23 (s, 1H), 7.21 – 7.09 (m, 2H), 7.04 (s, 1H), 7.00 (d, $J = 3.6$ Hz, 2H), 6.91 (s, 2H), 6.68 – 6.16 (m, 6H), 5.98 (d, $J = 12.7$ Hz, 3H), 4.64 (s, 1H), 4.41 (d, $J = 7.7$ Hz, 2H), 4.38 – 3.72 (m, 12H), 3.69 – 3.44 (m, 3H), 3.27 (s, 2H), 2.60 – 2.14 (m, 9H), 2.00 (d, $J = 23.9$ Hz, 9H), 1.80 (d, $J = 10.0$ Hz, 6H), 1.70 (s, 3H), 1.58 (d, $J = 19.0$ Hz, 9H), 1.49 – 1.02 (m, 43H), 0.87 (d, $J = 7.1$ Hz, 2H), 0.64 (d, $J = 6.4$ Hz, 3H), 0.43 (s, 3H). ^{13}C NMR (75 MHz, $CDCl_3$, 298 K, δ ppm): 164.3, 164.1, 163.5, 163.3, 162.8, 162.3, 162.1, 161.6, 161.3, 160.9, 160.7, 160.0, 154.8, 154.4, 154.0, 153.9, 152.3, 151.9, 150.9, 149.2, 148.9, 148.3, 148.2, 148.1, 147.0, 146.0, 144.7, 144.5, 138.9, 138.8, 137.8, 137.4, 137.2, 136.6, 136.4, 134.7, 134.5, 134.4, 130.9, 129.5, 129.5, 129.1, 128.2, 126.3, 125.8, 124.9, 124.9, 124.0, 123.4, 122.6, 120.8, 120.5, 120.4, 120.3, 119.1, 118.9, 117.1, 115.5, 115.2, 115.1, 115.0, 114.7, 114.4, 114.2, 109.1, 99.5, 98.9, 97.8, 95.9, 95.7, 94.8, 93.9, 90.5, 77.4, 76.0, 75.5, 75.4, 75.2, 75.0, 71.4, 70.6, 68.9, 67.0, 59.6, 42.8, 29.8, 28.6, 28.6, 28.4, 28.4, 28.2, 28.2, 27.6, 19.5, 19.4, 19.4, 19.2, 18.4, 17.8, 17.7, 14.2. HRMS (ESI): m/z calcd for $C_{161}H_{163}N_{31}O_{28}$ $[M+2H]^{2+}$ 1489.6153, Found 1489.5822.



12

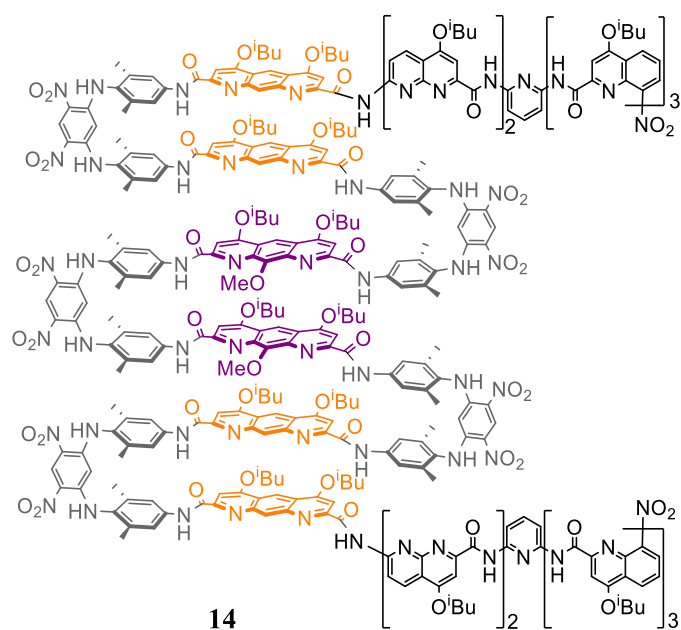
Oligomer 12. Dicarboxylic acid **40** (3 mg, 7 μmol) was suspended in anhydrous CHCl_3 (1 mL), then oxalyl chloride (6 μL , 68 μmol) was added and the reaction was allowed to stir at room temperature for 2 hours. The solvent and excess oxalyl chloride were removed under reduced pressure and the residue was dried under high vacuum for 3 hours to yield the corresponding acid chloride **40a** as a pale yellow solid. The solution of amine **39** (45 mg, 15 μmol) and distilled DIPEA (5 μL , 27 μmol) in anhydrous CHCl_3 (1 mL) was added dropwise via a syringe to solution of the freshly prepared **40a** dissolved in anhydrous CHCl_3 (0.5 mL). The reaction was allowed to proceed at room temperature for 16 hours. After evaporation of the solvents, the crude product was purified by recycling GPC. Oligomer **12** was obtained as a yellow product (20 mg, 46% yield). ^1H NMR (700 MHz, Acetone- d_6 , 298 K, δ ppm): 11.45 (s, 2H), 11.14 (s, 2H), 10.87 (s, 1H), 10.11 (s, 1H), 9.69 (s, 1H), 9.39 (d, $J = 6.5$ Hz, 2H), 9.34 (s, 2H), 9.29 (s, 2H), 9.14 (d, $J = 4.0$ Hz, 2H), 9.02 (s, 1H), 8.91 (s, 1H), 8.86 (d, $J = 8.1$ Hz, 1H), 8.73 (d, $J = 8.1$ Hz, 1H), 8.59 (d, $J = 7.9$ Hz, 1H), 8.51 (d, $J = 10.7$ Hz, 3H), 8.43 (d, $J = 7.9$ Hz, 1H), 8.27 (s, 1H), 8.26 – 8.21 (m, 2H), 8.19 (d, $J = 6.6$ Hz, 1H), 7.83 (d, $J = 8.0$ Hz, 2H), 7.75 (d, $J = 6.8$ Hz, 1H), 7.67 (s, 2H), 7.40 (d, $J = 5.5$ Hz, 4H), 7.35 (s, 1H), 7.26 (t, $J = 6.8$ Hz, 1H), 7.23 (s, 2H), 7.14 (t, $J = 6.9$ Hz, 1H), 7.06 (s, 2H), 7.01 (d, $J = 6.5$ Hz, 1H), 6.93 (d, $J = 6.8$ Hz, 1H), 6.83 – 6.77 (m, 3H), 6.72 (s, 2H), 6.68 (s, 1H), 6.51 (h, $J = 7.3$ Hz, 5H), 6.42 (s, 1H), 6.16 (m, 3H), 5.95 (s, 2H), 5.69 (s, 3H), 5.62 (s, 1H), 5.34 (s, 2H), 4.47 (s, 1H), 4.42 (m, 5H), 4.36 (t, $J = 5.9$ Hz, 4H), 4.33 – 4.18 (m, 15H), 4.12 (t, $J = 5.9$ Hz, 2H), 4.05 (m, 3H), 3.95 (m, 5H), 3.83 – 3.71 (m, 2H), 3.64 (d, $J = 5.7$ Hz, 2H), 3.53 – 3.45 (m, 2H), 3.41 (m, $J = 20.6, 5.1$ Hz, 4H), 3.21 (s, 1H), 2.54 (m, 5H), 2.36 (m, 7H), 2.27 (m, 2H), 2.24 – 2.13 (m, 7H), 2.09 (d, $J = 7.6$ Hz, 5H), 1.78 (s, 4H), 1.73 (s, 4H), 1.62 (s, 7H), 1.52 (s, 5H), 1.49 – 1.04 (m, 161H), 0.87 (m, 8H), 0.64 (d, $J = 7.3$ Hz, 6H), 0.58 (d, $J = 6.4$ Hz, 5H). ^{19}F NMR (376 MHz, Acetone- d_6 , 298 K, δ ppm): -131.8. HRMS (ESI): m/z calcd for $\text{C}_{344}\text{H}_{344}\text{FN}_{64}\text{O}_{60}$ $[\text{M}+3\text{H}]^{3+}$ 2117.8645, Found 2117.8780.



13

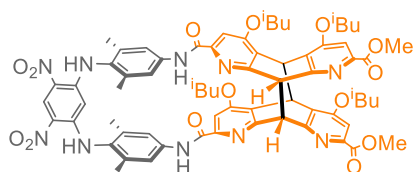
Oligomer 13. Dicarboxylic acid **24** (6.5 mg, 5.2 μmol) was suspended in anhydrous CHCl_3 (1 mL), then oxalyl chloride (5 μL , 52 μmol) was added and the reaction was allowed to stir at room temperature for 2 hours. The solvent and excess oxalyl chloride were removed under reduced pressure and the residue was dried under high vacuum for 3 hours to yield the corresponding acid chloride **24a** as a pale yellow solid. The solution of amine **39** (34 mg, 11.4 μmol) and distilled DIPEA (5 μL , 26 μmol) in anhydrous CHCl_3 (1 mL) was added dropwise via a syringe to solution of the freshly prepared **24a** dissolved in anhydrous CHCl_3 (0.5 mL). The reaction was allowed to proceed at room temperature for 16 hours. After evaporation of the solvents, the crude product was purified by recycling GPC. Oligomer **13** was obtained as a yellow solid (14 mg, 38% yield). ^1H NMR (700 MHz, Acetone- d_6 , 248 K, δ ppm; only peaks are given due to the presence of two conformers): 11.60, 11.49, 11.47, 11.28, 11.18, 11.14, 11.07, 11.02, 10.93, 10.64, 10.32, 10.21, 10.09, 10.08, 9.84, 9.83, 9.72, 9.62, 9.61, 9.51, 9.48, 9.43, 9.33, 9.29, 9.27, 9.22, 9.20, 9.19, 9.18, 9.16, 9.15, 9.13, 9.07, 9.07, 9.03, 9.02, 8.96, 8.95, 8.94, 8.94, 8.93, 8.92, 8.76, 8.75, 8.74, 8.72, 8.71, 8.67, 8.62, 8.61, 8.60, 8.56, 8.54, 8.53, 8.50, 8.45, 8.43, 8.41, 8.39, 8.31, 8.30, 8.29, 8.25, 8.24, 8.24, 8.21, 8.20, 8.19, 8.18, 8.08, 8.03, 7.93, 7.92, 7.90, 7.89, 7.87, 7.86, 7.85, 7.80, 7.74, 7.74, 7.72, 7.71, 7.69, 7.68, 7.66, 7.63, 7.62, 7.53, 7.47, 7.42, 7.37, 7.36, 7.35, 7.34, 7.33, 7.32, 7.31, 7.28, 7.27, 7.26, 7.23, 7.23, 7.22, 7.20, 7.18, 7.15, 7.11, 7.10, 7.10, 7.08, 7.07, 7.06, 7.03, 7.02, 7.01, 7.01, 6.96, 6.95, 6.95, 6.94, 6.88, 6.87, 6.86, 6.83, 6.83, 6.82, 6.78, 6.70, 6.69, 6.66, 6.66, 6.64, 6.63, 6.62, 6.58, 6.56, 6.54, 6.53, 6.52, 6.48, 6.47, 6.46, 6.37, 6.31, 6.20, 6.19, 6.18, 6.18, 6.15, 6.14, 6.13, 6.12, 6.03, 5.97, 5.88, 5.75, 5.73, 5.71, 5.68, 5.64, 5.62, 5.58, 5.55, 5.53, 5.32, 5.23, 4.58, 4.50, 4.48, 4.47, 4.45, 4.42, 4.40, 4.37, 4.35, 4.31, 4.25, 4.20, 4.18, 4.10, 4.04, 4.03, 4.03, 4.02, 4.02, 4.01, 4.01, 3.98, 3.97, 3.96, 3.95, 3.93, 3.90, 3.82, 3.80, 3.68, 3.63, 3.60, 3.56, 3.47, 3.42, 3.36, 3.32, 3.32, 3.31, 3.30, 3.25, 3.24, 3.23, 3.23, 3.21, 3.20, 3.19, 3.17, 3.14, 3.13, 3.13, 3.12, 3.11, 3.11, 3.10,

3.09, 3.08, 3.06, 3.02, 3.01, 2.83, 2.76, 2.75, 2.68, 2.61, 2.61, 2.60, 2.59, 2.53, 2.48, 2.47, 2.46, 2.45, 2.40, 2.39, 2.38, 2.38, 2.37, 2.36, 2.35, 2.34, 2.33, 2.29, 2.28, 2.27, 2.26, 2.25, 2.23, 2.22, 2.20, 2.17, 2.15, 2.14, 2.14, 2.14, 2.13, 2.13, 2.13, 2.08, 2.08, 2.07, 2.07, 2.06, 2.06, 2.05, 2.05, 2.05, 2.04, 2.03, 1.99, 1.99, 1.98, 1.97, 1.96, 1.96, 1.96, 1.96, 1.95, 1.93, 1.90, 1.89, 1.86, 1.80, 1.79, 1.78, 1.74, 1.70, 1.69, 1.66, 1.64, 1.63, 1.63, 1.61, 1.59, 1.58, 1.53, 1.51, 1.50, 1.49, 1.48, 1.47, 1.47, 1.46, 1.44, 1.43, 1.42, 1.40, 1.39, 1.38, 1.36, 1.35, 1.34, 1.33, 1.32, 1.31, 1.31, 1.30, 1.30, 1.29, 1.28, 1.25, 1.24, 1.23, 1.23, 1.22, 1.22, 1.21, 1.20, 1.20, 1.19, 1.19, 1.18, 1.18, 1.17, 1.17, 1.16, 1.15, 1.14, 1.12, 1.12, 1.12, 1.11, 1.11, 1.10, 1.10, 1.09, 1.08, 1.08, 1.06, 1.05, 1.04, 1.03, 1.03, 1.02, 1.02, 1.01, 1.00, 1.00, 0.99, 0.99, 0.98, 0.97, 0.95, 0.94, 0.93, 0.92, 0.88, 0.88, 0.87, 0.87, 0.86, 0.85, 0.84, 0.84, 0.83, 0.83, 0.82, 0.82, 0.81, 0.81, 0.80, 0.80, 0.79, 0.78, 0.70, 0.69, 0.68, 0.67, 0.66, 0.65, 0.63, 0.62, 0.58, 0.57, 0.16, 0.11, 0.09, 0.08, 0.07, 0.06, 0.05, 0.04, -0.01, -0.22, -0.23. ^{19}F NMR (376 MHz, Acetone- d_6 , 248 K, δ ppm): -130.9, 131.2, -132.8. HRMS (ESI): m/z calcd for $\text{C}_{388}\text{H}_{384}\text{F}_2\text{N}_{72}\text{O}_{68}$ $[\text{M}+3\text{H}]^{3+}$ 2394.63746, Found 2394.65375.



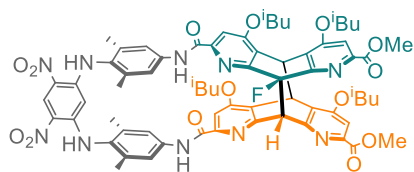
Oligomer 14. Dicarboxylic acid **25** (12.8 mg, 10 μmol) was suspended in anhydrous CHCl_3 (1 mL), then oxalyl chloride (20 μL , 200 μmol) was added and the reaction was allowed to stir at room temperature for 2 hours. The solvent and excess oxalyl chloride were removed under reduced pressure and the residue was dried under high vacuum for 3 hours to yield the corresponding acid chloride **25a** as a pale yellow solid. The solution of amine **39** (65.5 mg, 22 μmol) and distilled DIPEA (10 μL , 60 μmol) in anhydrous CHCl_3 (1 mL) was added dropwise via a syringe to solution of the freshly prepared **25a** dissolved in anhydrous CHCl_3 (0.5 mL). The reaction was allowed to proceed at room temperature for 16 hours. After evaporation of the solvents, the crude product was purified by recycling GPC. Oligomer **14** was obtained as a yellow product (38 mg, 54% yield). ^1H NMR (700 MHz, Acetone- d_6 ,

253 K, δ ppm; only peaks are given due to the presence of two conformers): 11.60, 11.50, 11.48, 11.28, 11.20, 11.14, 11.03, 10.92, 10.62, 10.33, 10.23, 10.19, 10.09, 9.99, 9.84, 9.64, 9.62, 9.59, 9.51, 9.47, 9.45, 9.38, 9.36, 9.32, 9.29, 9.27, 9.20, 9.18, 9.17, 9.12, 9.09, 9.05, 9.04, 9.01, 8.95, 8.92, 8.91, 8.88, 8.84, 8.72, 8.65, 8.54, 8.50, 8.45, 8.35, 8.30, 8.26, 8.25, 8.24, 8.21, 8.20, 8.19, 8.19, 8.17, 8.15, 8.04, 7.93, 7.87, 7.74, 7.71, 7.67, 7.64, 7.61, 7.56, 7.52, 7.49, 7.43, 7.41, 7.35, 7.31, 7.24, 7.22, 7.20, 7.15, 7.11, 7.07, 7.06, 7.03, 7.02, 6.94, 6.89, 6.84, 6.80, 6.77, 6.71, 6.66, 6.64, 6.61, 6.57, 6.45, 6.35, 6.24, 6.13, 6.10, 6.03, 6.01, 5.92, 5.89, 5.86, 5.71, 5.69, 5.65, 5.62, 5.57, 5.55, 5.41, 5.33, 4.59, 4.47, 4.40, 4.37, 4.35, 4.29, 4.26, 4.21, 4.16, 4.13, 4.09, 4.04, 4.03, 4.02, 4.01, 4.00, 3.96, 3.93, 3.90, 3.83, 3.70, 3.62, 3.55, 3.52, 3.44, 3.37, 3.18, 3.15, 2.93, 2.84, 2.78, 2.66, 2.60, 2.59, 2.54, 2.45, 2.40, 2.30, 2.23, 2.08, 2.07, 2.06, 2.05, 2.05, 2.05, 2.04, 2.02, 1.98, 1.96, 1.90, 1.89, 1.86, 1.82, 1.80, 1.71, 1.65, 1.64, 1.62, 1.60, 1.51, 1.50, 1.47, 1.47, 1.45, 1.43, 1.42, 1.41, 1.38, 1.36, 1.35, 1.34, 1.34, 1.33, 1.32, 1.30, 1.29, 1.27, 1.26, 1.25, 1.24, 1.21, 1.21, 1.20, 1.17, 1.17, 1.16, 1.15, 1.14, 1.13, 1.10, 1.09, 1.05, 1.05, 1.04, 1.01, 1.00, 1.00, 0.99, 0.96, 0.95, 0.94, 0.86, 0.85, 0.84, 0.83, 0.83, 0.82, 0.73, 0.71, 0.70, 0.69, 0.68, 0.66, 0.65, 0.61, 0.60, 0.57, 0.56, 0.14, 0.11, 0.07, 0.04, -0.29. HRMS (ESI): m/z calcd for $C_{390}H_{390}N_{72}O_{70}$ $[M+3H]^{3+}$ 2402.65079, Found 2402.65405.



1a

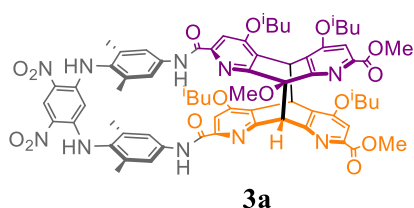
Photoproduct 1a. Compound **1** was irradiated in $CDCl_3$ as described above and the progress was followed by 1H NMR spectroscopy. After 60 min photoirradiation, 99% of photoproduct formation was observed. 1H NMR (300 MHz, $CDCl_3$, 298 K, δ ppm): 9.75 (s, 2H), 9.36 (s, 1H), 9.14 (s, 2H), 8.01 (s, 2H), 7.34 (s, 2H), 7.32 (s, 2H), 6.79 (s, 2H), 5.69 (s, 2H), 5.31 (s, 2H), 4.77 (s, 1H), 3.95 (s, 6H), 3.90 – 3.71 (m, 8H), 2.16 (m, 4H), 2.11 (s, 6H), 2.07 (s, 6H), 1.10 (m, 24H). ^{13}C NMR (75 MHz, $CDCl_3$, 298 K, δ ppm): 165.7, 163.5, 162.9, 161.9, 161.3, 148.9, 148.3, 147.3, 137.3, 137.0, 136.6, 130.0, 129.5, 128.1, 127.3, 124.8, 119.5, 118.9, 107.4, 103.9, 93.2, 93.2, 77.2, 75.1, 75.0, 56.1, 53.0, 33.2, 28.3, 28.3, 19.3, 19.0, 18.1, 17.8. HRMS (ESI): m/z calcd for $C_{68}H_{73}N_{10}O_{14}$ $[M+H]^+$ 1253.5302, found 1253.5358



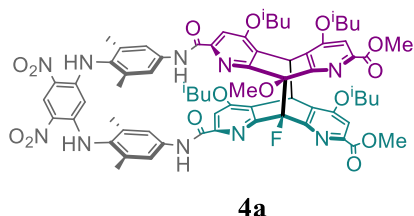
2a

Photoproduct 2a. Compound **2** was irradiated in $CDCl_3$ as described above and the progress was followed by 1H NMR spectroscopy. After 60 min photoirradiation, 99% of photoproduct formation was

observed. ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 9.93 (s, 1H), 9.75 (s, 1H), 9.14 (s, 1H), 8.00 (dd, $J = 6.7, 2.4$ Hz, 2H), 7.41 (s, 1H), 7.40 (s, 1H), 7.38 (s, 1H), 7.35 (s, 1H), 6.77 (t, $J = 2.7$ Hz, 2H), 5.81 – 5.67 (m, 2H), 5.39 (d, $J = 28.7$ Hz, 1H), 4.76 (s, 1H), 3.95 (s, 6H), 3.93 – 3.68 (m, 8H), 2.27 – 2.00 (m, 16H), 1.16 – 1.00 (m, 24H). ^{19}F NMR (282 MHz, CDCl_3 , 298 K, δ ppm): -159.2 (d, $J = 28.5$ Hz). $^{19}\text{F}\{^1\text{H}\}$ NMR (282 MHz, CDCl_3 , 298 K, δ ppm): -159.2. ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 165.8, 162.1, 161.5, 161.3, 160.5, 160.2, 159.9, 149.5, 149.3, 148.4, 148.4, 148.1, 147.9, 137.4, 137.1, 137.0, 136.8, 136.8, 130.3, 129.8, 127.8, 127.2, 124.9, 124.5, 119.7, 119.6, 119.3, 108.2, 107.7, 104.8, 104.4, 93.3, 75.5, 75.4, 75.3, 53.2, 53.1, 33.0, 32.8, 28.4, 19.4, 19.1, 18.2, 17.9. HRMS (ESI): m/z calcd for $\text{C}_{68}\text{H}_{72}\text{FN}_{10}\text{O}_{14}$ $[\text{M}+\text{H}]^+$ 1271.5208, Found 1271.5283.

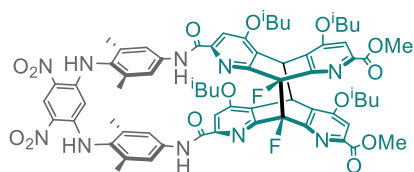


Photoproduct 3a. Compound **3** was irradiated in CDCl_3 as described above and the progress was followed by ^1H NMR spectroscopy. After 60 min photoirradiation, 99% of photoproduct formation was observed. ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 10.10 (s, 1H), 9.91 (s, 1H), 9.11 (s, 2H), 8.09 – 7.95 (m, 2H), 7.39 (s, 1H), 7.34 (s, 1H), 7.33 (s, 1H), 7.27 (s, 1H), 6.83 (d, $J = 2.4$ Hz, 1H), 6.77 (d, $J = 2.4$ Hz, 1H), 5.77 (d, $J = 10.9$ Hz, 1H), 5.64 (d, $J = 10.9$ Hz, 1H), 5.20 (s, 1H), 4.74 (s, 1H), 3.94 (s, 3H), 3.92 (s, 3H), 3.91 – 3.65 (m, 12H), 2.18 (m, 4H), 2.11 (d, $J = 1.4$ Hz, 6H), 2.07 (d, $J = 4.6$ Hz, 6H), 1.16 – 1.03 (m, 24H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 166.1, 166.0, 163.1, 162.6, 162.4, 161.9, 161.8, 161.7, 161.5, 161.4, 161.3, 149.2, 149.1, 148.5, 148.4, 147.8, 147.7, 137.4, 137.3, 137.3, 137.2, 136.7, 136.7, 130.3, 130.2, 129.6, 127.8, 127.2, 126.2, 126.1, 125.0, 124.9, 120.1, 119.9, 119.7, 119.6, 107.5, 107.3, 104.2, 104.1, 93.5, 88.9, 75.4, 75.2, 75.1, 67.3, 56.4, 53.1, 52.9, 33.4, 32.3, 28.4, 19.4, 19.4, 19.2, 19.1, 18.2, 17.9. HRMS (ESI): m/z calcd for $\text{C}_{69}\text{H}_{75}\text{N}_{10}\text{O}_{15}$ $[\text{M}+\text{H}]^+$ 1283.5408, Found 1283.5484.



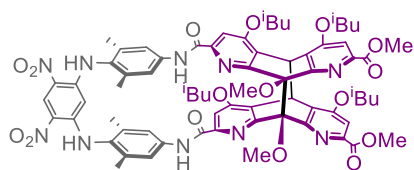
Photoproduct 4a. Compound **4** was irradiated in CDCl_3 as described above and the progress was followed by ^1H NMR spectroscopy. After 60 min photoirradiation, 99% of photoproduct formation was observed. ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 10.13 (s, 1H), 10.09 (s, 1H), 9.10 (s, 2H), 8.02 (d, $J = 2.4$ Hz, 1H), 7.97 (d, $J = 2.4$ Hz, 1H), 7.42 (s, 1H), 7.39 (s, 1H), 7.33 (d, $J = 1.8$ Hz, 2H), 6.88

(d, $J = 2.4$ Hz, 1H), 6.81 (d, $J = 2.4$ Hz, 1H), 5.82 (d, $J = 10.9$ Hz, 1H), 5.74 (d, $J = 10.8$ Hz, 1H), 4.73 (s, 1H), 3.93 (s, 3H), 3.91 (s, 3H), 3.90 – 3.69 (m, 12H), 2.27 – 2.12 (m, 4H), 2.10 (d, $J = 2.4$ Hz, 6H), 2.06 (d, $J = 4.1$ Hz, 6H), 1.16 – 1.04 (m, 24H). ^{19}F NMR (282 MHz, CDCl_3 , 298 K, δ ppm): -175.4. ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 166.1, 165.9, 162.3, 161.6, 161.5, 161.4, 160.6, 160.4, 149.6, 149.5, 148.5, 148.4, 148.2, 148.1, 137.2, 137.2, 137.1, 136.7, 136.6, 130.4, 130.2, 129.6, 126.4, 125.0, 124.9, 124.3, 120.4, 120.0, 119.9, 108.0, 107.6, 104.6, 93.5, 75.5, 75.4, 56.7, 53.0, 52.9, 33.6, 32.2, 28.4, 19.4, 19.4, 19.1, 18.2, 17.9. HRMS (ESI): m/z calcd for $\text{C}_{69}\text{H}_{74}\text{FN}_{10}\text{O}_{15}$ $[\text{M}+\text{H}]^+$ 1301.5314, Found 1301.5402.



5a

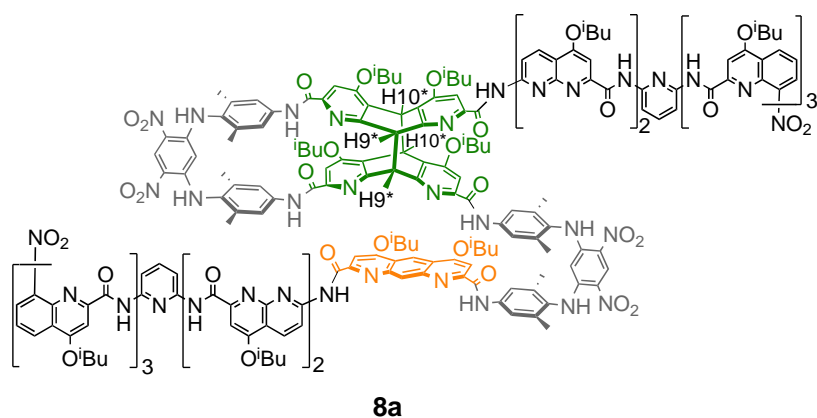
Photoproduct 5a. Compound **5** was irradiated in CDCl_3 as described above and the progress was followed by ^1H NMR spectroscopy. After 60 min photoirradiation, 99% of photoproduct formation was observed. ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 9.94 (s, 2H), 9.36 (s, 1H), 9.14 (s, 2H), 7.99 (d, $J = 2.4$ Hz, 2H), 7.44 (s, 2H), 7.41 (s, 2H), 6.81 (d, $J = 2.4$ Hz, 2H), 5.81 (s, 2H), 4.76 (s, 1H), 3.95 (s, 6H), 3.93 – 3.71 (m, 8H), 2.18 (m, 4H), 2.12 (s, 6H), 2.07 (s, 6H), 1.15 – 1.03 (m, 24H). ^{19}F NMR (282 MHz, CDCl_3 , 298 K, δ ppm): -178.0. ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 165.7, 161.9, 161.3, 161.1, 149.7, 148.4, 137.3, 136.9, 136.8, 130.3, 129.7, 124.9, 124.7, 119.7, 108.4, 105.1, 93.3, 75.6, 75.5, 53.2, 32.9, 28.4, 19.4, 19.1, 18.2, 17.9. HRMS (ESI): m/z calcd for $\text{C}_{68}\text{H}_{71}\text{F}_2\text{N}_{10}\text{O}_{14}$ $[\text{M}+\text{H}]^+$ 1289.5114, found 1289.5207.



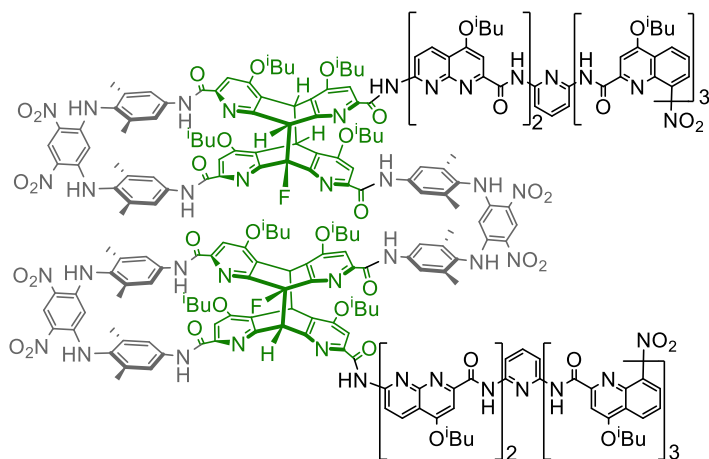
6a

Photoproduct 6a. Compound **7** was irradiated in CDCl_3 as described above and the progress was followed by ^1H NMR spectroscopy. After 60 min photoirradiation, 99% of photoproduct formation was observed. ^1H NMR (300 MHz, CDCl_3 , 298 K, δ ppm): 10.38 (s, 2H), 9.34 (s, 1H), 9.06 (s, 2H), 8.02 (d, $J = 2.4$ Hz, 2H), 7.37 (s, 2H), 7.23 (s, 2H), 6.91 (d, $J = 2.4$ Hz, 2H), 5.74 (s, 2H), 4.71 (s, 1H), 3.90 (s, 6H), 3.88 – 3.67 (m, 8H), 3.59 (s, 6H), 2.22 – 2.13 (m, 4H), 2.10 (s, 6H), 2.06 (s, 6H), 1.17 – 1.03 (m, 24H). ^{13}C NMR (75 MHz, CDCl_3 , 298 K, δ ppm): 166.4, 162.2, 162.1, 161.6, 161.5, 161.2, 149.3, 148.4, 147.8, 137.5, 137.1, 136.5, 130.3, 129.5, 126.3, 124.9, 120.5, 107.1, 104.2, 93.8, 89.5, 75.4, 75.2, 56.6,

52.8, 33.0, 28.5, 28.4, 19.4, 19.4, 19.1, 18.2, 18.0. HRMS (ESI): m/z calcd for $C_{70}H_{77}N_{10}O_{16}$ $[M+H]^+$ 1313.5514, Found 1313.5618.



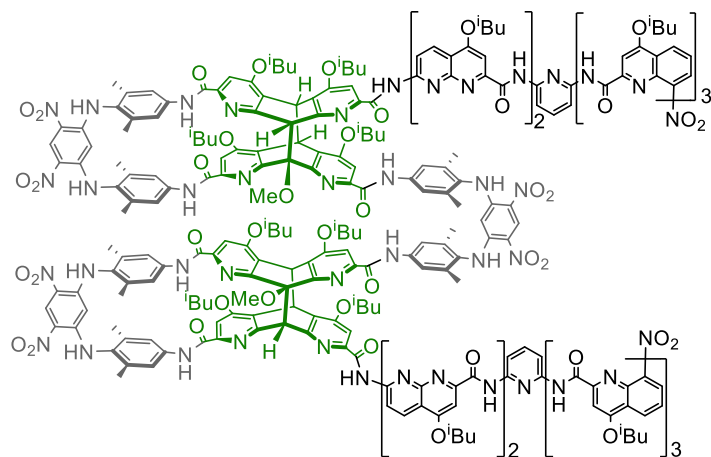
Photoproduct 8a. Compound **8** was irradiated in $CDCl_3$ as described above and the progress was followed by 1H NMR spectroscopy. After 90 min photoirradiation, photoproduct **8a** was obtained. 1H NMR (700 MHz, $CDCl_3$, 298 K, δ ppm): 11.81, 11.71, 11.69, 11.60, 11.58, 11.54, 11.45, 11.28, 11.21, 11.01, 10.99, 10.97, 10.90, 10.80, 10.26, 9.98, 9.89, 9.86, 9.53, 9.44, 9.37, 9.20, 9.15, 9.07, 9.05, 9.02, 8.94, 8.93, 8.90, 8.86, 8.85, 8.83, 8.81, 8.80, 8.75, 8.74, 8.73, 8.67, 8.66, 8.57, 8.48, 8.42, 8.36, 8.33, 8.32, 8.31, 8.28, 8.27, 8.26, 8.22, 8.21, 8.19, 8.15, 8.14, 8.12, 8.08, 8.05, 8.04, 7.94, 7.86, 7.83, 7.81, 7.75, 7.74, 7.73, 7.66, 7.59, 7.57, 7.51, 7.47, 7.46, 7.45, 7.41, 7.40, 7.28, 7.26, 7.22, 7.20, 7.19, 7.18, 7.15, 7.14, 7.12, 7.10, 7.08, 7.04, 7.02, 7.01, 7.00, 6.99, 6.97, 6.97, 6.96, 6.96, 6.93, 6.93, 6.89, 6.88, 6.87, 6.85, 6.84, 6.79, 6.78, 6.70, 6.57, 6.52, 6.39, 6.27, 6.24, 6.23, 6.22, 6.19, 6.18, 6.17, 6.13, 6.05, 6.00, 5.88, 5.80, 5.71, 5.64, 5.35, 5.19, 5.18, 5.06, 5.04, 4.68, 4.32, 4.31, 4.28, 4.27, 4.22, 4.21, 4.20, 4.19, 4.19, 4.18, 4.17, 4.16, 4.15, 4.13, 4.12, 4.11, 4.11, 4.10, 4.09, 4.08, 3.96, 3.95, 3.91, 3.90, 3.89, 3.88, 3.87, 3.85, 3.83, 3.82, 3.80, 3.75, 3.75, 3.74, 3.74, 3.73, 3.66, 3.65, 3.64, 3.63, 3.61, 3.54, 3.53, 3.48, 3.45, 3.27, 3.24, 3.03, 3.01, 3.00, 2.82, 2.74, 2.46, 2.45, 2.44, 2.43, 2.42, 2.41, 2.40, 2.39, 2.39, 2.35, 2.34, 2.33, 2.32, 2.31, 2.27, 2.26, 2.25, 2.24, 2.23, 2.22, 2.21, 2.13, 2.12, 2.12, 2.01, 2.00, 1.97, 1.96, 1.91, 1.86, 1.86, 1.85, 1.85, 1.84, 1.82, 1.82, 1.80, 1.75, 1.60, 1.45, 1.44, 1.43, 1.41, 1.40, 1.39, 1.35, 1.34, 1.34, 1.33, 1.33, 1.32, 1.31, 1.30, 1.29, 1.26, 1.25, 1.24, 1.24, 1.23, 1.22, 1.22, 1.21, 1.21, 1.21, 1.20, 1.19, 1.17, 1.17, 1.16, 1.15, 1.14, 1.14, 1.13, 1.11, 1.11, 1.10, 1.10, 0.99, 0.89, 0.89, 0.88, 0.88, 0.87, 0.87, 0.86, 0.85, 0.84, 0.78, 0.77, 0.69, 0.57, 0.54, 0.53, 0.53, 0.52, 0.48, 0.47, 0.42, 0.34, 0.33, 0.29, 0.28, 0.25, 0.24, 0.07. HRMS (ES⁺): m/z calcd for $C_{256}H_{257}N_{48}O_{44}$ $[M+3H]^{3+}$ 1569.6466 Found 1569.6514.



9a

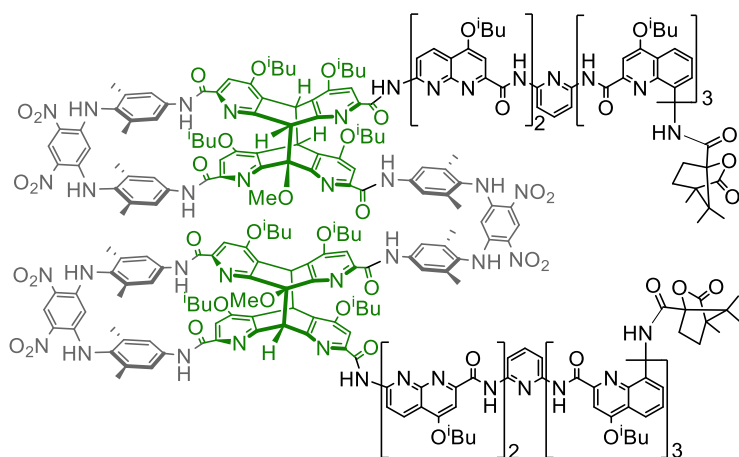
Photoproduct 9a. Compound **9** was irradiated in acetone- d_6 as described above and the progress was followed by ^1H NMR spectroscopy. After 5 hours of photoirradiation, 98% of photoproduct formation was observed. ^1H NMR (700 MHz, Acetone- d_6 , 298 K, δ ppm; only peaks are given due to the presence of two conformers): 11.76, 11.67, 11.61, 11.54, 11.49, 11.48, 11.26, 11.02, 10.97, 10.85, 10.73, 10.62, 10.51, 10.11, 9.95, 9.90, 9.86, 9.84, 9.82, 9.77, 9.75, 9.70, 9.54, 9.51, 9.50, 9.47, 9.41, 9.31, 9.26, 9.25, 9.23, 9.23, 9.21, 9.18, 9.12, 9.08, 9.06, 9.05, 9.00, 8.98, 8.94, 8.92, 8.90, 8.89, 8.87, 8.86, 8.85, 8.84, 8.84, 8.82, 8.80, 8.77, 8.76, 8.67, 8.66, 8.65, 8.60, 8.58, 8.55, 8.46, 8.46, 8.45, 8.44, 8.44, 8.42, 8.41, 8.40, 8.39, 8.38, 8.37, 8.34, 8.32, 8.31, 8.29, 8.27, 8.26, 8.25, 8.23, 8.20, 8.18, 8.17, 8.13, 8.11, 7.92, 7.90, 7.87, 7.83, 7.82, 7.80, 7.78, 7.77, 7.76, 7.74, 7.71, 7.70, 7.69, 7.67, 7.64, 7.60, 7.56, 7.54, 7.52, 7.49, 7.48, 7.47, 7.44, 7.43, 7.42, 7.41, 7.38, 7.32, 7.31, 7.29, 7.27, 7.26, 7.25, 7.25, 7.23, 7.20, 7.19, 7.13, 7.10, 7.05, 7.04, 7.02, 7.00, 6.99, 6.97, 6.96, 6.95, 6.91, 6.90, 6.88, 6.87, 6.86, 6.82, 6.79, 6.76, 6.75, 6.72, 6.65, 6.62, 6.59, 6.56, 6.53, 6.51, 6.49, 6.44, 6.43, 6.42, 6.38, 6.37, 6.36, 6.33, 6.30, 6.25, 6.24, 6.23, 6.21, 6.16, 6.12, 6.10, 6.06, 6.01, 5.98, 5.76, 5.67, 5.62, 5.51, 5.49, 5.45, 5.45, 5.43, 5.42, 5.40, 5.39, 5.37, 5.35, 5.35, 5.34, 5.28, 5.27, 5.07, 4.85, 4.79, 4.75, 4.69, 4.68, 4.68, 4.67, 4.66, 4.59, 4.58, 4.55, 4.52, 4.50, 4.40, 4.39, 4.38, 4.37, 4.36, 4.35, 4.34, 4.33, 4.32, 4.31, 4.28, 4.27, 4.27, 4.26, 4.25, 4.23, 4.22, 4.21, 4.20, 4.19, 4.16, 4.15, 4.14, 4.13, 4.12, 4.11, 4.10, 4.09, 4.07, 4.06, 4.05, 4.04, 4.04, 4.03, 4.02, 4.01, 3.99, 3.98, 3.97, 3.96, 3.96, 3.92, 3.91, 3.90, 3.88, 3.87, 3.86, 3.83, 3.82, 3.81, 3.76, 3.75, 3.73, 3.72, 3.71, 3.65, 3.64, 3.61, 3.58, 3.44, 3.38, 3.30, 3.28, 3.27, 3.25, 3.24, 3.24, 3.23, 3.23, 3.22, 3.22, 3.16, 3.15, 3.14, 3.11, 2.97, 2.92, 2.91, 2.90, 2.81, 2.43, 2.42, 2.41, 2.40, 2.39, 2.38, 2.37, 2.36, 2.35, 2.34, 2.33, 2.32, 2.31, 2.30, 2.29, 2.29, 2.28, 2.26, 2.25, 2.24, 2.22, 2.17, 2.16, 2.15, 2.14, 2.14, 2.14, 2.13, 2.13, 2.12, 2.11, 2.10, 2.08, 2.07, 2.07, 2.06, 2.06, 2.05, 2.05, 2.05, 2.04, 2.02, 1.99, 1.99, 1.98, 1.97, 1.96, 1.96, 1.96, 1.95, 1.95, 1.93, 1.88, 1.86, 1.84, 1.82, 1.78, 1.74, 1.70, 1.67, 1.64, 1.63, 1.62, 1.61, 1.60, 1.59, 1.57, 1.56, 1.54, 1.48, 1.47, 1.45, 1.44, 1.42, 1.41, 1.40, 1.38, 1.37, 1.37, 1.36, 1.36, 1.35, 1.35, 1.34, 1.32, 1.31, 1.30, 1.30, 1.29, 1.27, 1.26, 1.25, 1.25, 1.24, 1.24, 1.23, 1.22, 1.21, 1.21, 1.20, 1.19, 1.17, 1.16, 1.15, 1.14, 1.13, 1.13, 1.12, 1.10, 1.03, 1.02, 1.00, 0.97, 0.96,

0.95, 0.93, 0.92, 0.91, 0.90, 0.89, 0.88, 0.87, 0.86, 0.85, 0.85, 0.83, 0.82, 0.77, 0.76, 0.69, 0.68, 0.67, 0.63, 0.60, 0.59, 0.58, 0.56, 0.56, 0.47, 0.46, 0.45, 0.44, 0.21, 0.17, 0.15, 0.13, 0.12, 0.10, 0.09, 0.09, 0.08, 0.07, 0.04. HRMS (ES⁺): m/z calcd for C₃₀₀H₂₉₉F₂N₅₆O₅₂ [M+3H]³⁺: 1852.7509 found 1852.7573.



10a

Photoproduct 10a. Compound **10** was irradiated in CDCl₃ as described above and the progress was followed by ¹H NMR spectroscopy. After 60 min of photoirradiation, 99% of photoproduct formation was observed. ¹H NMR (700 MHz, CDCl₃, 298 K, δ ppm; only peaks are given due to the presence of two conformers): 11.82, 11.75, 11.60, 11.53, 11.00, 10.87, 10.83, 10.72, 9.83, 9.81, 9.75, 9.37, 9.35, 9.31, 9.24, 9.14, 8.94, 8.91, 8.86, 8.79, 8.78, 8.74, 8.73, 8.63, 8.58, 8.50, 8.45, 8.44, 8.40, 8.39, 8.39, 8.38, 8.36, 8.35, 8.33, 8.32, 8.25, 8.17, 8.08, 8.00, 7.96, 7.85, 7.79, 7.77, 7.70, 7.64, 7.63, 7.61, 7.61, 7.59, 7.50, 7.48, 7.45, 7.41, 7.31, 7.30, 7.26, 7.24, 7.23, 7.22, 7.21, 7.20, 7.19, 7.18, 7.15, 7.14, 7.11, 7.09, 7.06, 7.03, 6.99, 6.98, 6.95, 6.94, 6.91, 6.84, 6.79, 6.77, 6.74, 6.70, 6.55, 6.43, 6.33, 6.32, 6.31, 6.27, 6.22, 6.17, 5.97, 5.92, 5.37, 5.36, 5.32, 5.30, 5.23, 4.70, 4.60, 4.51, 4.42, 4.26, 4.25, 4.24, 4.21, 4.20, 4.19, 4.18, 4.17, 4.13, 4.11, 4.10, 4.09, 4.03, 4.02, 4.01, 3.99, 3.98, 3.97, 3.94, 3.93, 3.92, 3.89, 3.88, 3.81, 3.75, 3.73, 3.72, 3.71, 3.70, 3.69, 3.68, 3.65, 3.57, 3.56, 3.55, 3.50, 3.49, 3.48, 3.39, 3.31, 3.30, 3.29, 3.05, 2.99, 2.65, 2.55, 2.41, 2.40, 2.39, 2.38, 2.37, 2.37, 2.36, 2.35, 2.34, 2.33, 2.32, 2.31, 2.30, 2.29, 2.24, 2.23, 2.22, 2.21, 2.20, 2.19, 2.18, 2.17, 2.13, 2.12, 2.11, 2.10, 2.03, 2.02, 2.01, 2.00, 1.99, 1.95, 1.92, 1.88, 1.85, 1.82, 1.80, 1.61, 1.56, 1.53, 1.40, 1.33, 1.31, 1.30, 1.29, 1.28, 1.26, 1.25, 1.24, 1.23, 1.22, 1.20, 1.19, 1.19, 1.18, 1.18, 1.17, 1.16, 1.15, 1.15, 1.15, 1.14, 1.12, 1.11, 1.10, 1.09, 1.08, 1.05, 1.02, 1.01, 0.97, 0.95, 0.94, 0.93, 0.92, 0.89, 0.88, 0.87, 0.85, 0.84, 0.84, 0.76, 0.75, 0.74, 0.73, 0.59, 0.58, 0.56, 0.42, 0.15, 0.09, 0.07, 0.04, -0.02. HRMS (ES⁺): m/z calcd for C₃₀₂H₃₀₂N₅₆O₅₄ [M+3H]³⁺: 1860.7642 found 1860.7721.



11a

Photoproduct 11a. Compound **11** was irradiated in CDCl_3 as described above and the progress was followed by ^1H NMR spectroscopy. After 40 min of photoirradiation, a quantitative photoproduct formation was observed. ^1H NMR (700 MHz, CDCl_3 , 298 K, δ ppm): 11.89, 11.82, 11.13, 10.88, 9.86, 9.77, 9.65, 9.60, 9.39, 9.37, 8.93, 8.78, 8.75, 8.46, 8.38, 8.24, 7.97, 7.88 - 7.76, 7.70, 7.68, 7.58, 7.49, 7.36, 7.31, 7.11 - 7.01, 6.95, 6.89, 6.74, 6.62, 6.42, 6.39, 6.29, 6.15, 5.32, 5.26, 4.54, 4.43, 4.24, 4.18, 4.09, 4.03, 3.89, 3.70, 3.66, 3.61, 3.53, 3.47, 3.27, 3.06, 2.78, 2.36, 2.19, 2.05 - 1.94, 1.92, 1.80, 1.50, 1.45, 1.40, 1.37 - 1.12, 1.09, 0.92, 0.89 - 0.78, 0.72, 0.63 - 0.55, 0.53, 0.41, 0.15, 0.07. HRMS (ES⁺): m/z calcd for $\text{C}_{322}\text{H}_{333}\text{N}_{56}\text{O}_{56}$ $[\text{M}+3\text{H}]^{3+}$: 1960.8338 found 1960.8473.

3. Photoirradiation of aromatic sheets and thermal reversibility

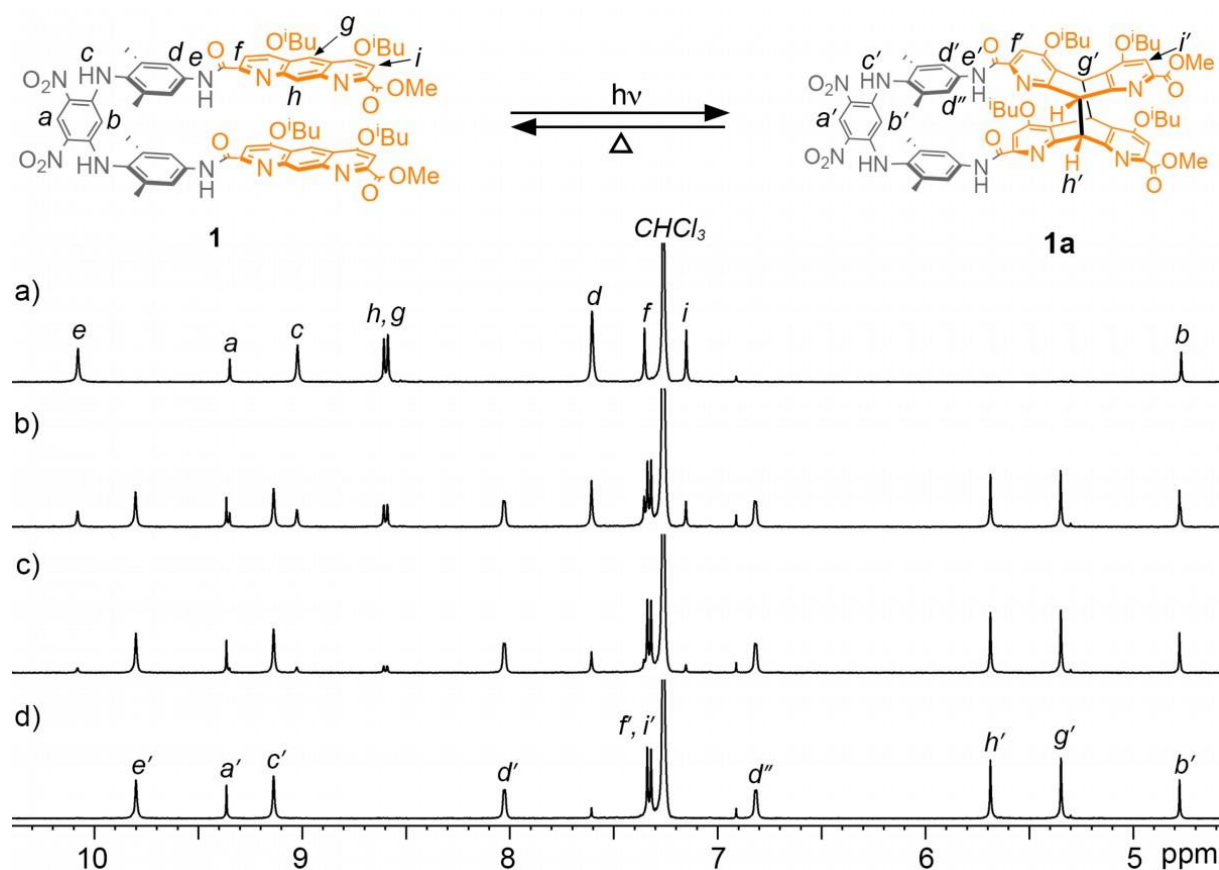


Figure S1. Excerpts from the ¹H NMR spectra (300 MHz, 298K) in CDCl₃ showing the photoproduct formation of **1** (5 mM): (a) before irradiation and after (b): 5 min; (c) 10 min; (d) 20 min of irradiation with assignment of proton resonances.

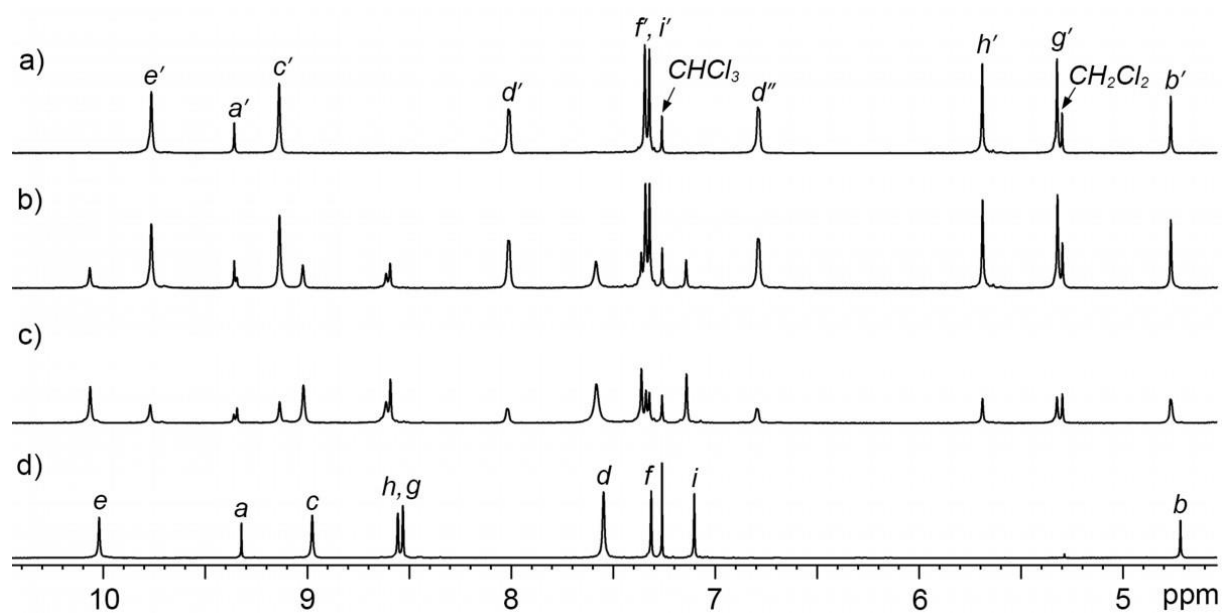


Figure S2. Excerpts from the ¹H NMR spectra (300 MHz, 298K) of **1a** (5 mM) in (a) CDCl₃ and after heating it for (b) 5 h; (c) 15 h; and (d) 30 h in CDCl₃ at 333 K.

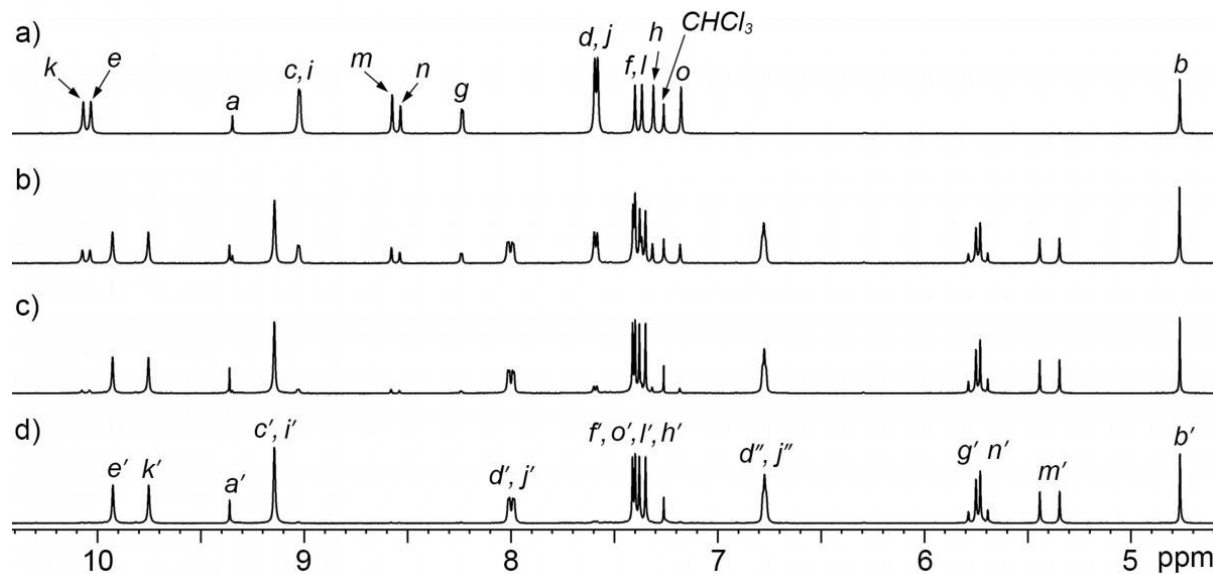
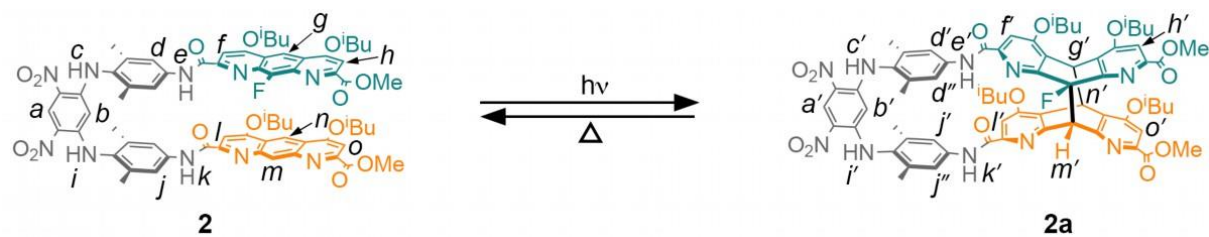


Figure S3. Excerpts from the ^1H NMR spectra (300 MHz, 298K) in CDCl_3 showing the photoproduct formation of **2** (5 mM): (a) before irradiation and after (b): 20 min; (c) 40 min; (d) 60 min of irradiation with assignment of proton resonances.

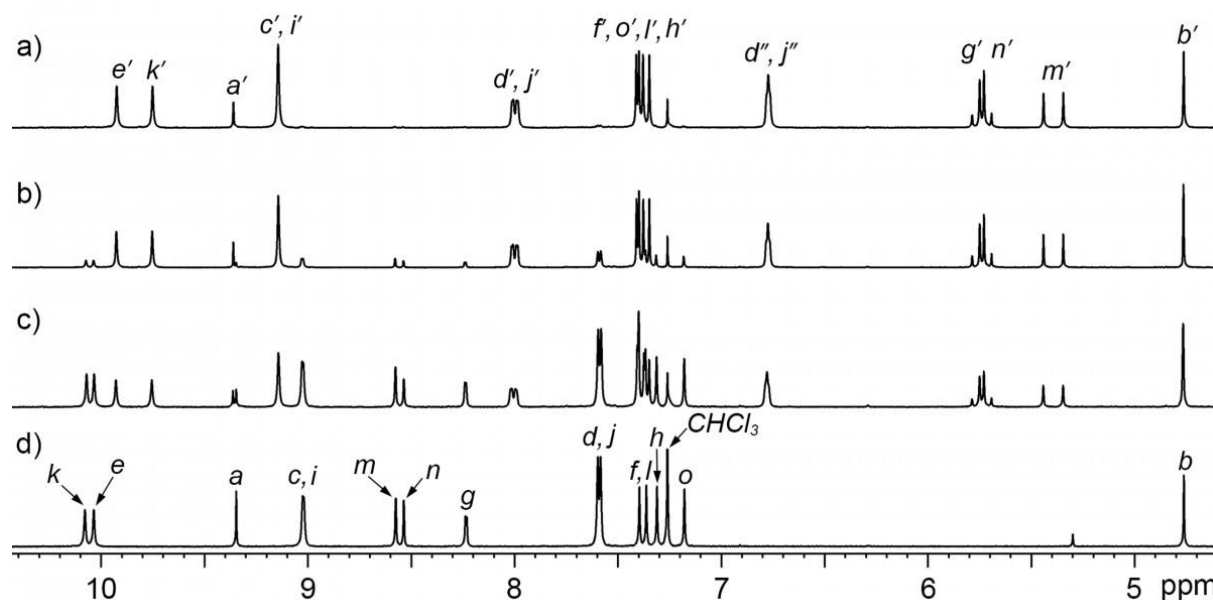


Figure S4. Excerpts from the ^1H NMR spectra (300 MHz, 298K) of **2a** (5 mM) in (a) CDCl_3 and after heating it for (b) 5 h; (c) 15 h; and (d) 30 h in CDCl_3 at 333 K.

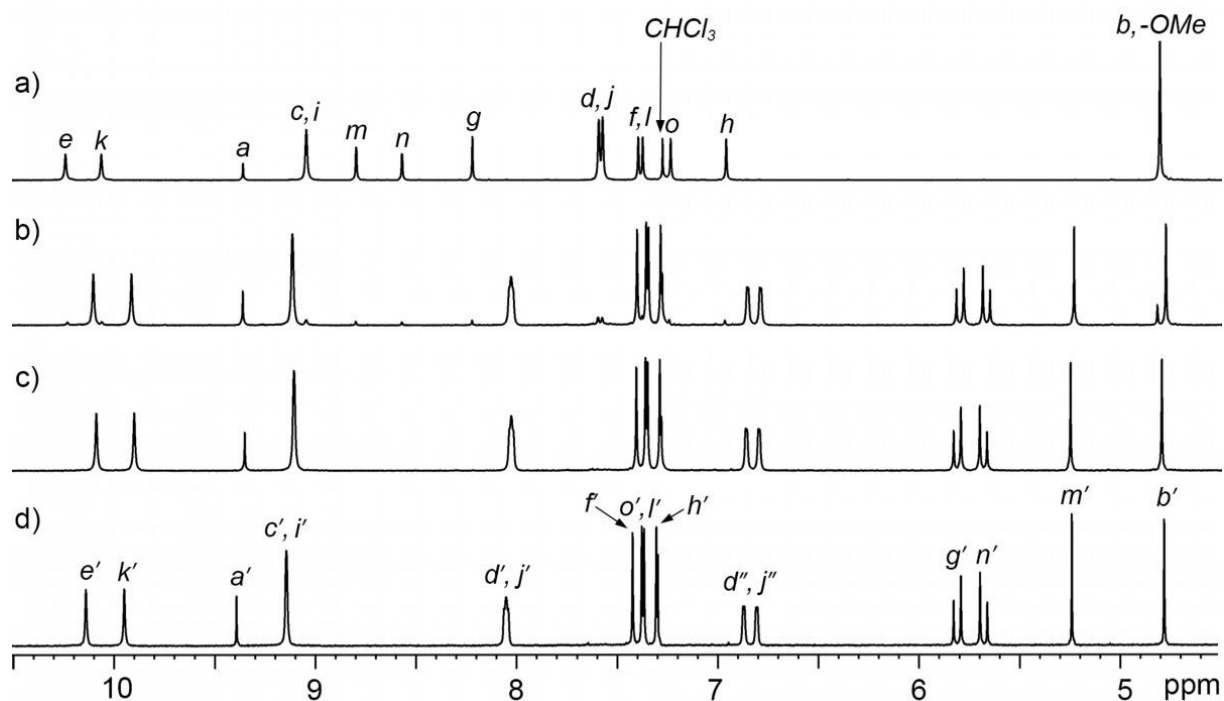
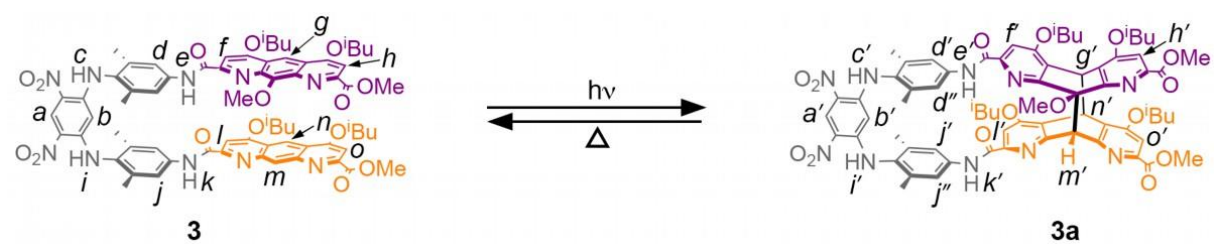


Figure S5. Excerpts from the ^1H NMR spectra (300 MHz, 298K) in CDCl_3 showing the photoproduct formation of **3** (5 mM): (a) before irradiation and after (b): 20 min.; (c) 40 min.; (d) 60 min of irradiation with assignment of proton resonances.

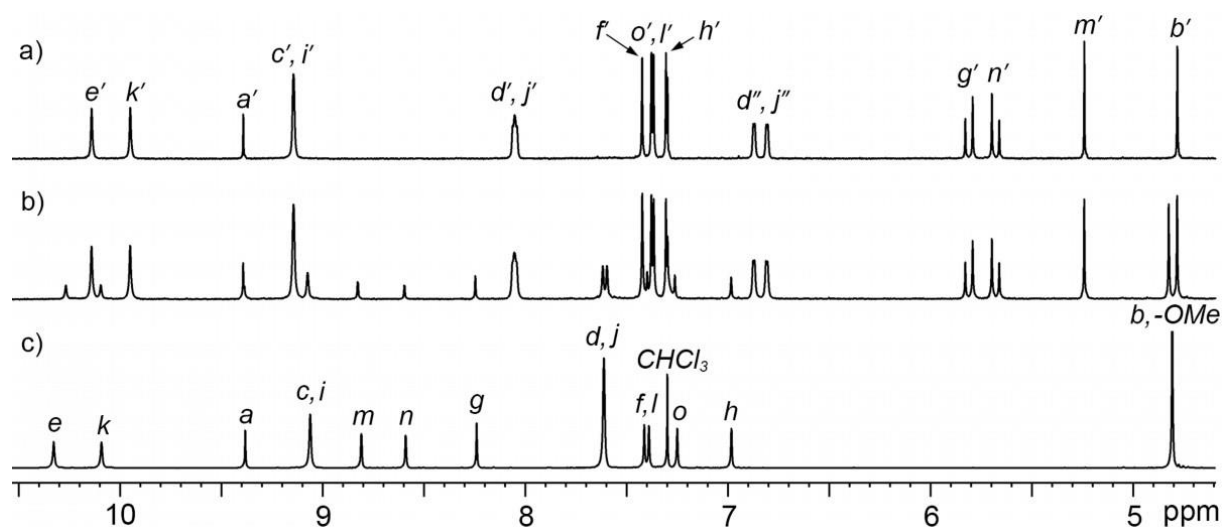


Figure S6. Excerpts from the ^1H NMR spectra (300 MHz, 298K) of **3a** (5 mM) in (a) CDCl_3 and after heating it for (b) 5 h; and (c) 30 h in CDCl_3 at 333 K.

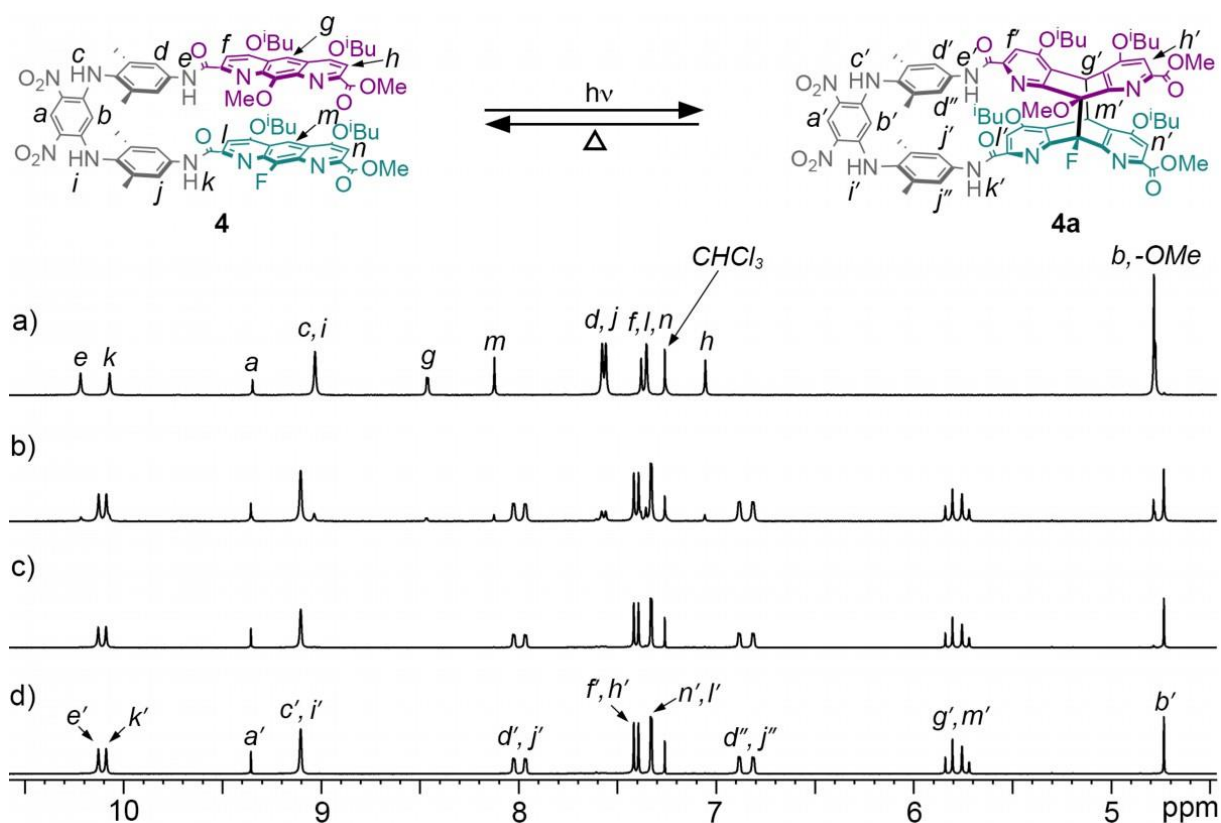


Figure S7. Excerpts from the ^1H NMR spectra (300 MHz, 298K) in CDCl_3 showing the photoproduct formation of **4** (5 mM): (a) before irradiation and after (b): 20 min.; (c) 40 min.; (d) 60 min of irradiation with assignment of proton resonances.

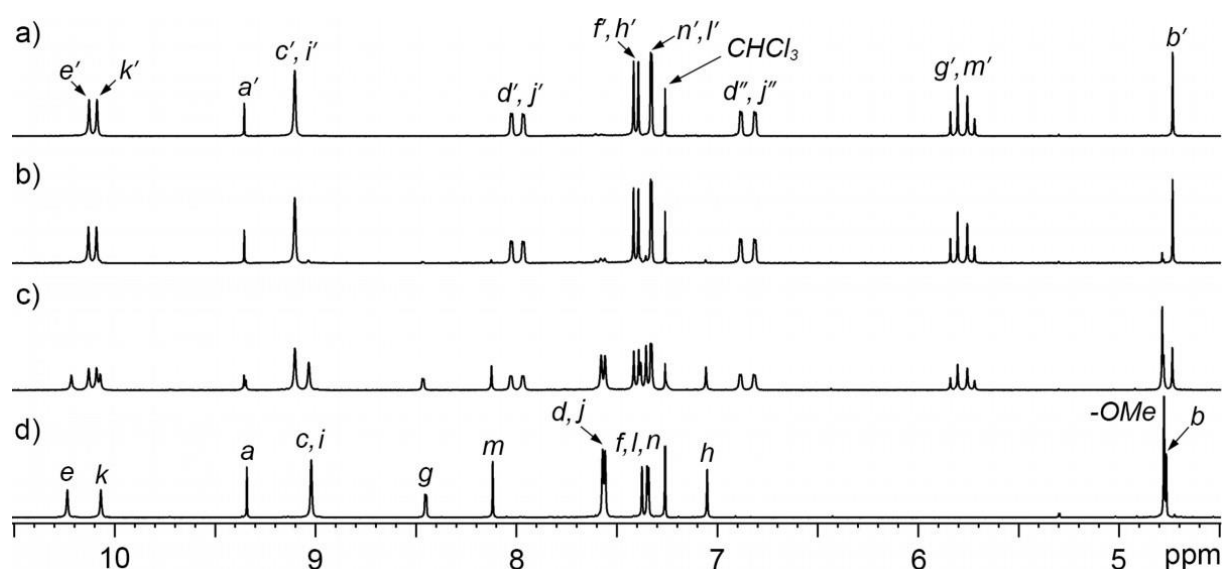


Figure S8. Excerpts from the ^1H NMR spectra (300 MHz, 298K) of **4a** (5 mM) in (a) CDCl_3 and after heating it for (b) 5 h; (c) 15 h; and (d) 30 h in CDCl_3 at 333 K.

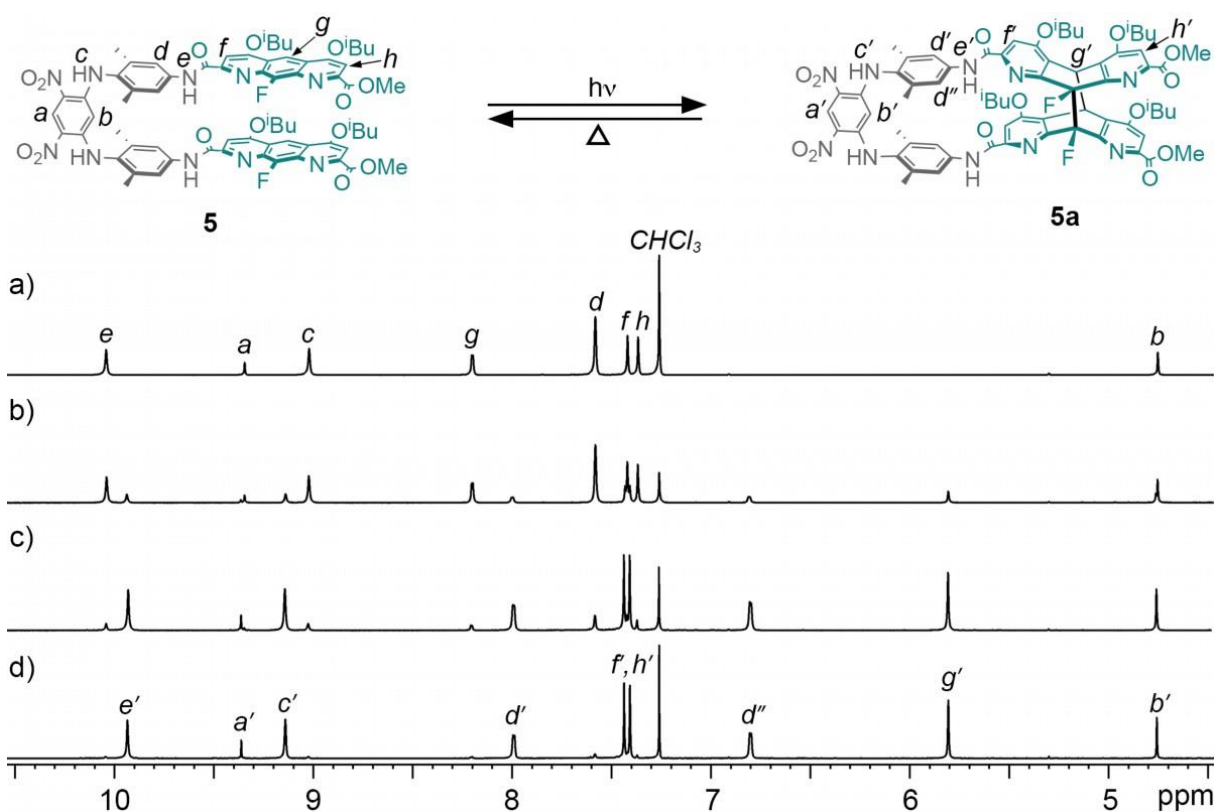


Figure S9. Excerpts from the ^1H NMR spectra (300 MHz, 298K) in CDCl_3 showing the photoproduct formation of **5** (5 mM): (a) before irradiation and after (b): 1 hour; (c) 4 hours; (d) 8 hours of irradiation with assignment of proton resonances.

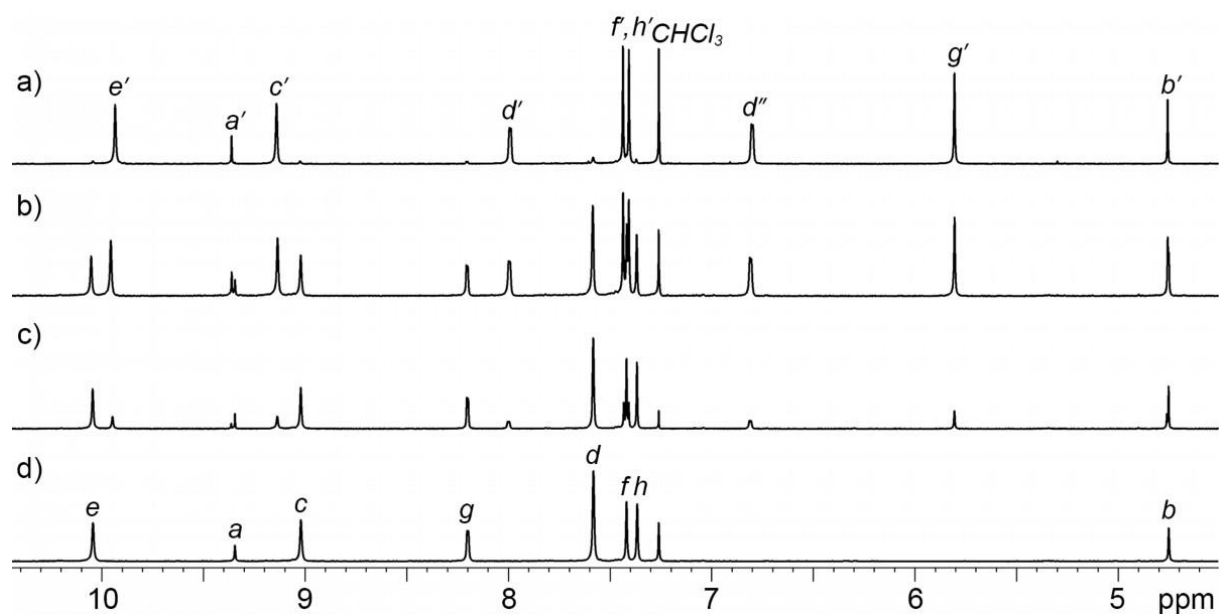


Figure S10. Excerpts from the ^1H NMR spectra (300 MHz, 298K) of **5a** (5 mM) in (a) CDCl_3 and after heating it for (b) 5 h; (c) 15 h; and (d) 30 h in CDCl_3 at 333 K.

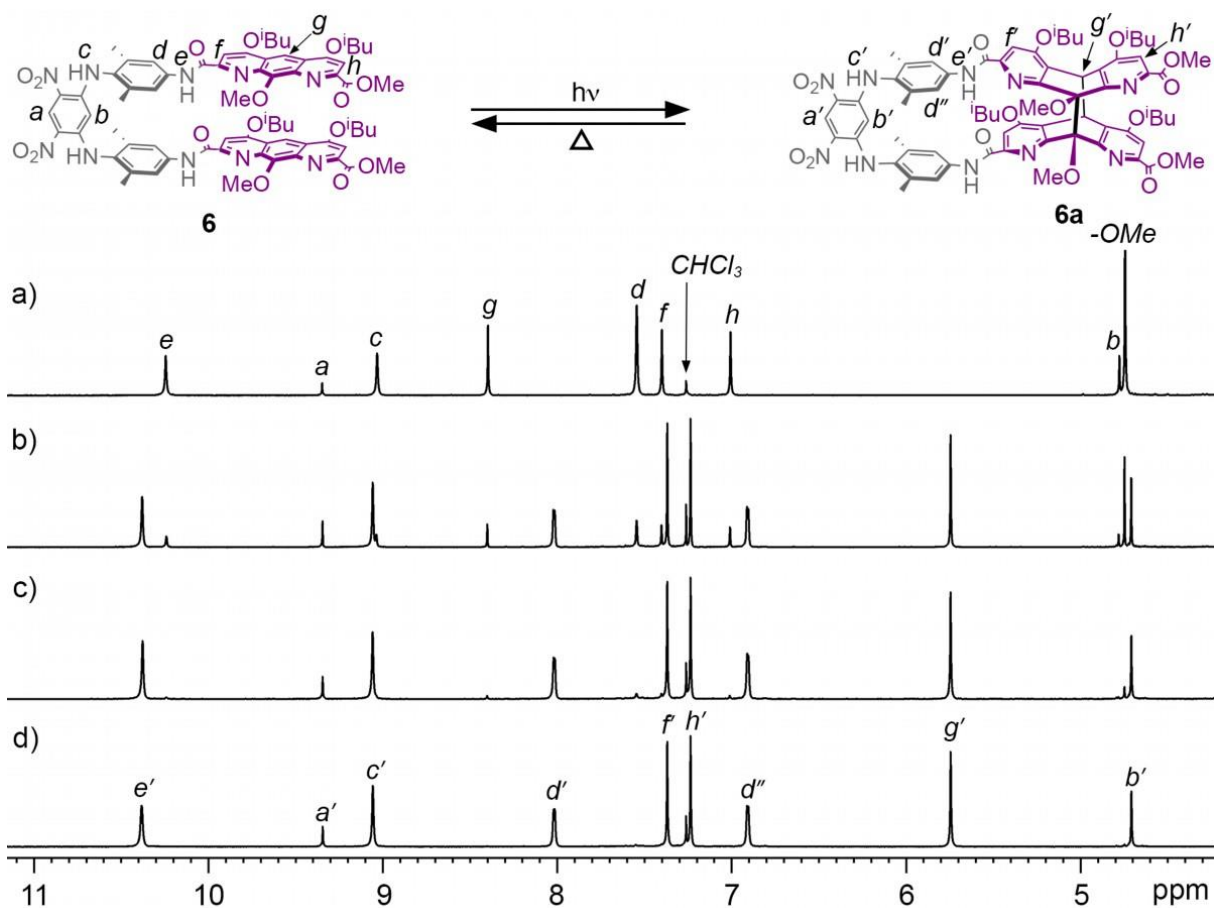


Figure S11. Excerpts from the ^1H NMR spectra (300 MHz, 298K) in CDCl_3 showing the photoproduct formation of **6** (5 mM): (a) before irradiation and after (b): 20 min.; (c) 40 min.; (d) 60 min of irradiation with assignment of proton resonances.

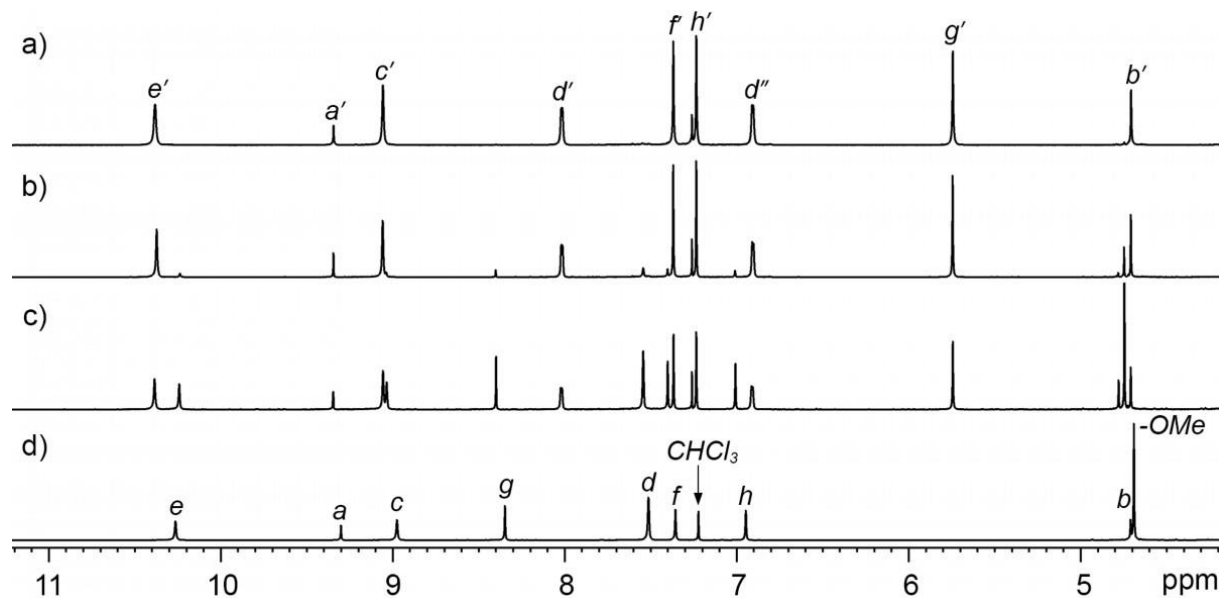


Figure S12. Excerpts from the ^1H NMR spectra (300 MHz, 298K) of **6a** (5 mM) in (a) CDCl_3 and after heating it for (b) 5 h; (c) 15 h; and (d) 30 h in CDCl_3 at 333 K.

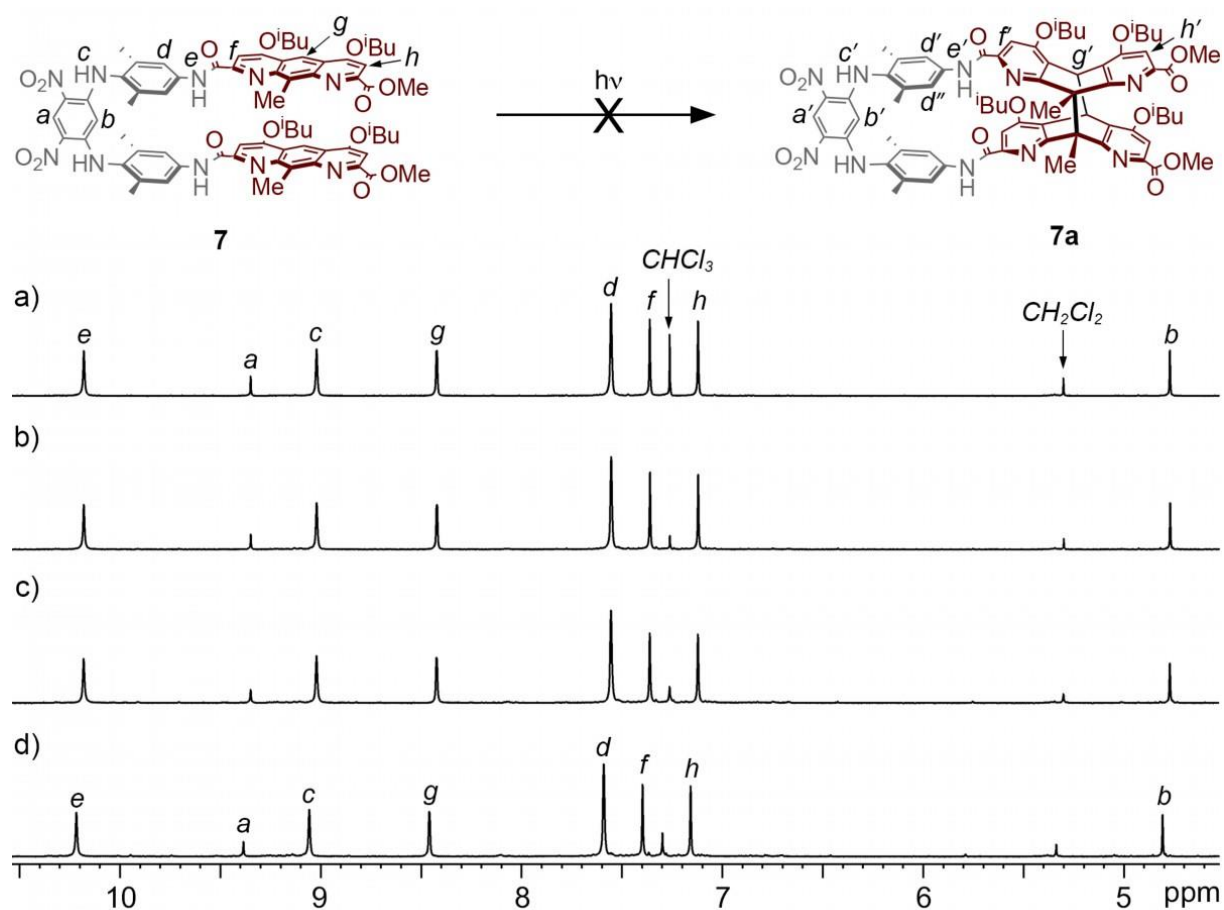


Figure S13. Excerpts from the ^1H NMR spectra (300 MHz, 298K) in CDCl_3 showing the photoirradiation process of **7** (5 mM): (a) before irradiation and after (b): 20 min; (c) 35 min; (d) 50 min of irradiation with assignment of proton resonances.

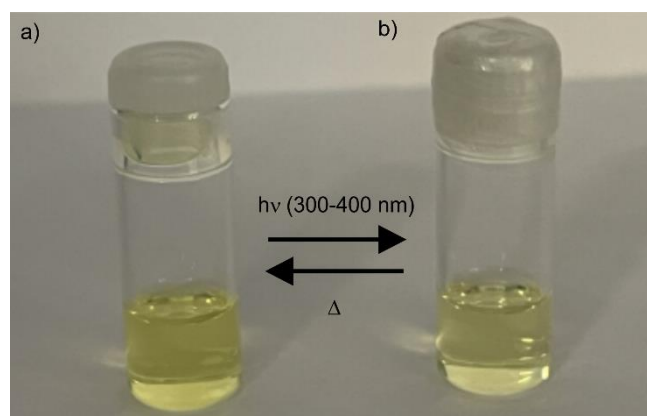


Figure S14. Photography illustrating the slight color change of a solution of **6** (0.5 mM) after irradiation and yielding the corresponding photoproduct **6a** which is less yellow.

5. Photochemistry

4.1 Fluorescence Quantum Yield

The fluorescence and reaction quantum yield were determined in degassed dichloromethane and air-equilibrated solutions as follows. The luminescence quantum yield (Φ) was calculated by using the equation $\Phi_{\text{LUM}} = \Phi_{\text{r}}(I/I_{\text{r}})(A_{\text{r}}/A)(\eta^2/\eta_{\text{r}}^2)$ in which Φ_{r} refers to the quantum yield of the reference, I is the integrated emission intensity, A is the absorbance at the excitation wavelength and η is the refractive index of the solvent. An optically dilute solution of quinine sulphate ($\lambda_{\text{exc}} = 370$ nm) in 1N sulphuric acid was used as the reference, $\Phi_{\text{f}} = 0.54$.

4.2 Luminescence Decay

Luminescence lifetimes were measured via time-correlated single photon counting spectrometry on a Horiba Jobin-Yvon Fluorolog-3 spectrofluorometer, exciting with a 371 nm NanoLED (FWHM = ca. 1 ns).

4.3 Photoreaction quantum yields

Photoreaction quantum yields were determined upon excitation at 405 nm using the couple potassium ferrioxalate-phenanthroline as a chemical actinometer^[8] on an optical bench equipped with a 150 W Hg-Xe lamp and a monochromator. Samples (1.09-1.52 mM for intermolecular photodimérisation in 1 mm cell and 0.022-0.027 mM for intramolecular photodimérisation in 10 mm cell) were stirred during the irradiation and the amount of converted material was determined at 2 min intervals by UV-vis following the disappearance of the absorption band of the diaazanthracene moieties at 400 nm. The error in photodimerisation quantum yield determination was estimated at $\pm 10\%$.

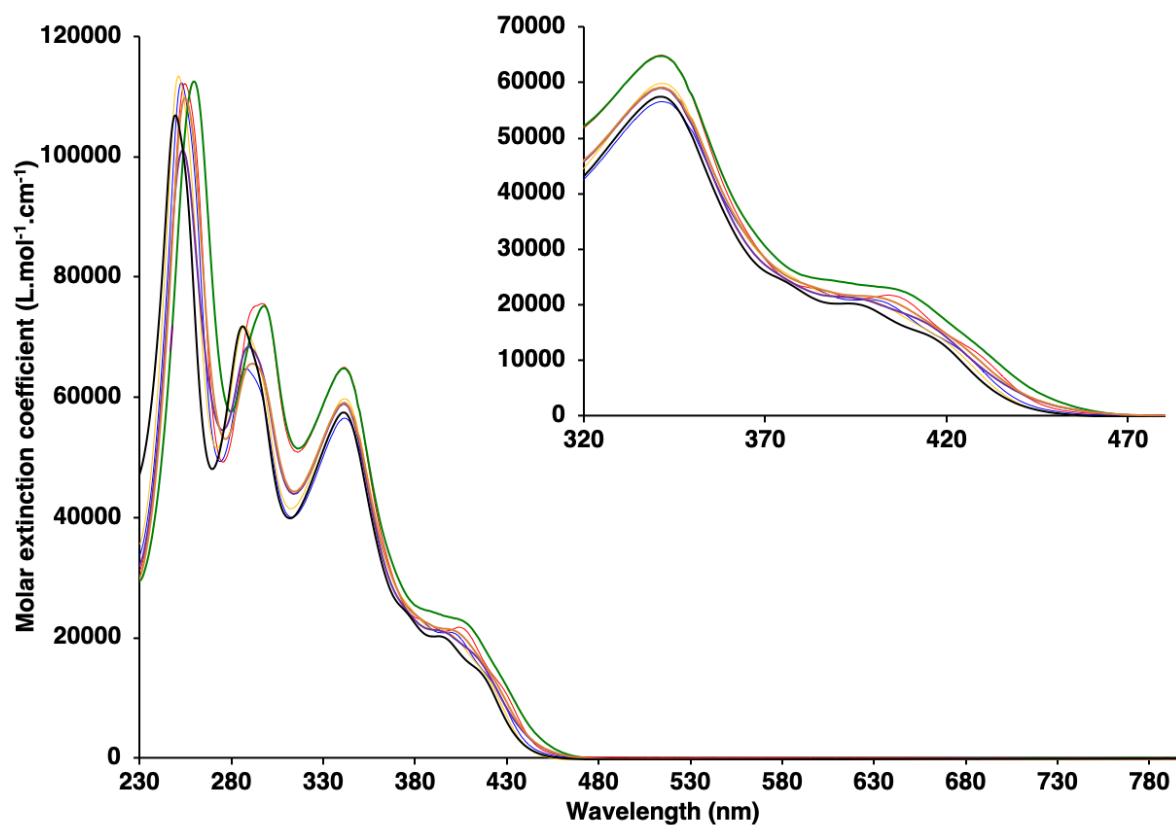


Figure S15: Electronic absorption spectra of a degassed solution of **1** (black), **2** (yellow), **3** (purple), **4** (orange), **5** (blue), **6** (green), and **7** (red) in CH_2Cl_2 .

6. Nuclear Magnetic Resonance

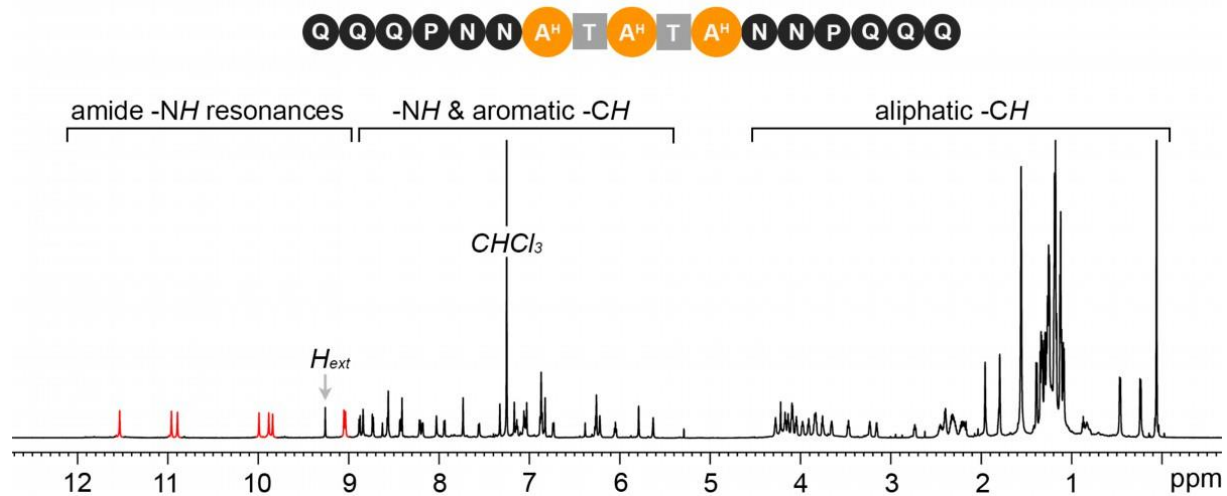


Figure S16. 700 MHz ^1H NMR spectra at 298 K of oligomer **8** in CDCl_3 . The amide $-\text{NH}$ signals are highlighted in red.

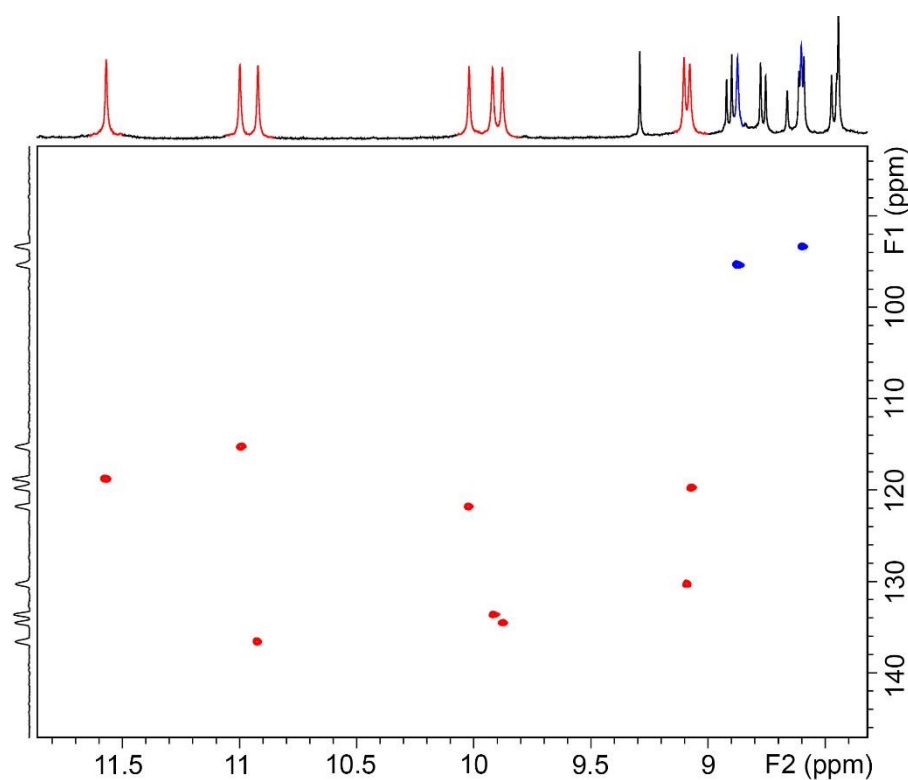


Figure S17. ^1H - ^{15}N HSQC spectrum (400 MHz, CDCl_3) of **8** (1 mM) at 298K. The amide and amine $-\text{NH}$ signals are highlighted in red and blue colors, respectively.

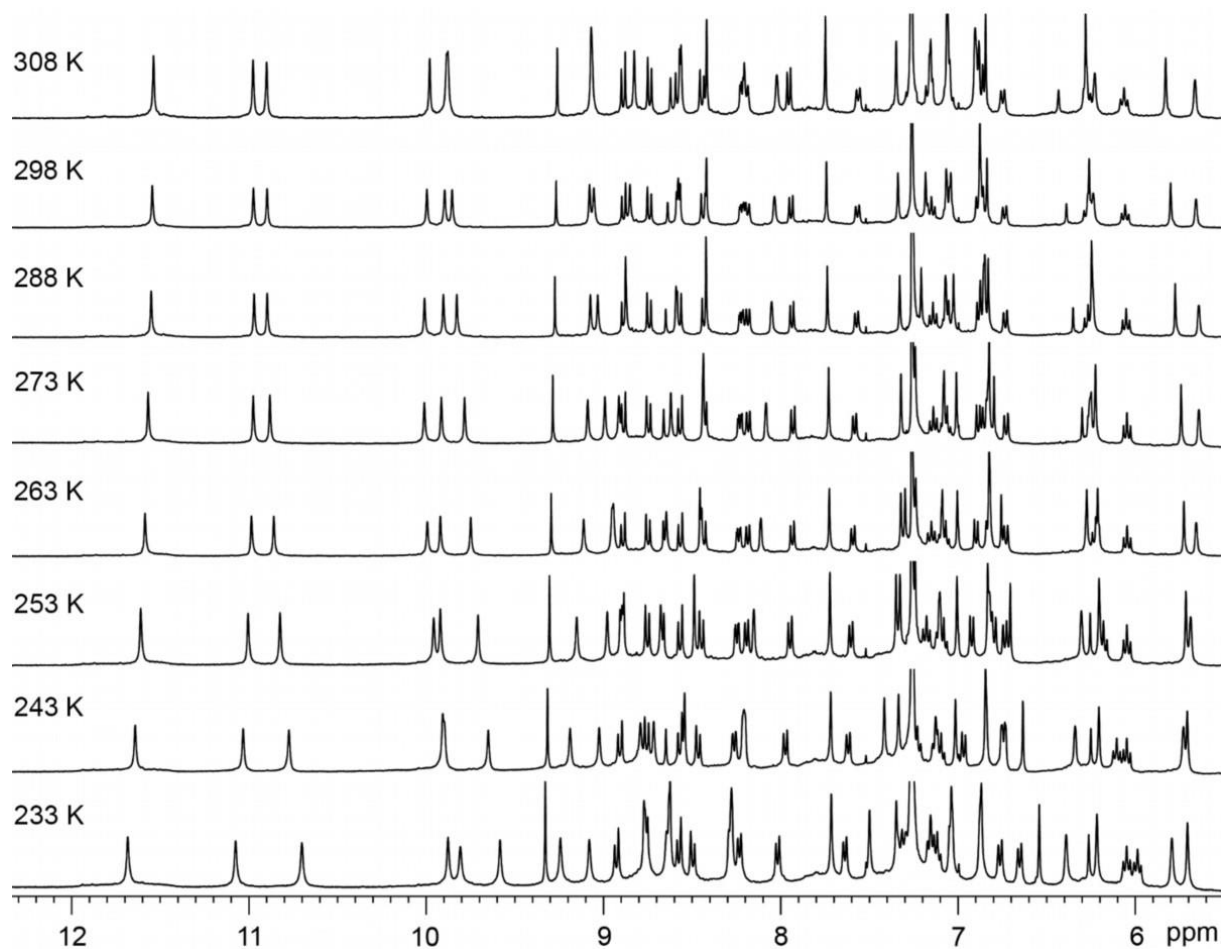


Figure S18. Part of the variable temperature ¹H NMR spectra (400 MHz, CDCl₃) of **8** (1 mM) from 308 K to 233 K.

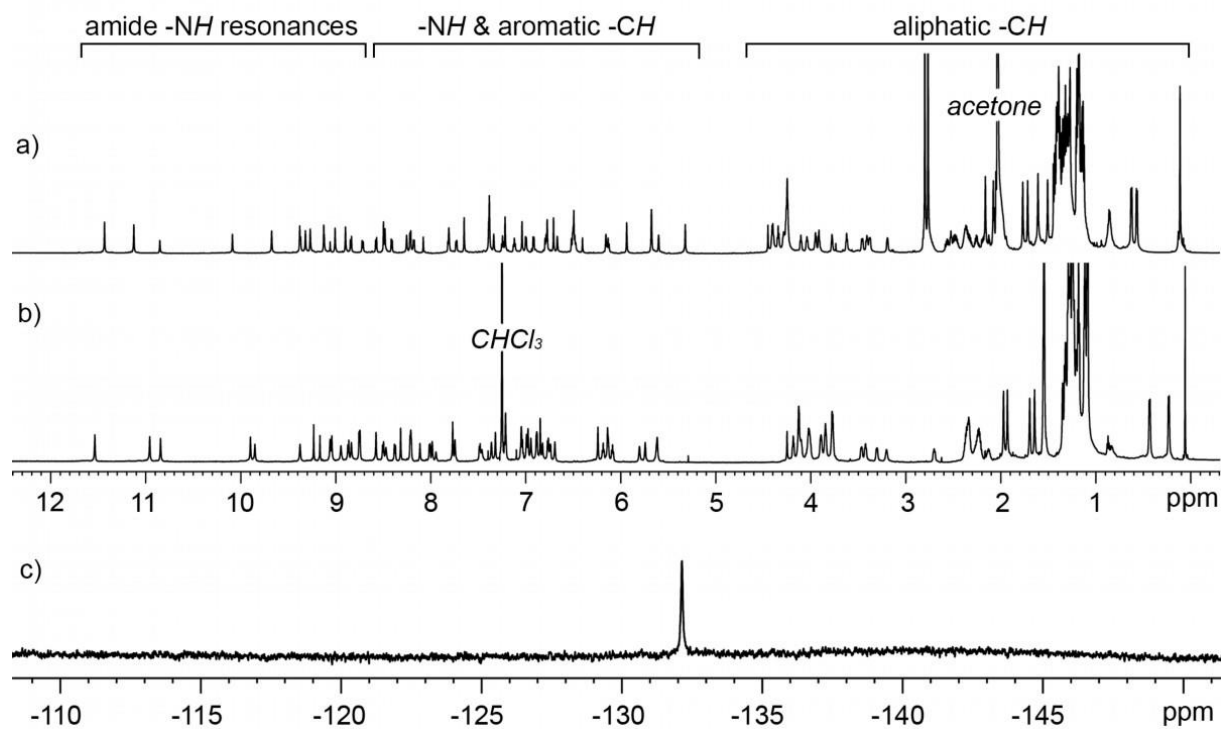


Figure S19. 700 MHz ¹H NMR spectra at 298 K of oligomer **12** in: (a) acetone-*d*₆; and (b) CDCl₃. (c) 376 MHz ¹⁹F NMR spectra at 298 K of oligomer **12** in CDCl₃.

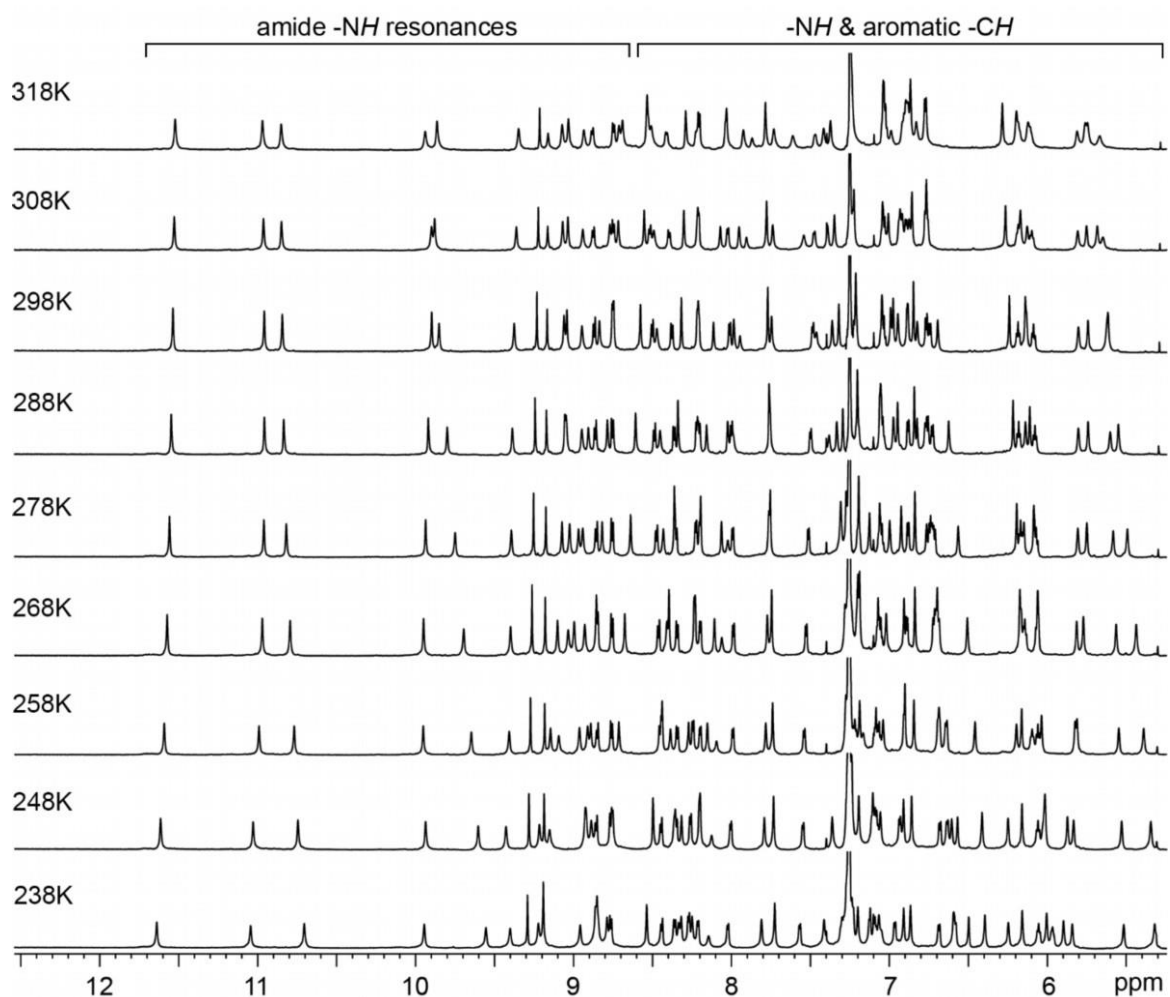


Figure S20. Part of the variable temperature ^1H NMR spectra (700 MHz, CDCl_3) of **12** (1 mM) from 318K to 238K.

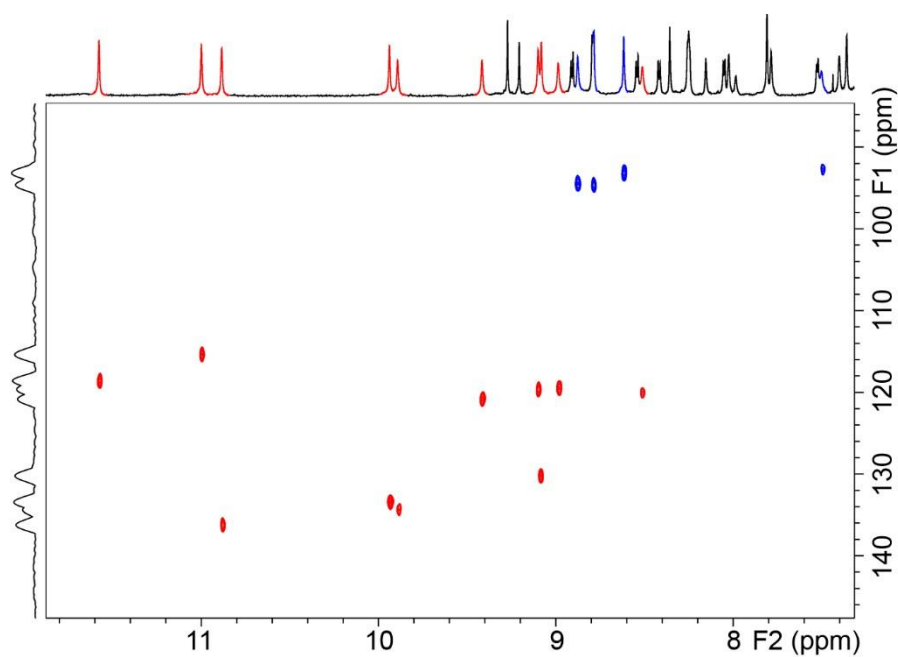


Figure S21. ^1H - ^{15}N HSQC spectrum (400 MHz, CDCl_3) of **12** (1 mM) at 298K. The -NH amide signals are depicted in red and turn unit amine protons are shown in blue colors.

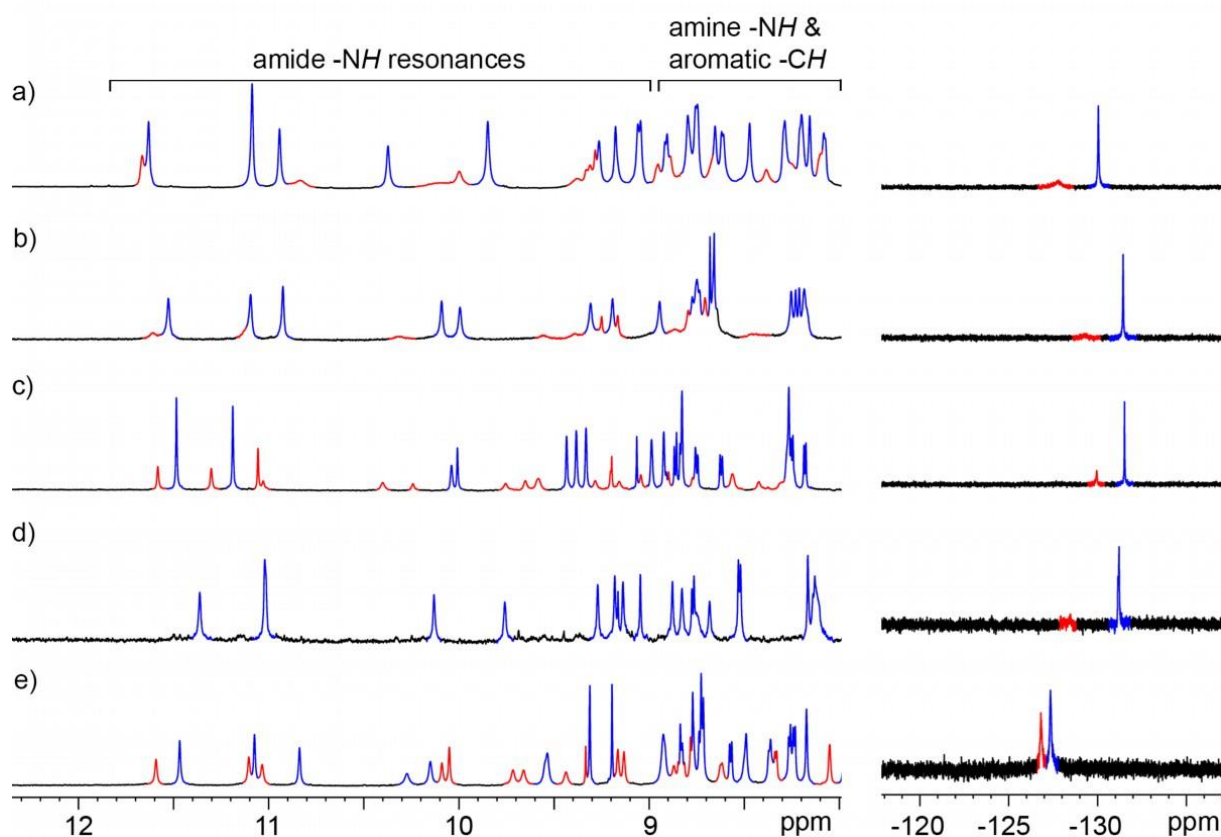


Figure S22. Part of 700 MHz ^1H NMR spectra at 298 K of oligomer **9** in: (a) CDCl_3 , (b) CD_2Cl_2 , (c) acetone- d_6 , (d) CD_3CN , and (e) d_2 -tetrachloroethane. The corresponding ^{19}F NMR in respective solvents are given in right side. Existence of two conformers has been represented by highlighting signals corresponding to those by red and blue respectively.

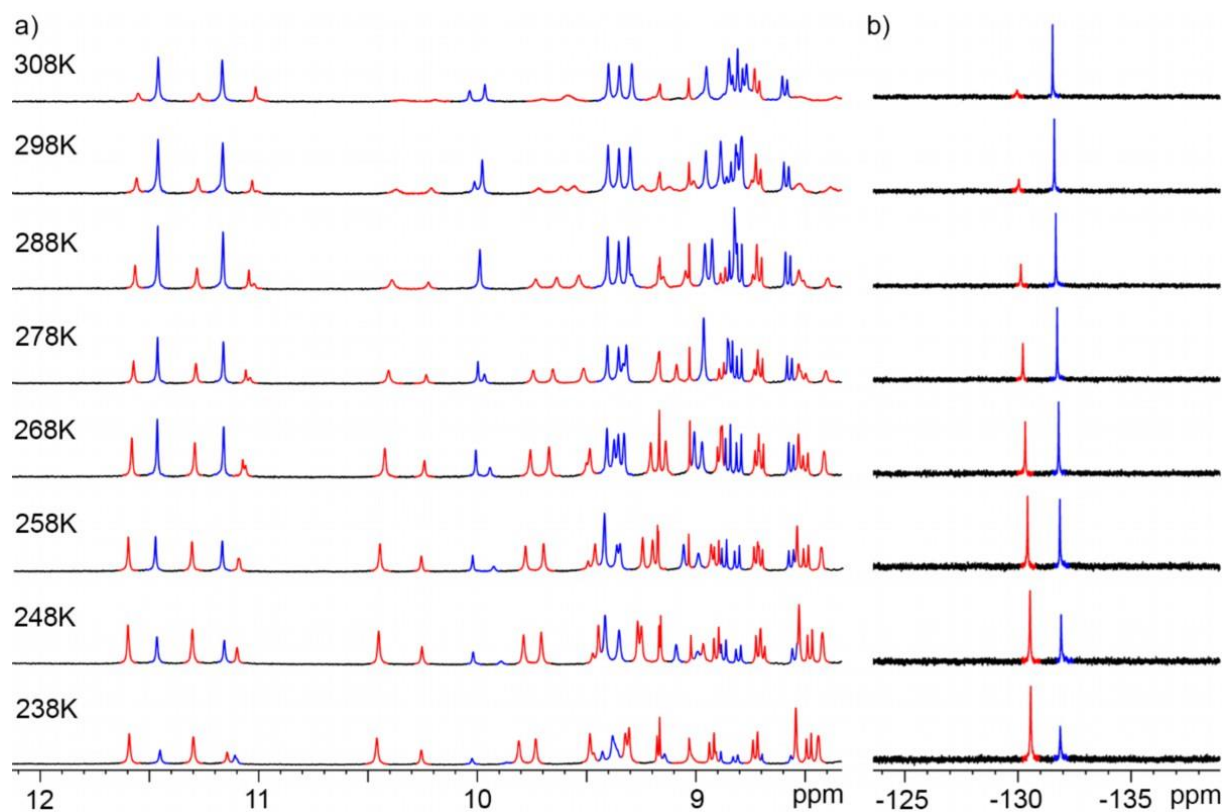


Figure S23. Part of the variable temperature (a) ¹H NMR spectra (400 MHz) and (b) ¹⁹F-NMR in acetone-*d*₆ of **9** (1 mM) from 308 K to 238 K. Two conformers present at room temperature are in slow exchange at the NMR time scale are highlighted in red and blue, respectively.

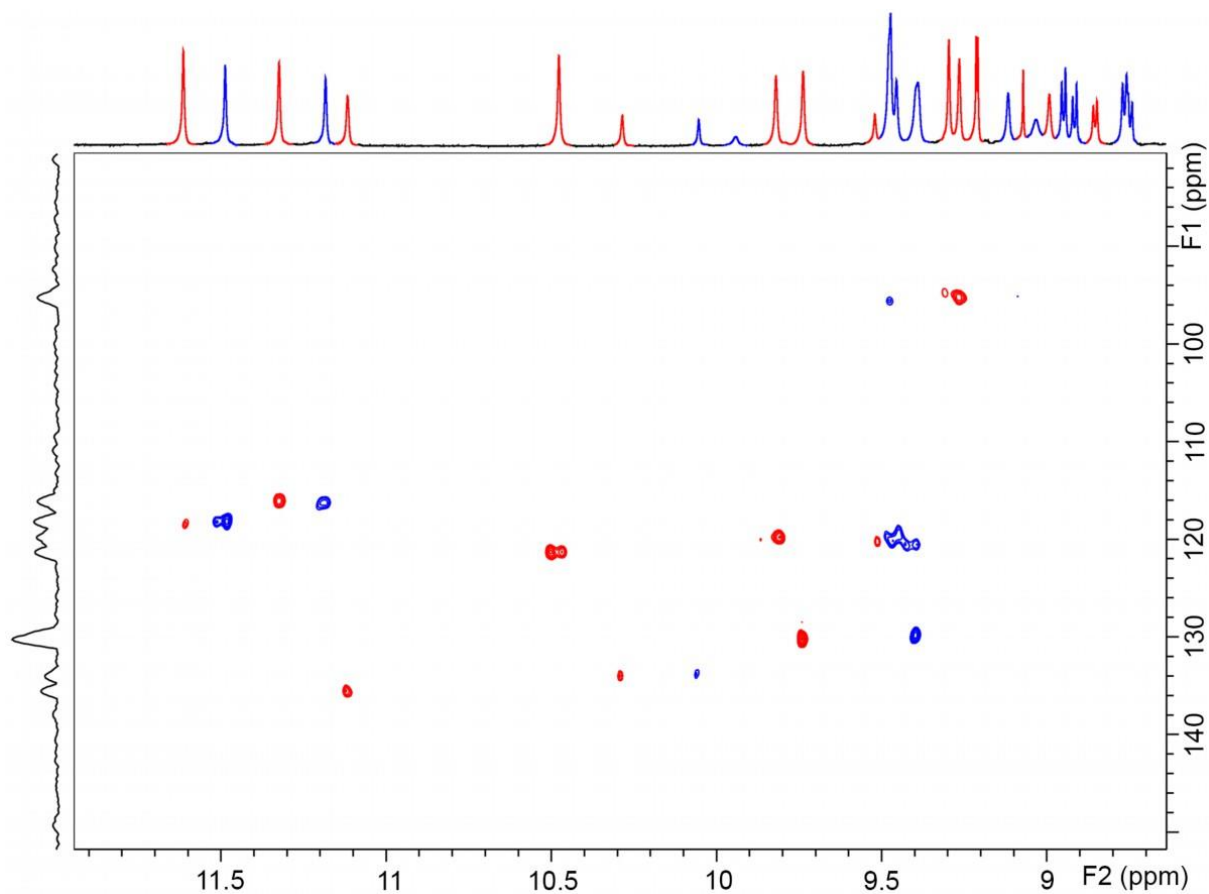


Figure S24. ^1H - ^{15}N HSQC spectrum (700 MHz, acetone- d_6) of **9** (1 mM) at 258K. Signals corresponding to two conformers are shown in red and blue colors respectively.

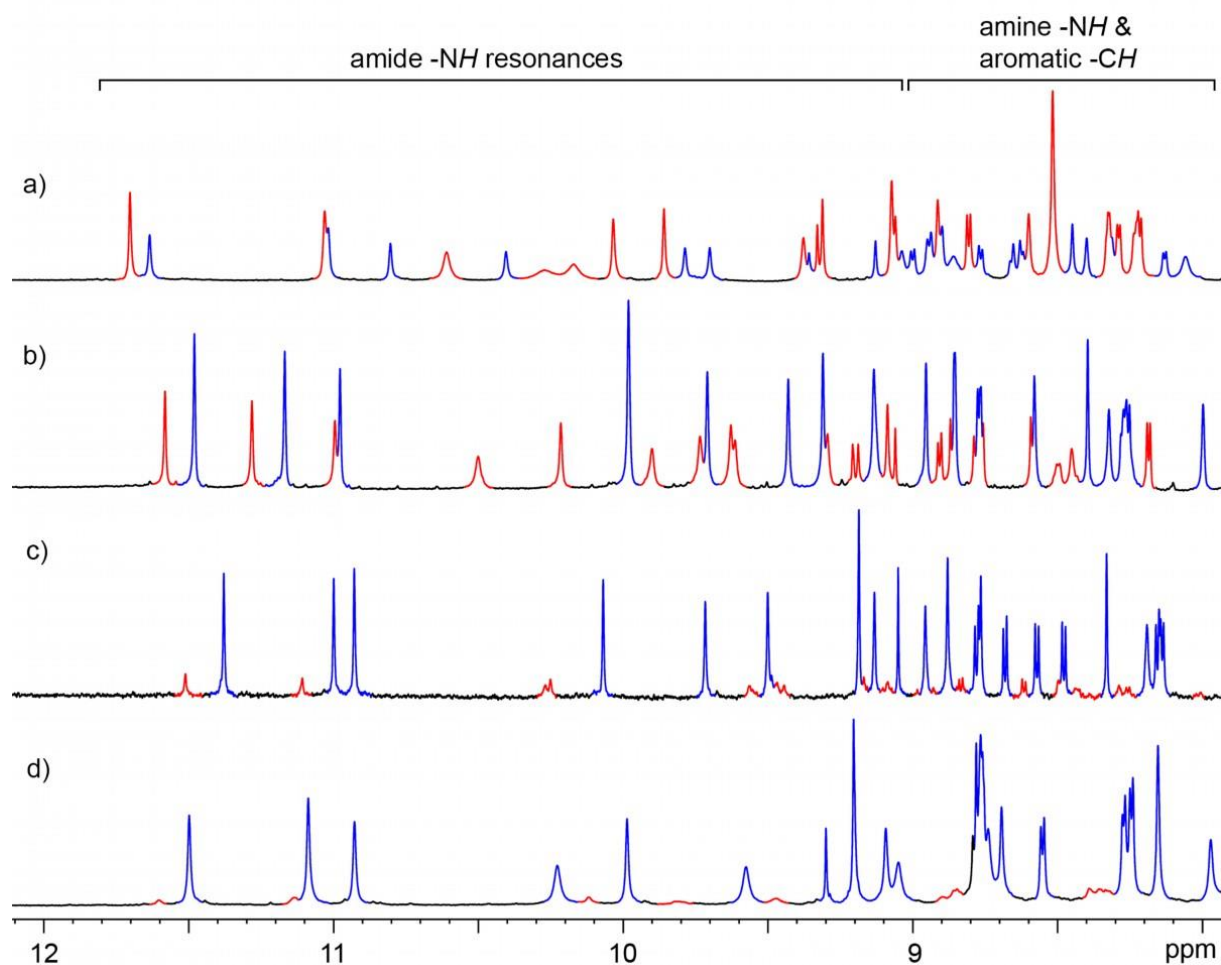


Figure S25. Part of 700 MHz ^1H NMR spectra at 298 K of oligomer **10** (1mM) in: (a) CDCl_3 , (b) d_6 -acetone, (c) CD_3CN , and (d) d_2 -tetrachloroethane. Two conformers present at room temperature are in slow exchange at the NMR time scale are highlighted in red and blue, respectively.

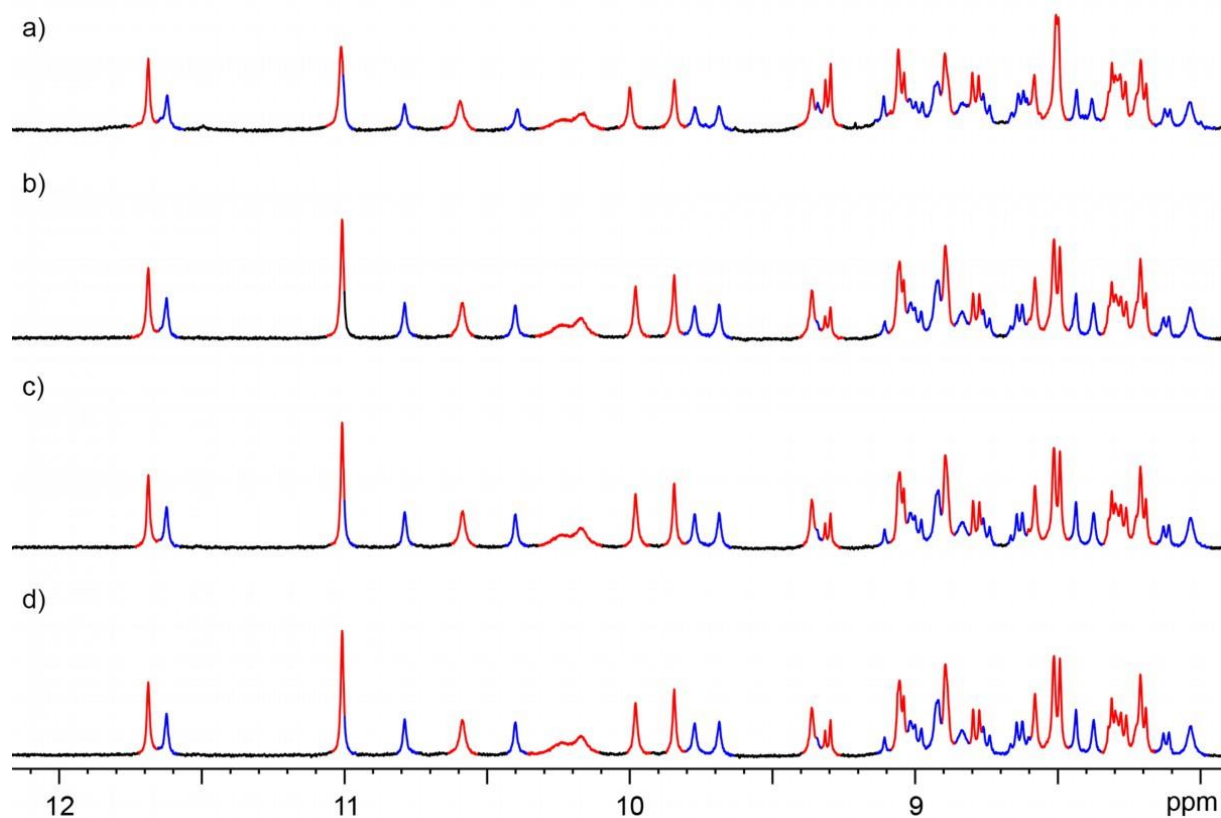


Figure S26. Part of 400 MHz ¹H NMR spectra at 298 K of oligomer **10**; a) in CDCl₃, (b) Freshly dissolved crystals after 5 min, (c) after 25 min and (d) 50 min. Signals corresponds to two conformers are highlighted in red and blue, respectively. No difference in their proportion after freshly dissolution of crystals compared to reference NMR as well as no change of it over time indicating significantly fast dynamics among the conformers.

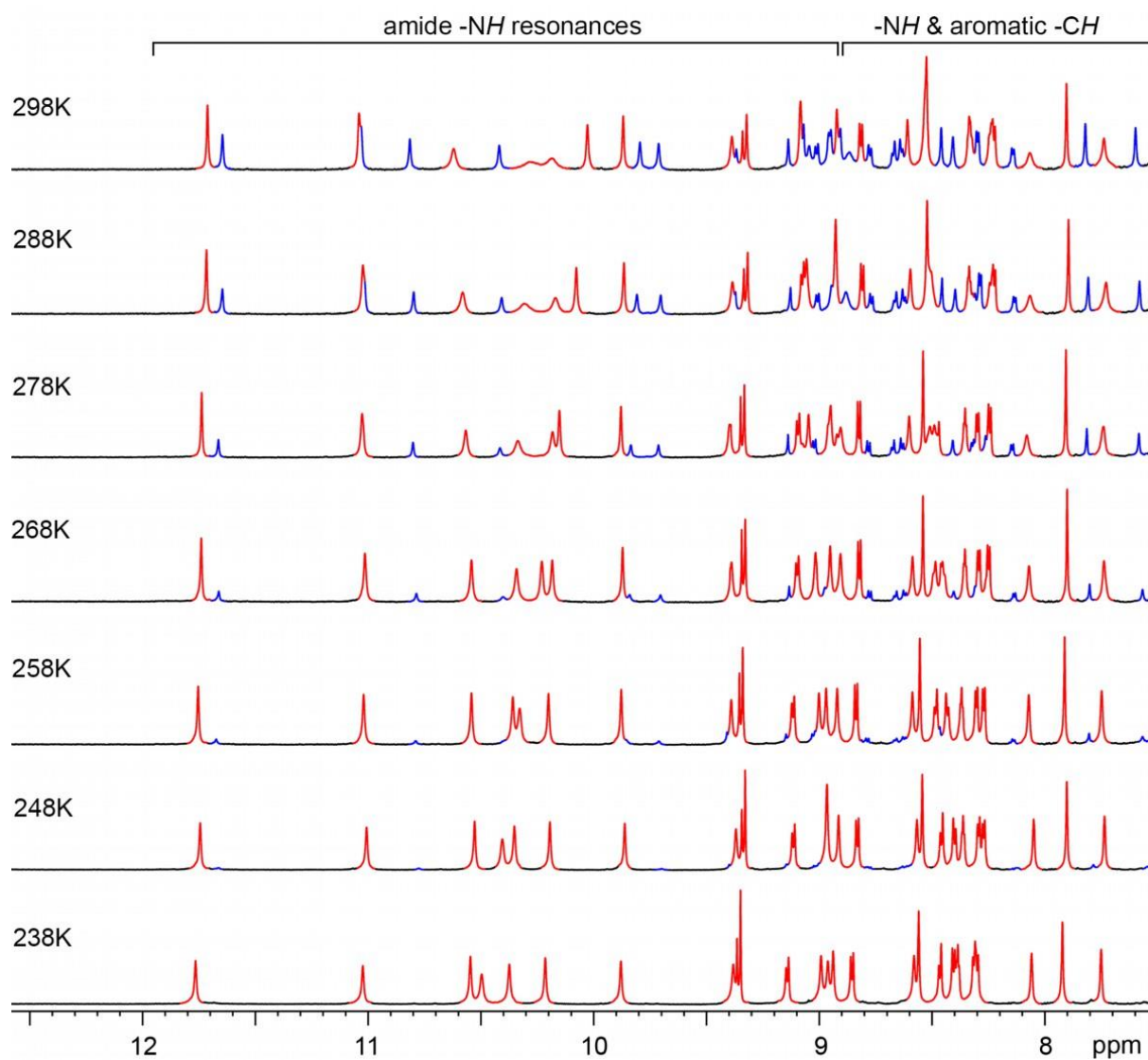


Figure S27. Part of the variable temperature ^1H NMR spectra (700 MHz, CDCl_3) of **10** (1 mM) from 298 K to 238 K. Two conformers present at room temperature are in slow exchange at the NMR time scale are highlighted in red and blue peaks, respectively. One of the conformers highlighted in blue disappear at 238 K.

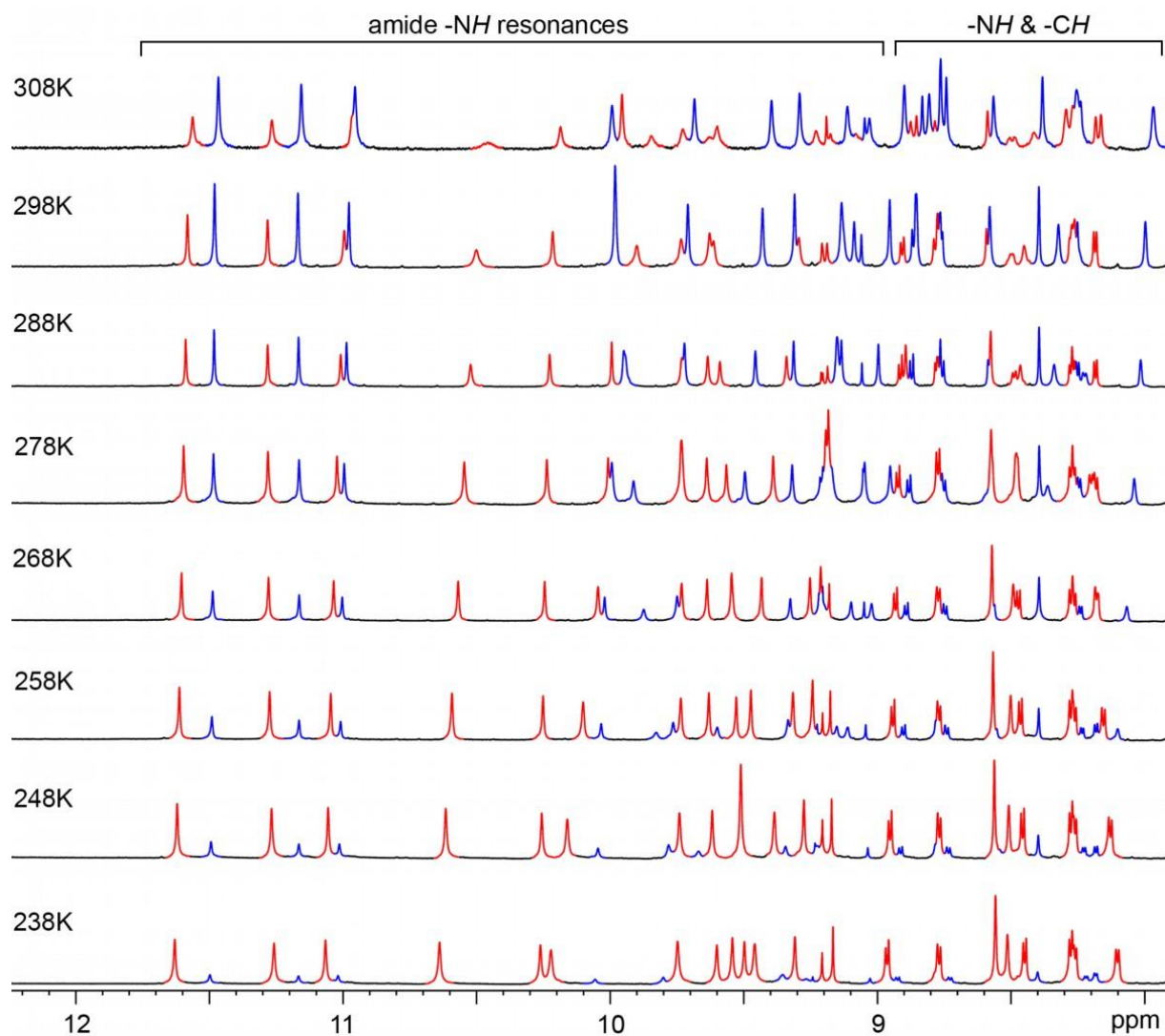


Figure S28. Part of the variable temperature ¹H NMR spectra (700 MHz, acetone-*d*₆) of **10** (1 mM) from 308K to 238K. The peaks highlighted in red and blue are corresponding to two conformers present at room temperature. Those are in slow exchange at the NMR time scale. Proportion of two conformers changes with temperature variation.

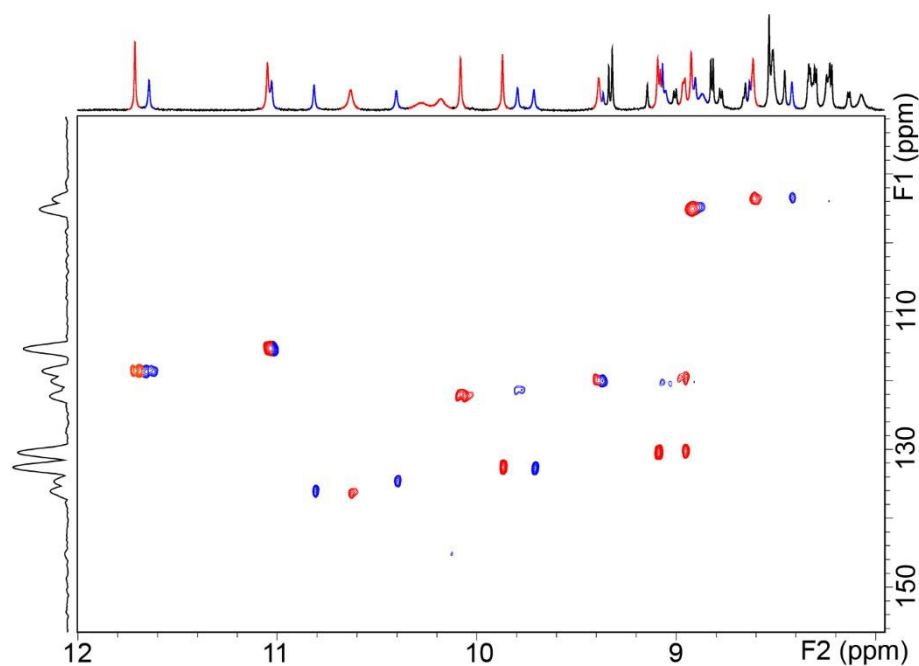


Figure S29. ^1H - ^{15}N HSQC spectrum (700 MHz, CDCl_3) of **10** (1 mM) at 298K. The -NH signals present in two conformers are shown in red and blue colors. Two NH correlations could be detected due to resonance broadness.

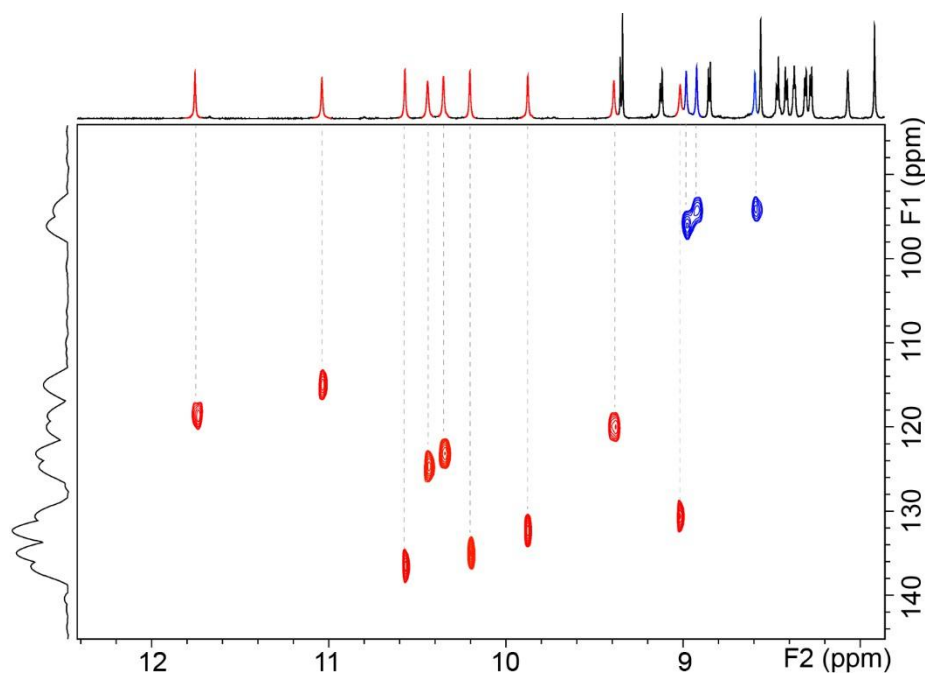


Figure S30. ^1H - ^{15}N HSQC spectrum (700 MHz, CDCl_3) of **10** (1 mM) at 248K. At this temperature only one conformer was observed. The NH amide signals are depicted in red and turn unit amine protons are shown in blue colors.

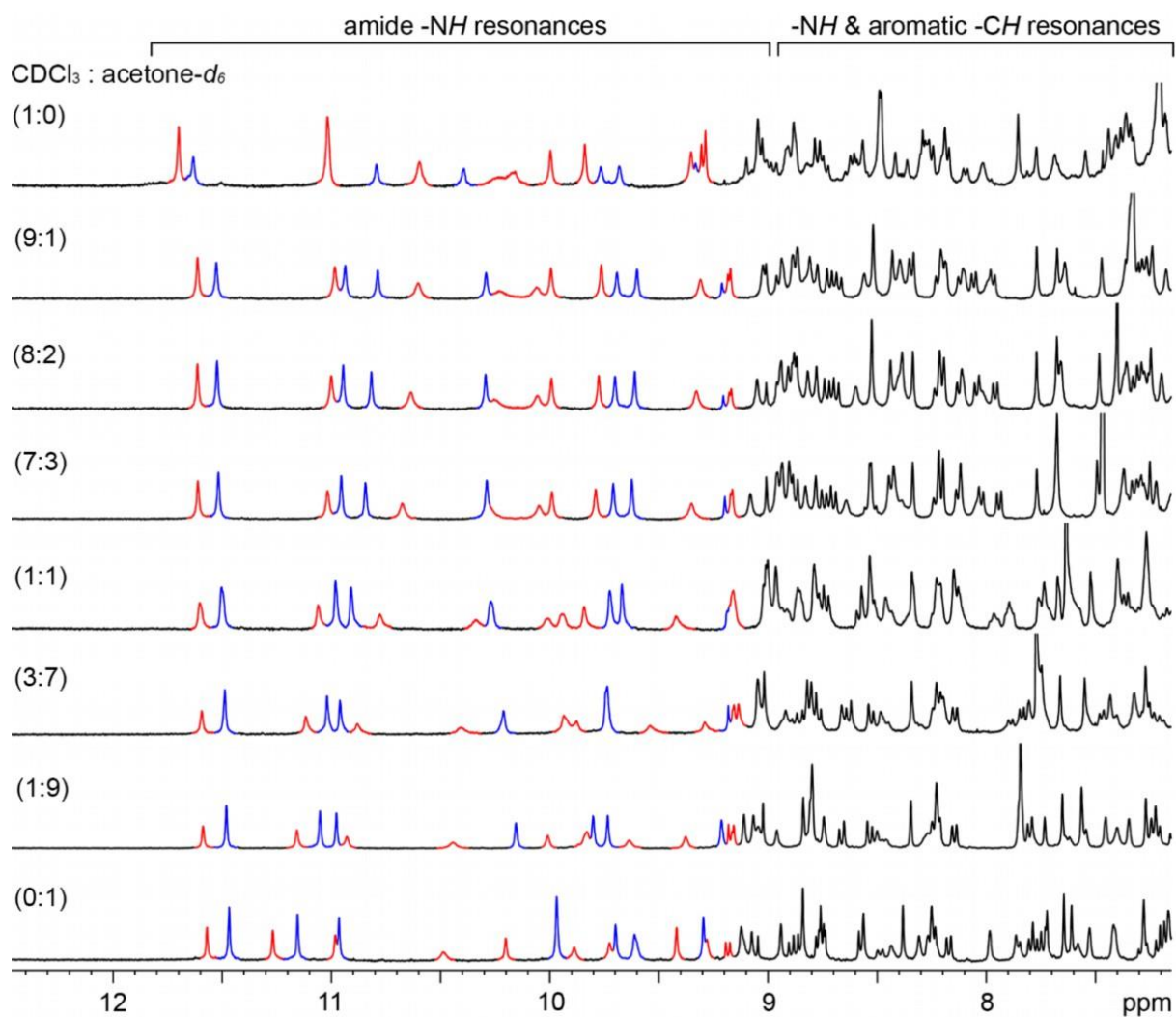


Figure S31. Part of ¹H NMR spectra (400 MHz) of **10** (1 mM) in varying CDCl₃/acetone-*d*₆ mixtures at 298K. Proportion of the two conformers, highlighted with red and blue peaks, can be seen changes from CDCl₃ to acetone-*d*₆.

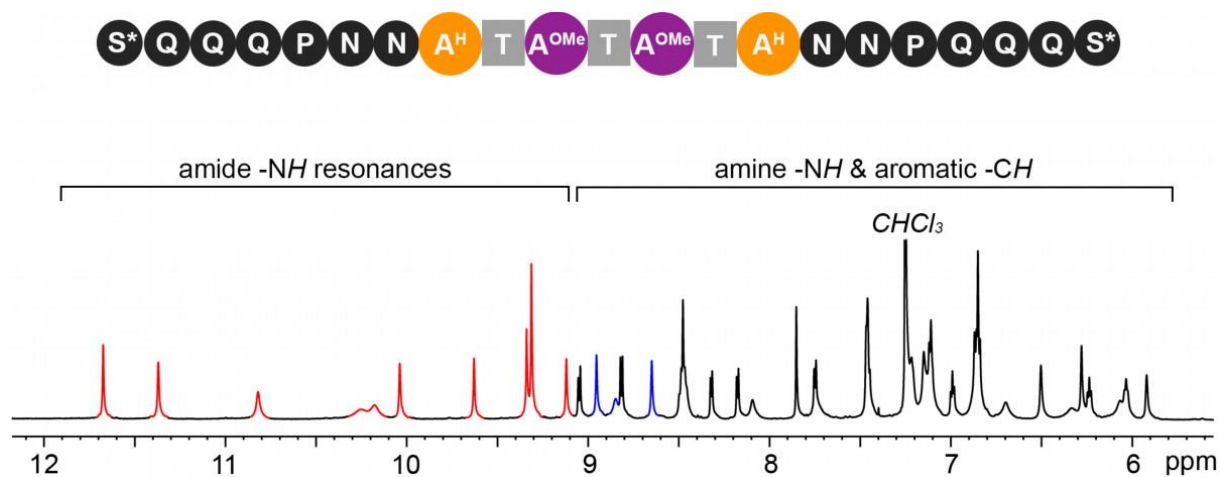


Figure S32. Part of 700 MHz ^1H NMR spectra at 298 K of oligomer **11** (1 mM) in CDCl_3 , showing presence of single conformer. Peaks highlighted in red and blue corresponds to the amide and amine proton signals respectively.

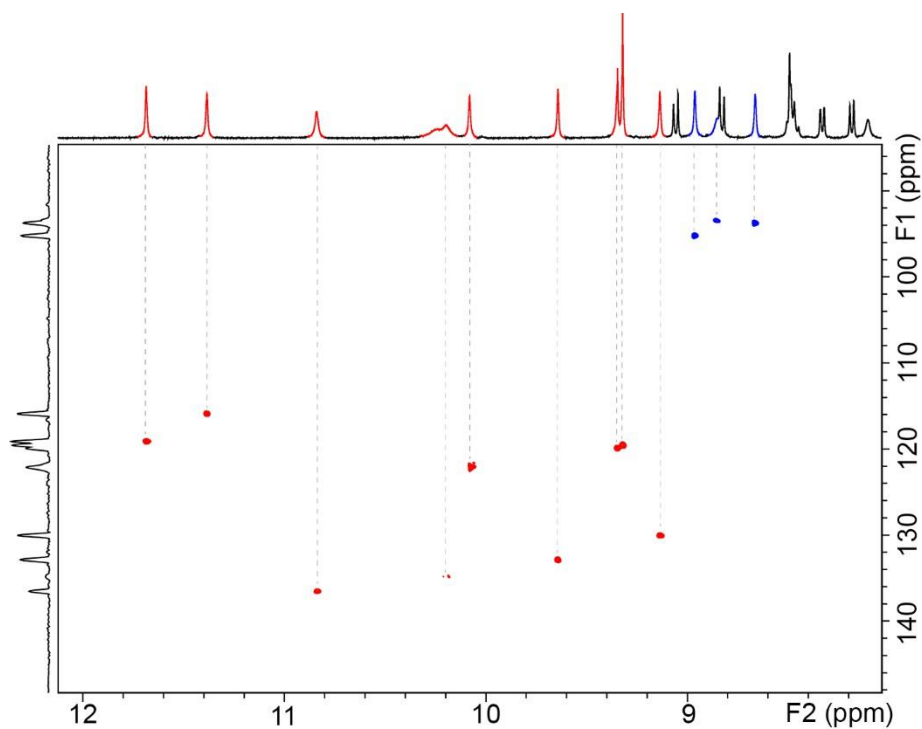


Figure S33. ^1H - ^{15}N HSQC spectrum (700 MHz, CDCl_3) of **11** (1 mM) at 298K. The NH amide signals are depicted in red and turn unit amine protons are shown in blue colors.

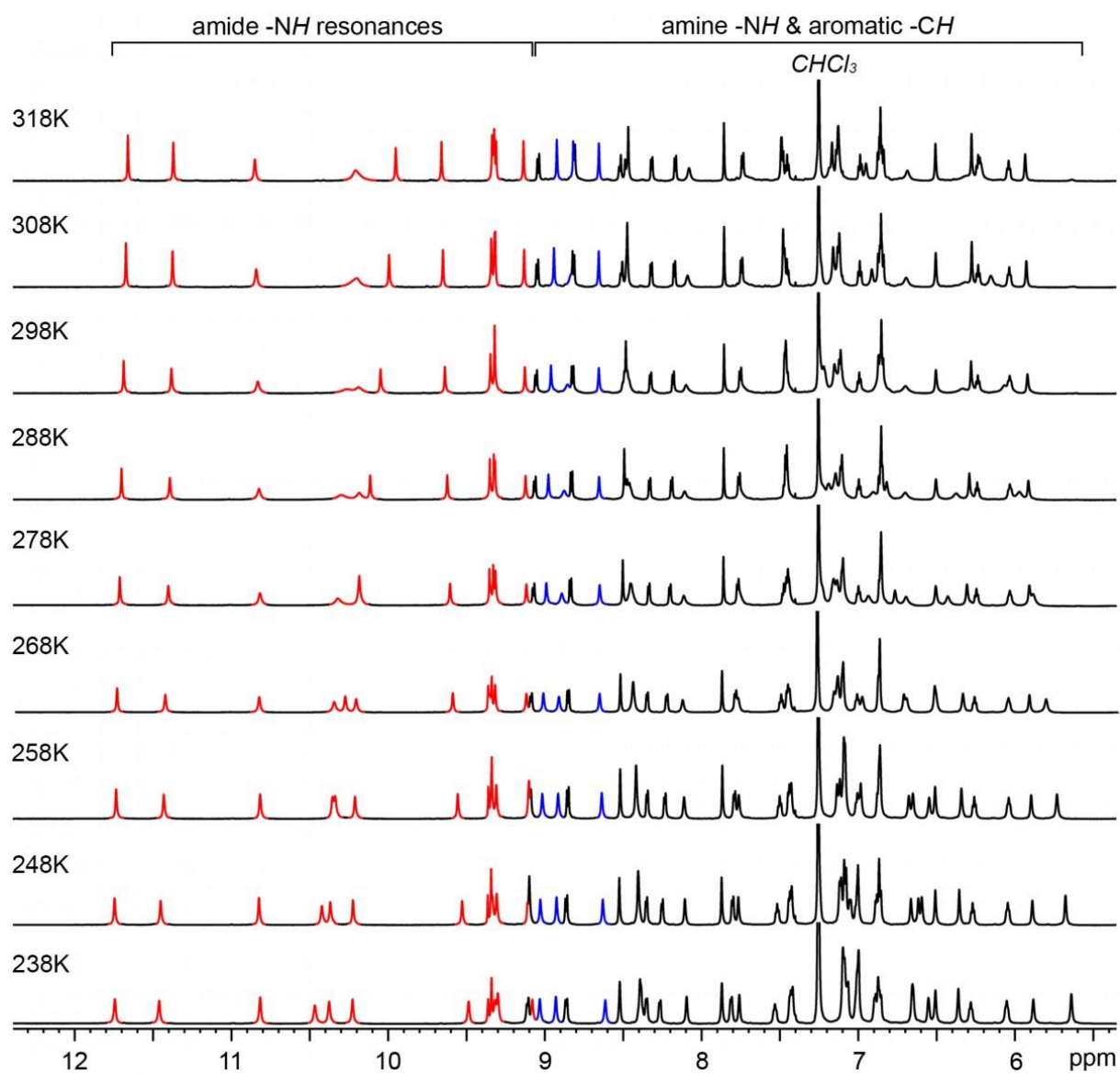


Figure S34. Part of the variable temperature ^1H NMR spectra (700 MHz, CDCl_3) of **11** (1 mM) from 318K to 238K. Peaks highlighted in red and blue corresponds to the amide and amine proton signals respectively.

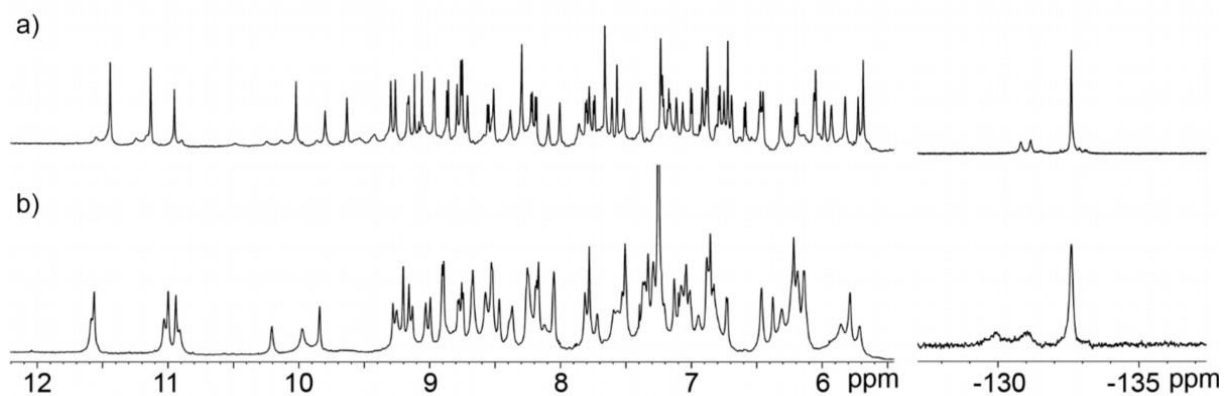


Figure S35. Part of 700 MHz ¹H NMR spectra at 298 K of oligomer **13** (1mM) in: (a) acetone-*d*₆, (b) CDCl₃ and corresponding ¹⁹F-NMR in respective solvents are given at right side.

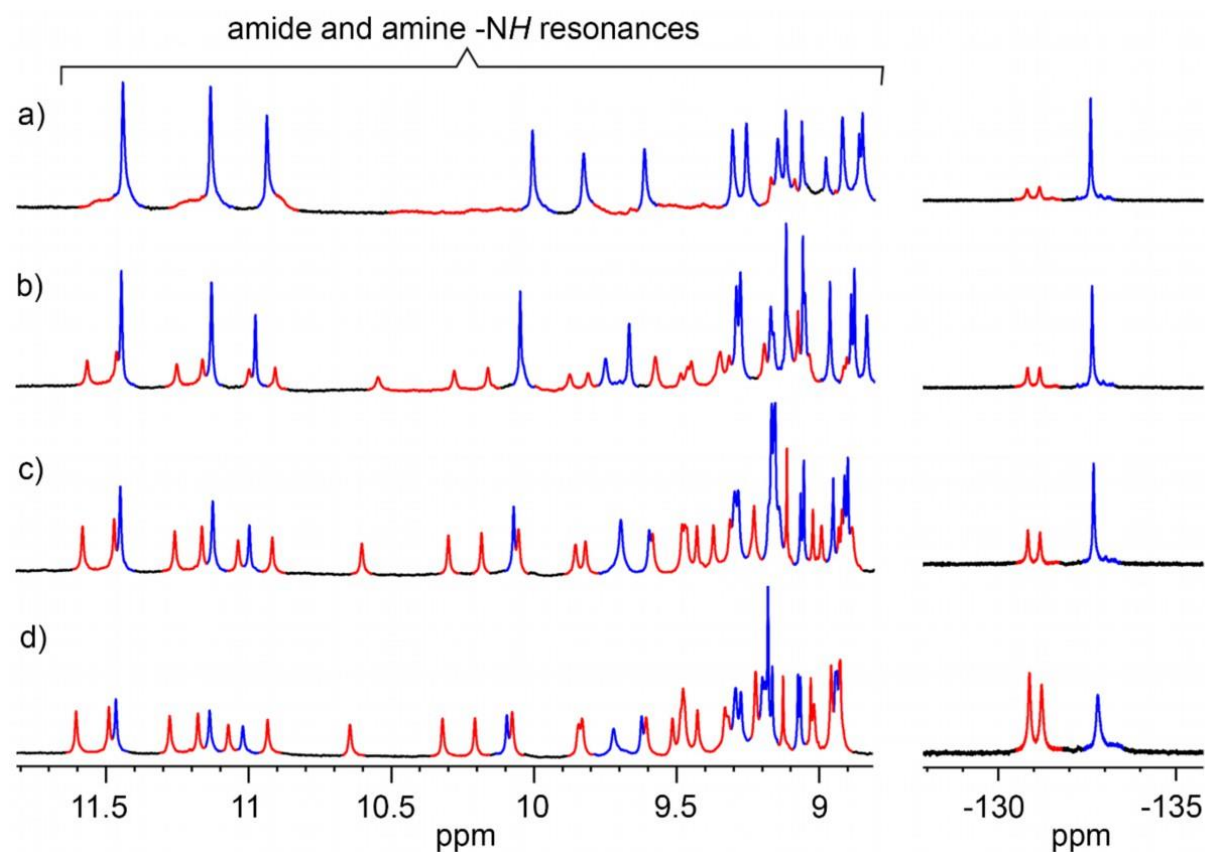


Figure S36. The 400 MHz ¹H-NMR (left) and side by side 376 MHz ¹⁹F NMR (right) spectra of **13** (1 mm) at: a) 298 K and; b) 278 K; c) 258 K; and d) 248 K in acetone-*d*₆. Signals assigned to two different conformers **13-sym** and **13-asm** are highlighted in blue and red, respectively.

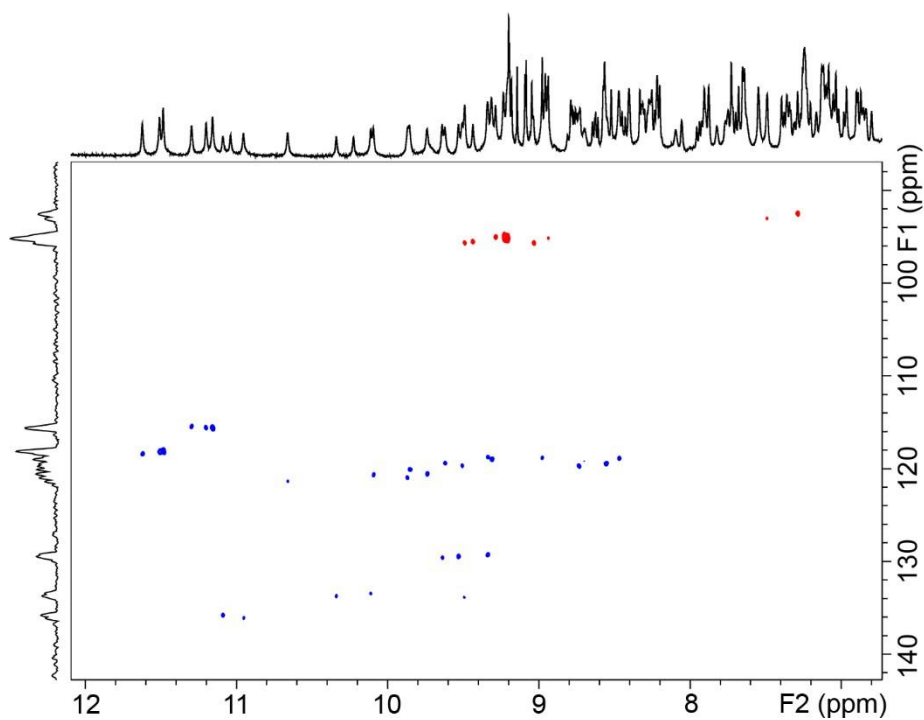


Figure S37. ^1H - ^{15}N HSQC spectrum (700 MHz, acetone- d_6) of **13** (1 mM) at 258K. The -NH amide signals are depicted in blue and turn unit amine protons are shown in red colors.

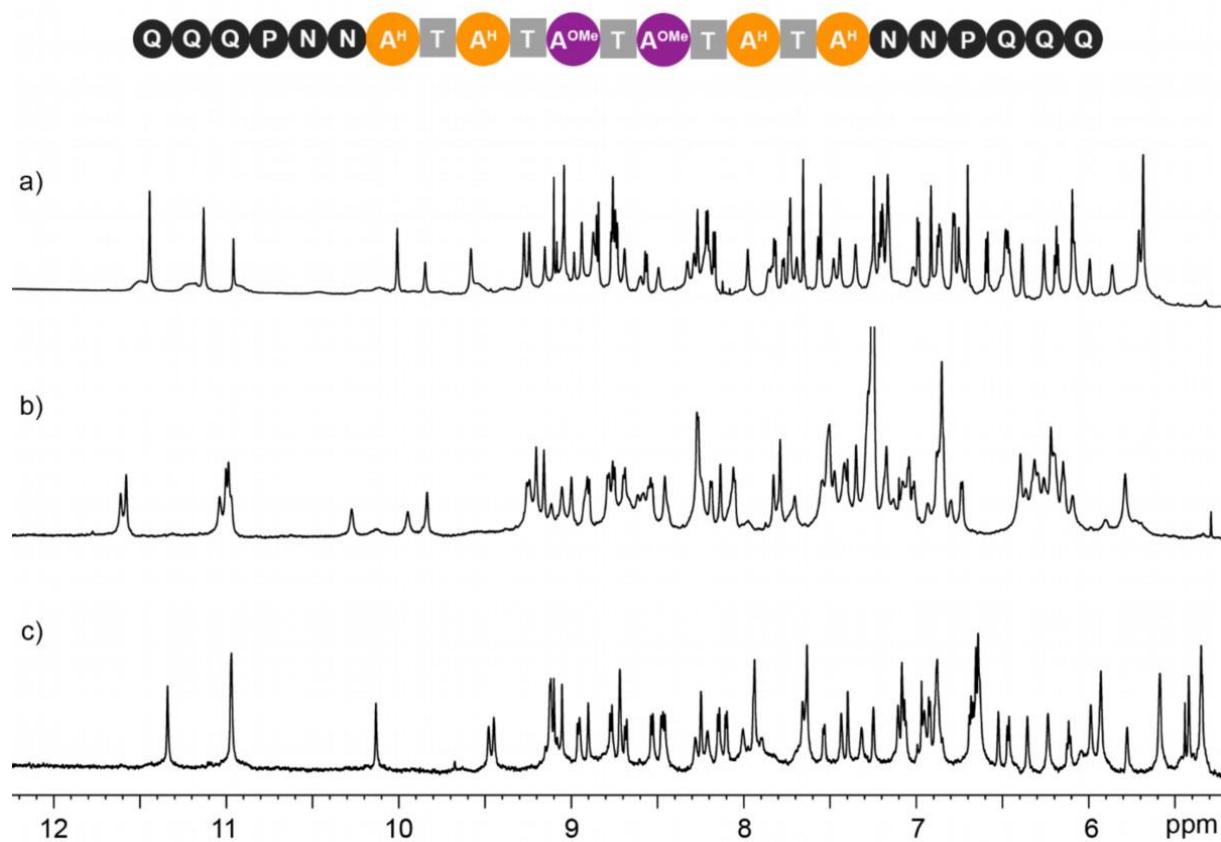


Figure S38. Part of 700 MHz ^1H NMR spectra at 298 K of oligomer **14** (1mM) in: (a) acetone- d_6 , (b) CDCl_3 , and (c) CD_3CN .

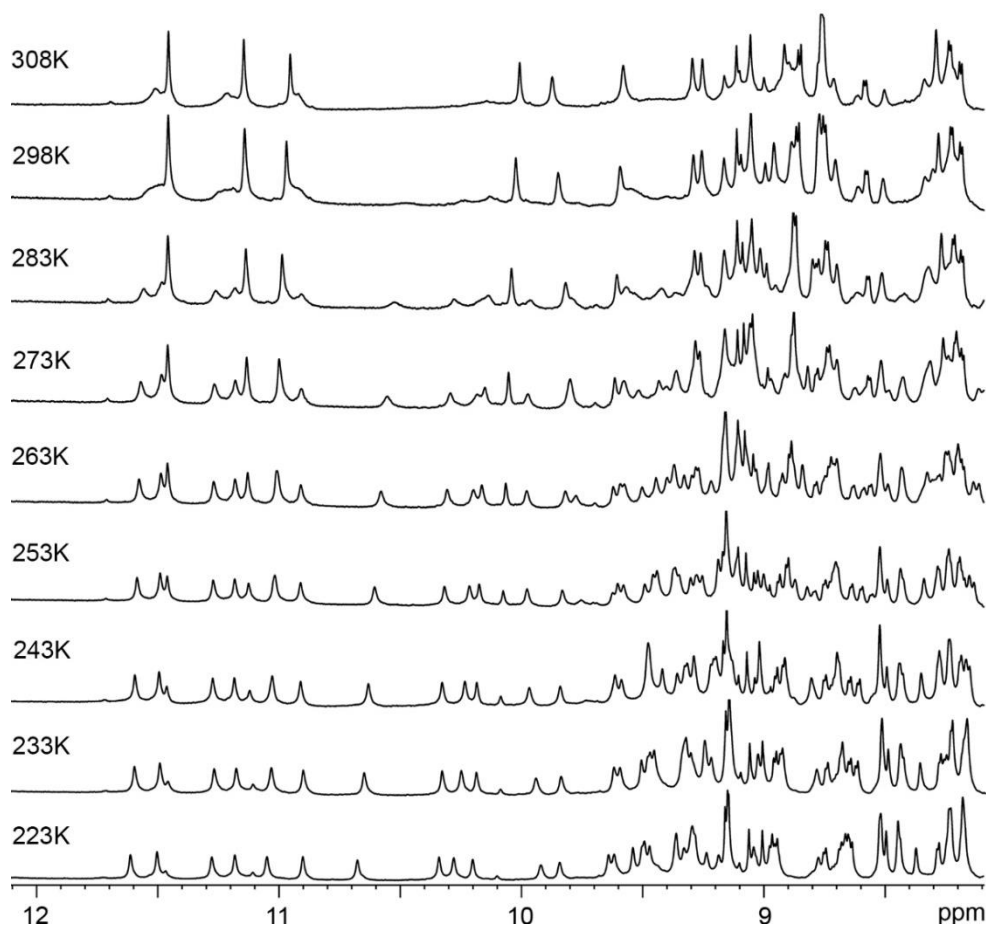


Figure S39. Part of the variable temperature ^1H NMR spectra (700 MHz, acetone- d_6) of **14** (1 mM) from 308K to 223K.

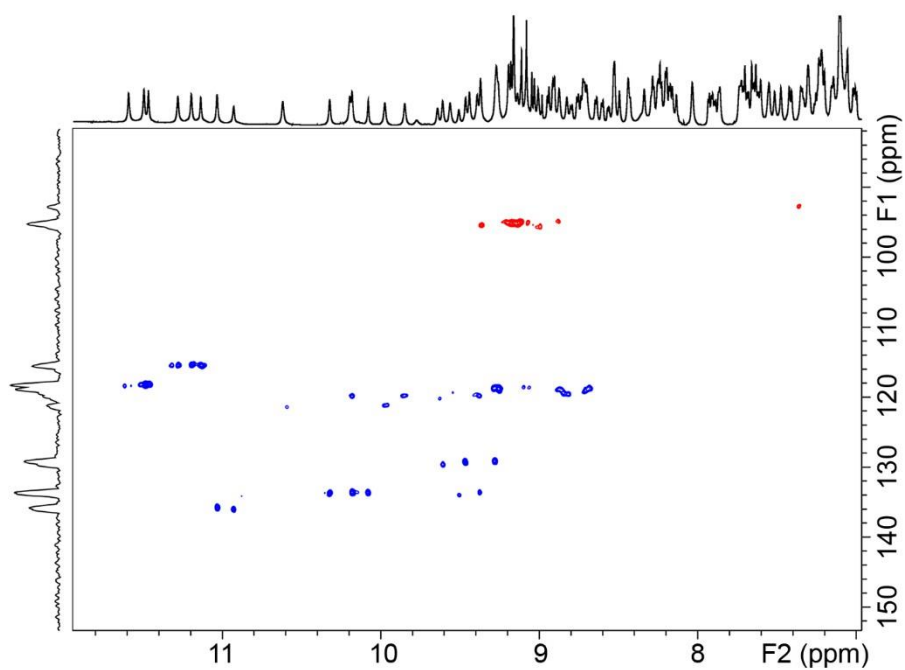


Figure S40. ^1H - ^{15}N HSQC spectrum (700 MHz, acetone- d_6) of **14** (1 mM) at 258K. The NH amide signals are depicted in blue and turn unit amine protons are shown in red colors.

7. Photoirradiation of oligomers

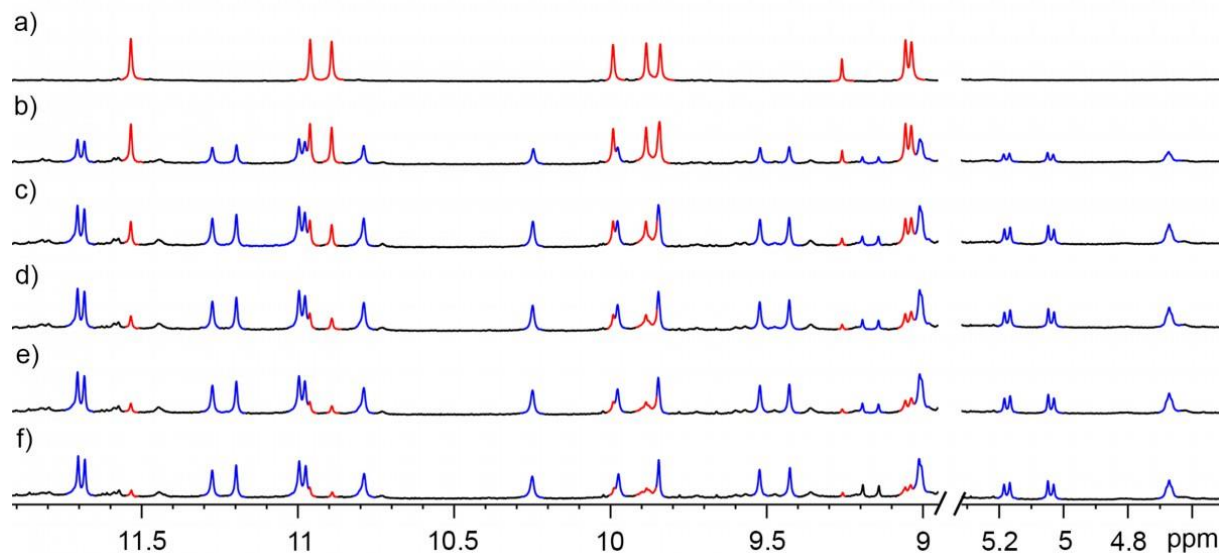
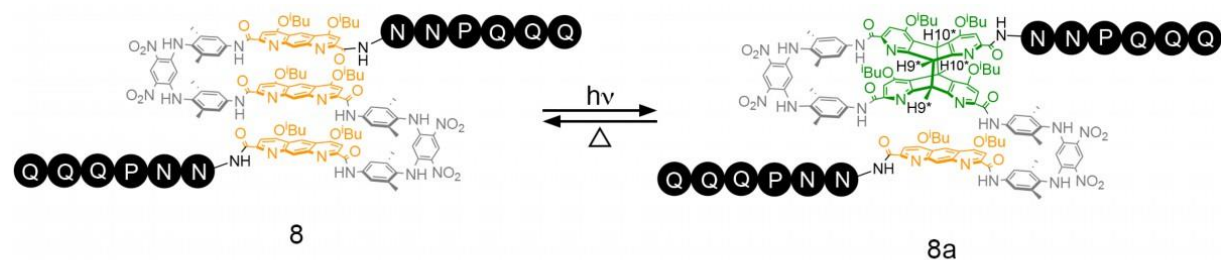


Figure S41. Part of the 700 MHz ^1H NMR spectra of **8** at 298K (1 mM in acetone- d_6) under photoirradiation after (a) 0 min; (b) 15 min.; (c) 30 min; (d) 45 min; (e) 60 min; and (f) 90 min. The signals correspond to the starting material are shown in red and for photoproducts, **8a**, in blue. The presence of negligible amount of non identified byproduct cannot be assigned.

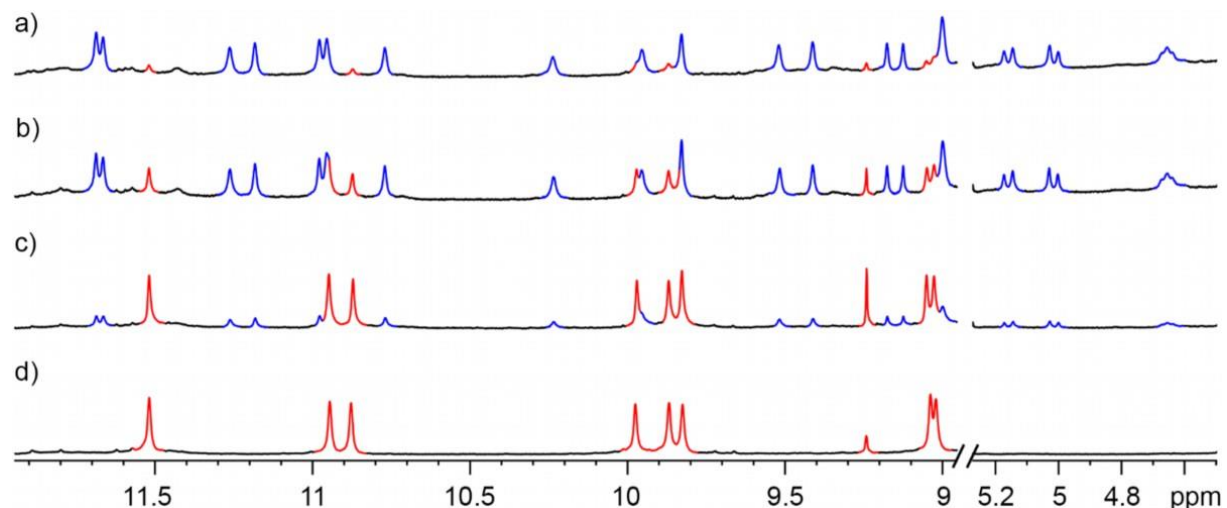


Figure S42. Part of the 400 MHz ^1H NMR spectra of **8a** at 298K after (a) 0 min; (b) 3 h; (c) 15 h; and (d) 36 h of heating in CDCl_3 at 333 K. The signals correspond to the **8a** and **8** are shown in blue and red respectively.

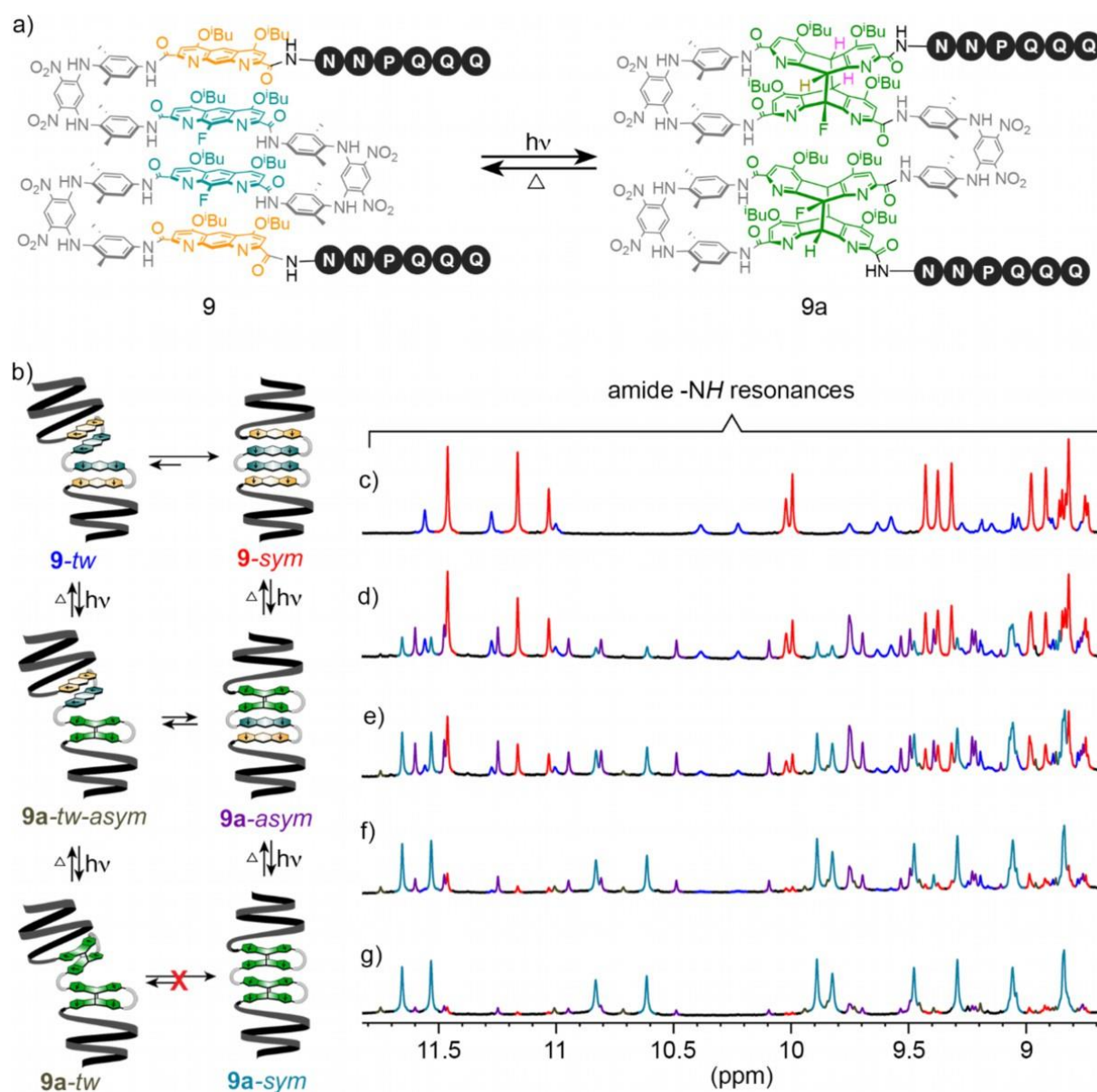


Figure S43. a) Photoproduct formation of **9** and thermal reversibility. b) Schematic illustration of stepwise photoadduct formation of different conformers of **9**. Initially two conformers, **9-*tw*** and **9-*sym***, are present in an equilibrium at 298 K. Both have equal possibility to undergo stepwise photoadduct formation to final photoproduct **9a-*tw*** and **9a-*sym*** through intermediate monoadducts **9a-*tw-asym*** and **9a-*asym***, respectively. Part of the 700 MHz ^1H NMR spectra of at 298K of **9** (1 mM in acetone- d_6) under photoirradiation after (b) 0 min; (c) 30 min.; (d) 1 h.; (e) 2 h.; and (f) 5 h. The signals corresponding to the starting two conformers **9-*tw*** and **9-*sym*** are shown in blue and red respectively. Signals in the photoirradiation steps are assigned to individual species are in accordance with color as depicted in (a).

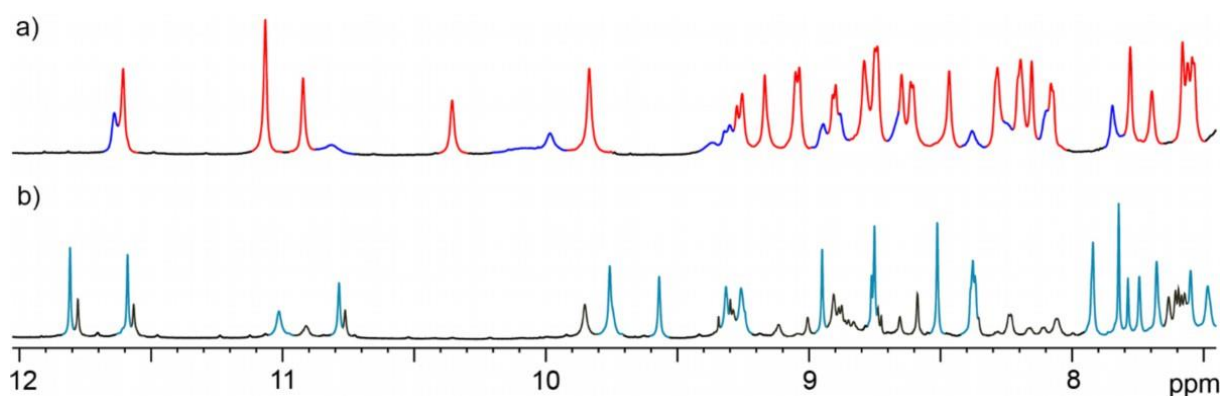


Figure S44. Part of the 700 MHz ^1H NMR spectra of at 298K of **9** (1 mM in CDCl_3) under photoirradiation. (a) before irradiation; and (b) after 5 h irradiation. The signals corresponding to the starting two conformers **9**-*tw* and **9**-*sym* are shown in blue and red respectively and signals corresponds to their respective photoproducts, **9a**-*tw* and **9a**-*sym*, are shown in grey and cyan.

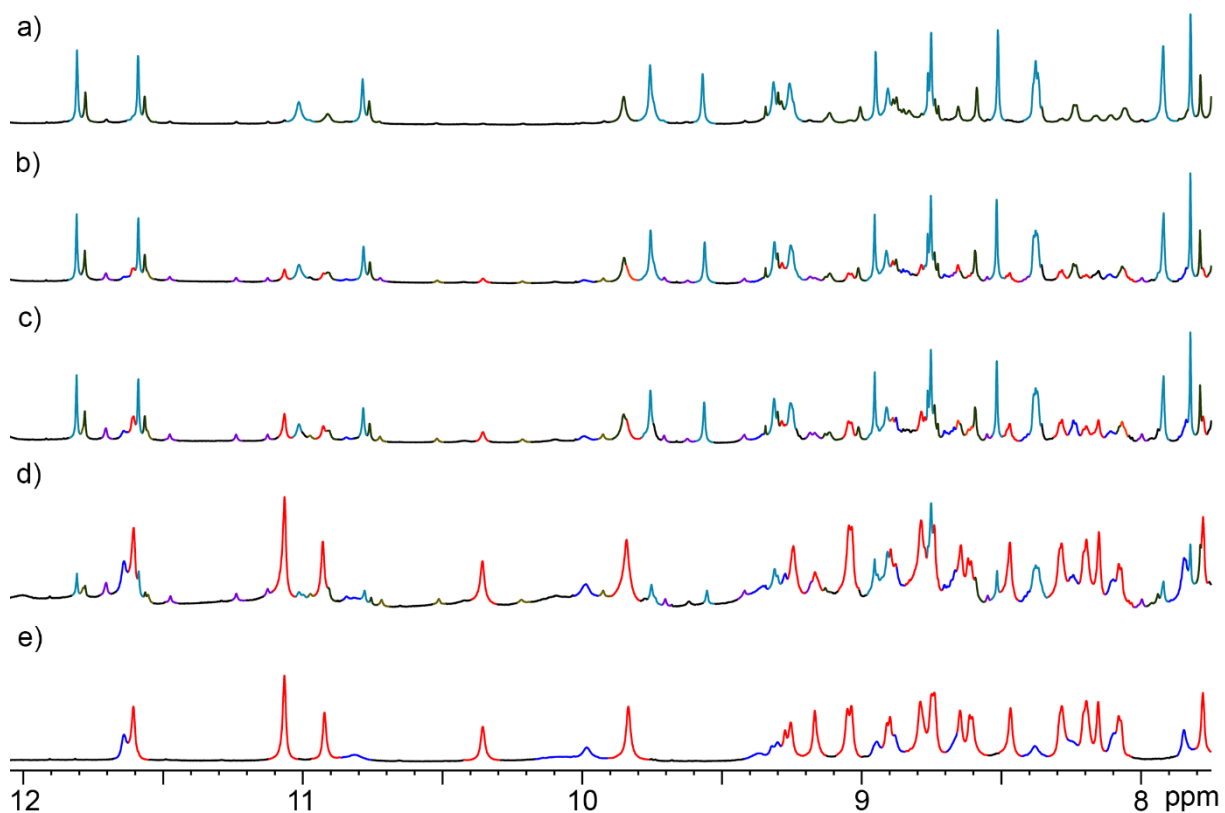


Figure S45. Thermal degradation of photoproduct **9a**. Part of the 400 MHz ^1H NMR spectra of **9a** at 298K (a) in CDCl_3 ; and after heating the solution for (b) 3 h; (c) 10 h; (d) 20 h; and (e) 48 h of heating in CDCl_3 at 333 K. The starting signals corresponding to two photoproduct conformers **9a**-*tw* and **9a**-*sym*, shown in brown and cyan respectively. After 48 h of heating at 333 K, the photoproducts are converted back to the initial products **9**-*tw* and **9**-*sym*. Their signals are shown in blue and red respectively. In between, intermediates are observed which are the monoadducts and their color code is maintained according to Figure S43b.

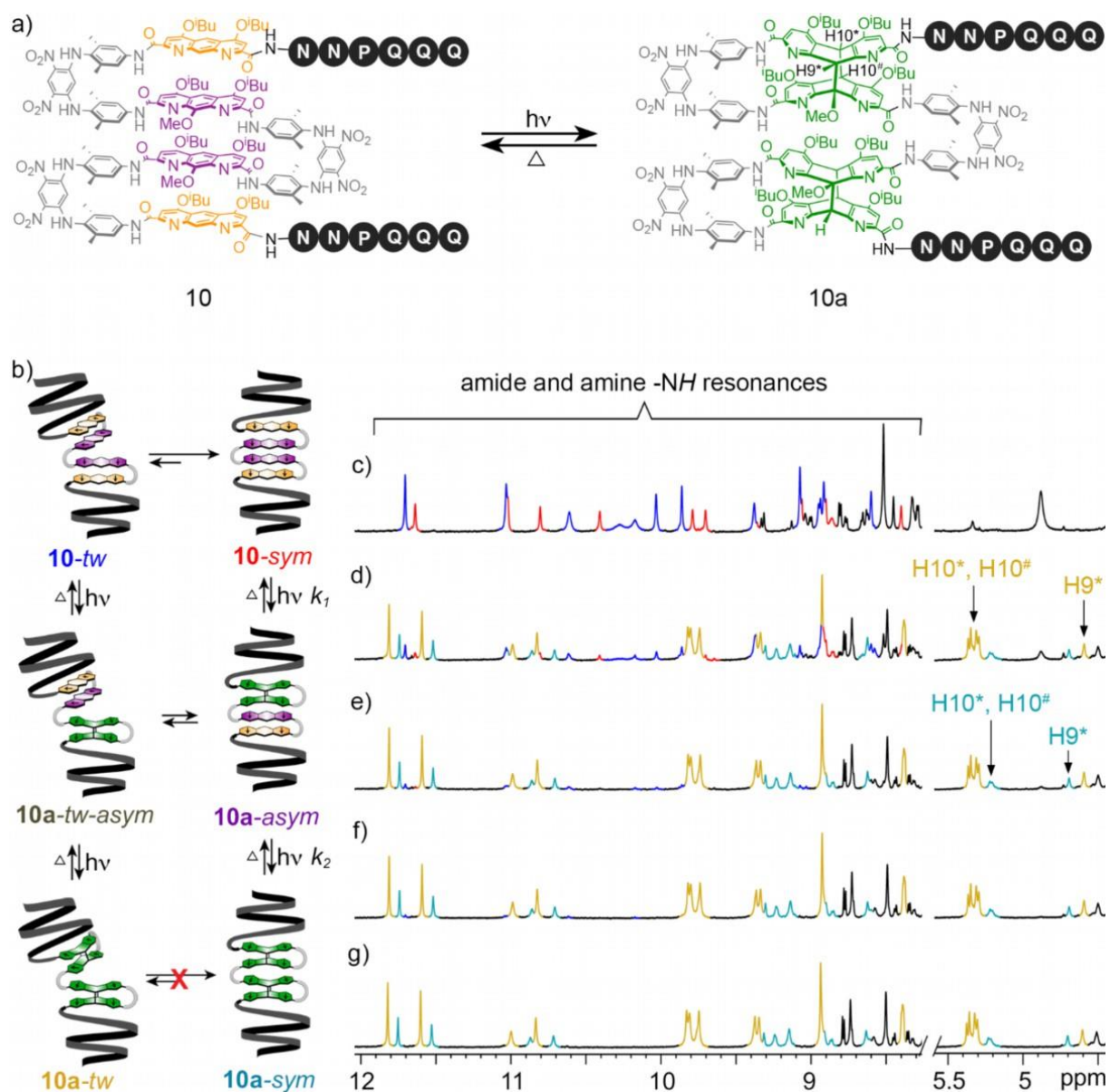


Figure S46. (a) Photoproduct formation of **10** and thermal reversibility. (b) Schematic illustration of stepwise photoadduct formation of different conformers of **10**. Initially two conformers, **10-tw** and **10-sym**, are present in an equilibrium at 298 K. Both have equal possibility to undergo stepwise photoadduct formation to final photoproduct **10a-tw** and **10a-sym** through intermediate monoadducts **10a-tw-asy** and **10a-asy**, respectively. Part of the 700 MHz ^1H NMR spectra at 298K of **10** (1 mM in CDCl_3) under photoirradiation after (c) 0 min; (d) 10 min; (e) 20 min; (f) 30 min; and (g) 60 min. The signals corresponding to the starting two conformers **10-tw** and **10-sym** are shown in blue and red respectively. Signals corresponding to photoproducts, **10a-tw** and **10a-sym**, are shown in olive and cyan respectively. The terms k_1 and k_2 are represent respective formation rate constants with respect to time of irradiation for monoadduct and di-adducts. Clearly, a cooperativity in the second photoadduct was observed with $k_2 \gg k_1$.

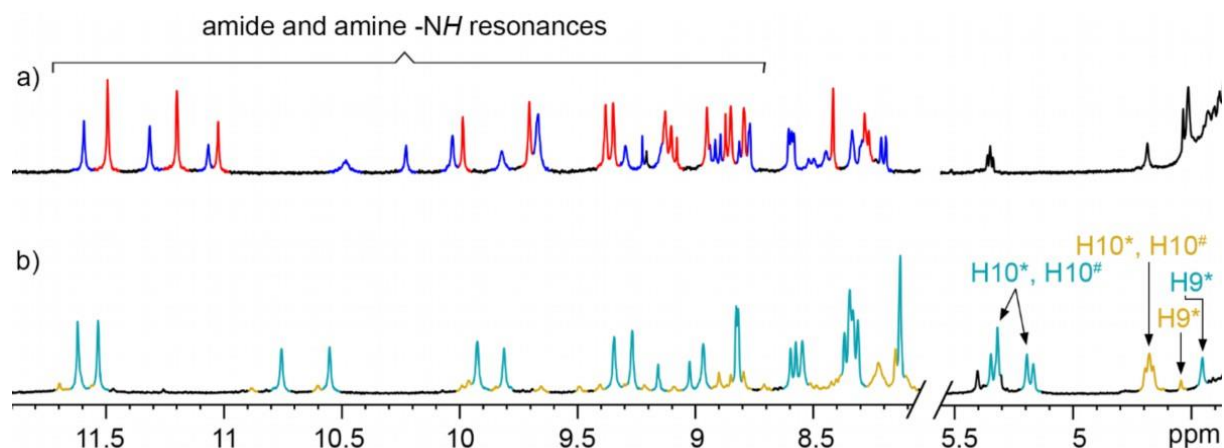


Figure S47. Part of the 400 MHz ^1H NMR spectra at 298K of **10** (1 mM in acetone- d_6) under photoirradiation after (a) 0 min; and (b) 60 min. The signals corresponding to the starting two conformers **10-*tw*** and **10-*sym*** are shown in blue and red respectively. Signals corresponding to photoproducts, **10a-*tw*** and **10a-*sym***, are shown in olive and cyan respectively. Note that proportion of two photoproduct conformers, **10a-*tw*** and **10a-*sym***, is different than that of their parent conformers.

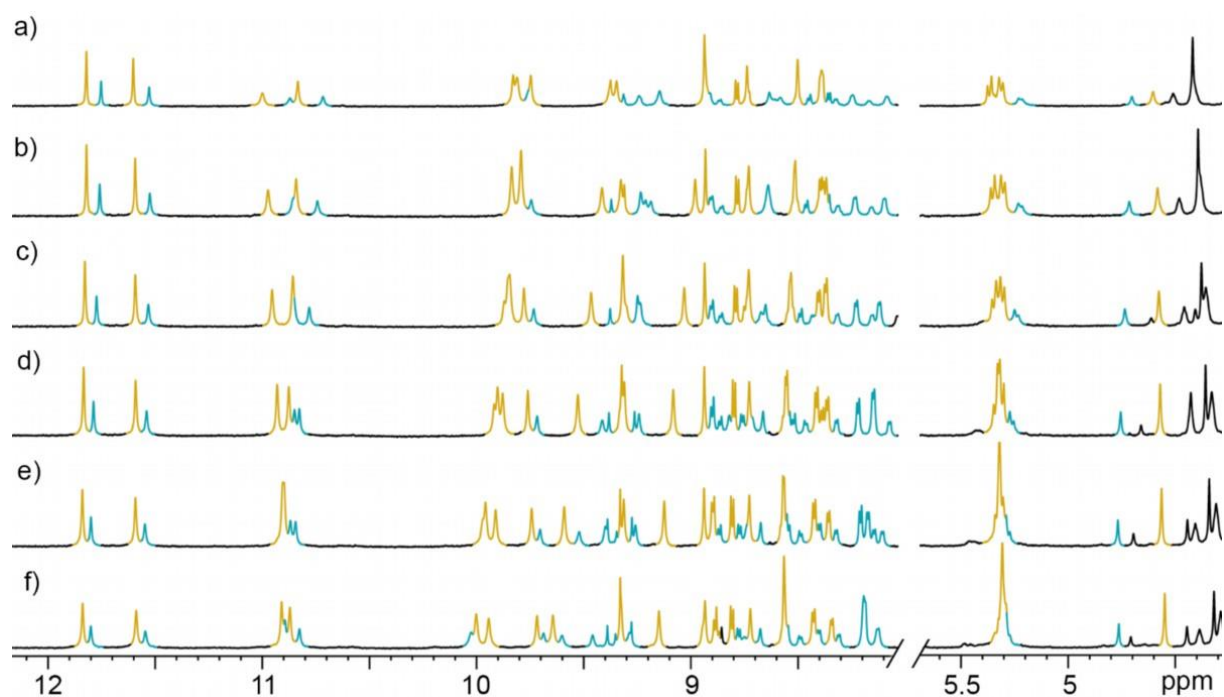


Figure S48. Part of the variable temperature ^1H NMR spectra (700 MHz, CDCl_3) of **10a** (1 mM) at different temperature. a) 298K; b) 288K; c) 278K; d) 268K; e) 258K; and f) 248K. Peaks highlighted in olive and cyan corresponds to the **10a-*tw*** and **10a-*sym*** respectively. After photoproduct formation conformers have negligible influence of temperature variation.

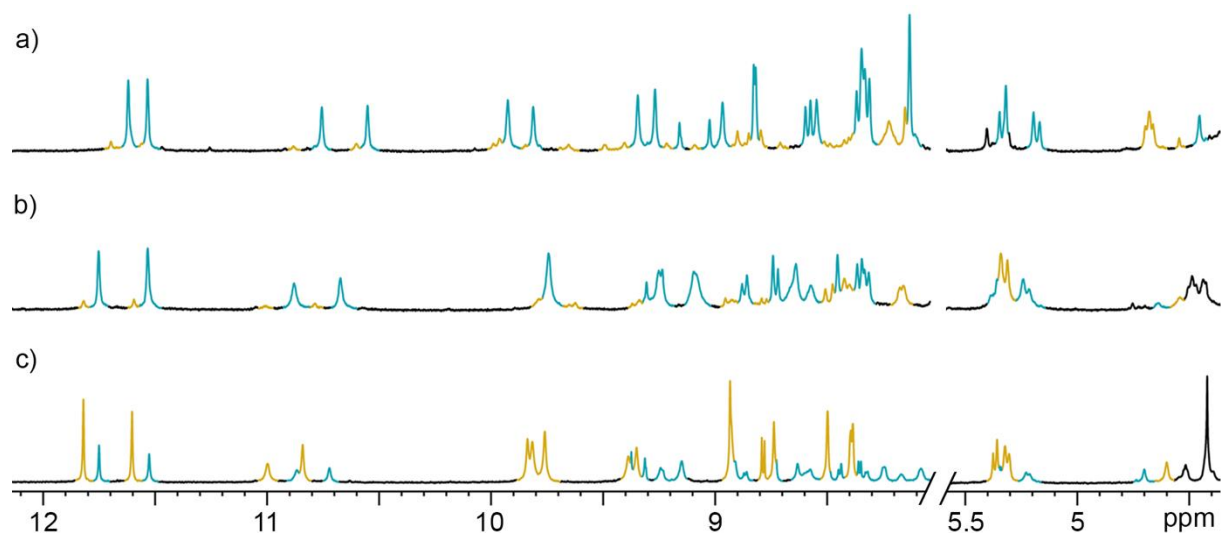


Figure S49. (a) Part of the 400 MHz ^1H NMR spectra of **10a** at 298K in acetone- d_6 . (b) The ^1H NMR spectra of **10a** was recorded in CDCl_3 after 60 min photoirradiation in acetone. (c) In comparison, the ^1H NMR spectra was recorded in CDCl_3 after 60 min irradiation in CDCl_3 . The signals correspond to two conformers, **10a-*tw*** and **10a-*sym***, are shown in olive and cyan, respectively. The proportion of two photoproducts has significant difference when their starting conformers were irradiated in acetone- d_6 and CDCl_3 . It is also evident that the photoproducts have no solvent effects.

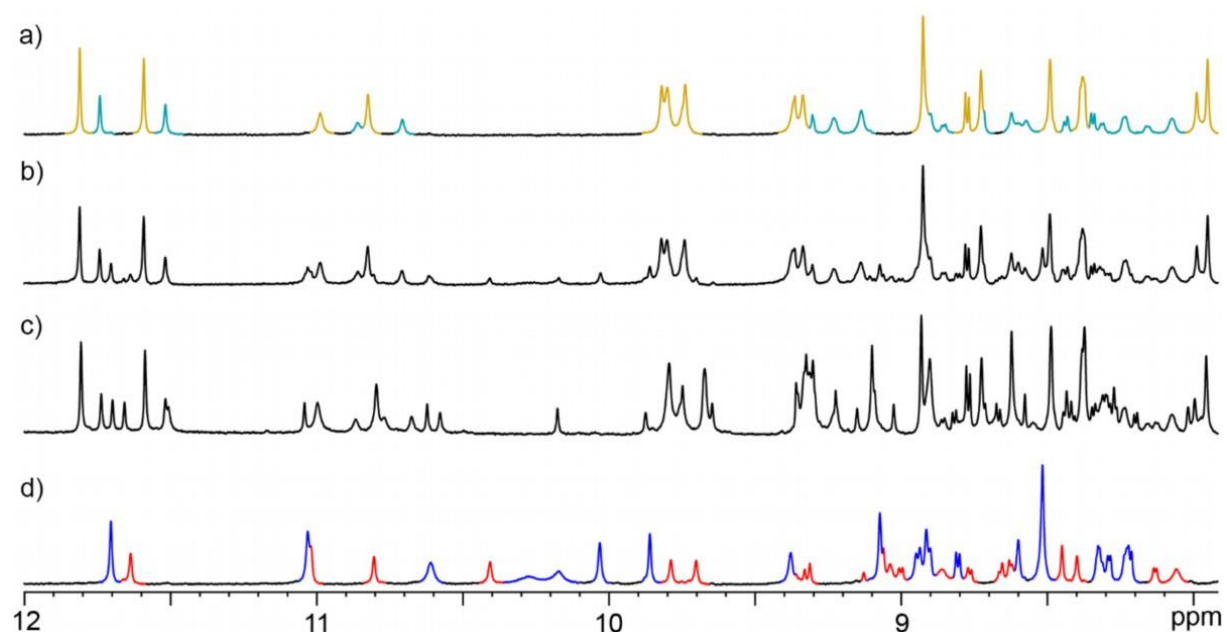


Figure S50. Part of the 700 MHz ^1H NMR spectra of **10a** at 298K (a) in CDCl_3 ; and after (b) 3 h; (c) 10 h; and (d) 48 h of heating in CDCl_3 at 333 K. The signals corresponding to the starting two conformers **10-*tw*** and **10-*sym*** are shown in blue and red respectively and signals corresponds to their respective photoproducts, **10a-*tw*** and **10a-*sym***, are shown in olive and cyan.

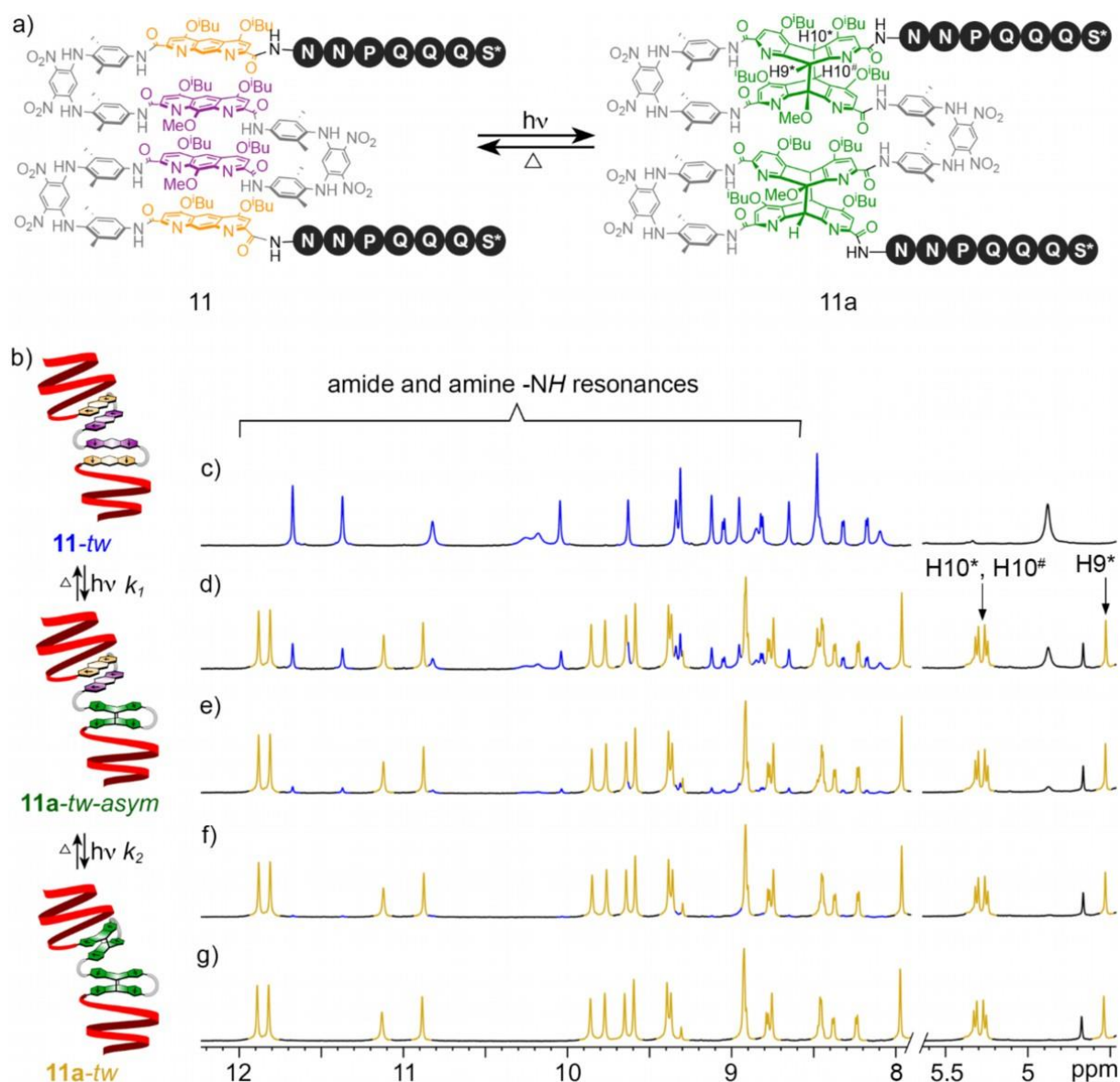


Figure S51. (a) Photoproduct formation of **11** and thermal reversibility. (b) Schematic illustration of stepwise photoadduct formation of **11**. Initially, conformer **11-tw** was present in the solution at 298 K. It can undergo stepwise photoadduct formation to final photoproduct **11a-tw** through intermediate monoadducts **11a-tw-asy**. Part of the 700 MHz ^1H NMR spectra at 298K of **11** (1 mM in CDCl_3) under photoirradiation after (c) 0 min; (d) 10 min; (e) 20 min; (f) 30 min; and (g) 40 min. The signals corresponding to the starting compound, **11-tw**, have been shown in blue and signals corresponding to photoproduct, **11a-tw**, are shown in olive. The terms k_1 and k_2 are represent respective formation rate constants with respect to time of irradiation for monoadduct and di-adducts. Interestingly, no monoadduct, **11a-tw-asy**, was identified which is indicating a high cooperativity in the second photoadduct.

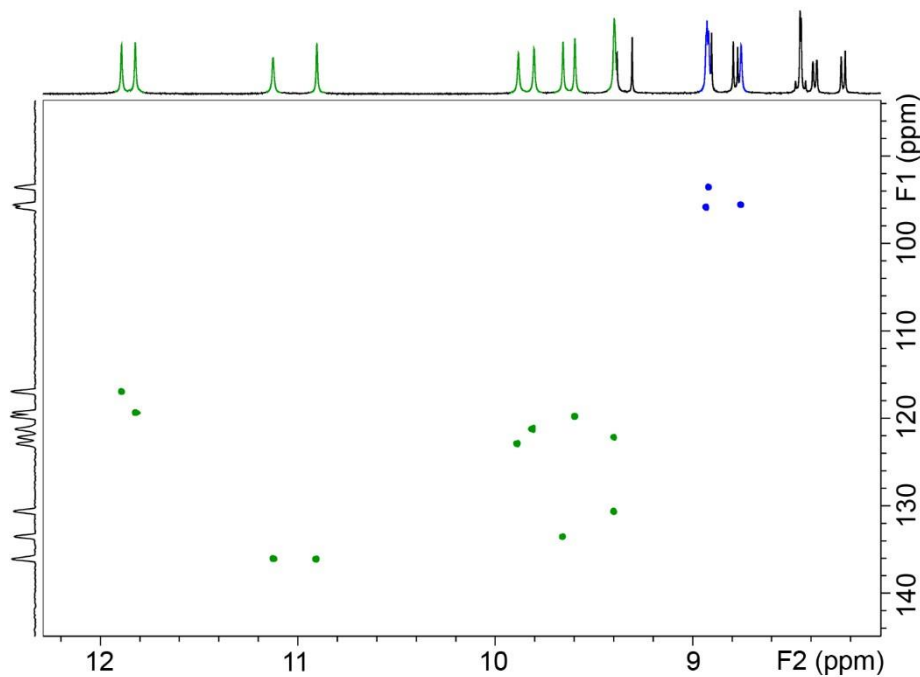


Figure S52. ^1H - ^{15}N HSQC spectrum (400 MHz, CDCl_3) of photoproduct **11a** (1 mM) at 298K. The $-\text{NH}$ amide signals are depicted in green and amine protons from turn units are shown in blue colors.

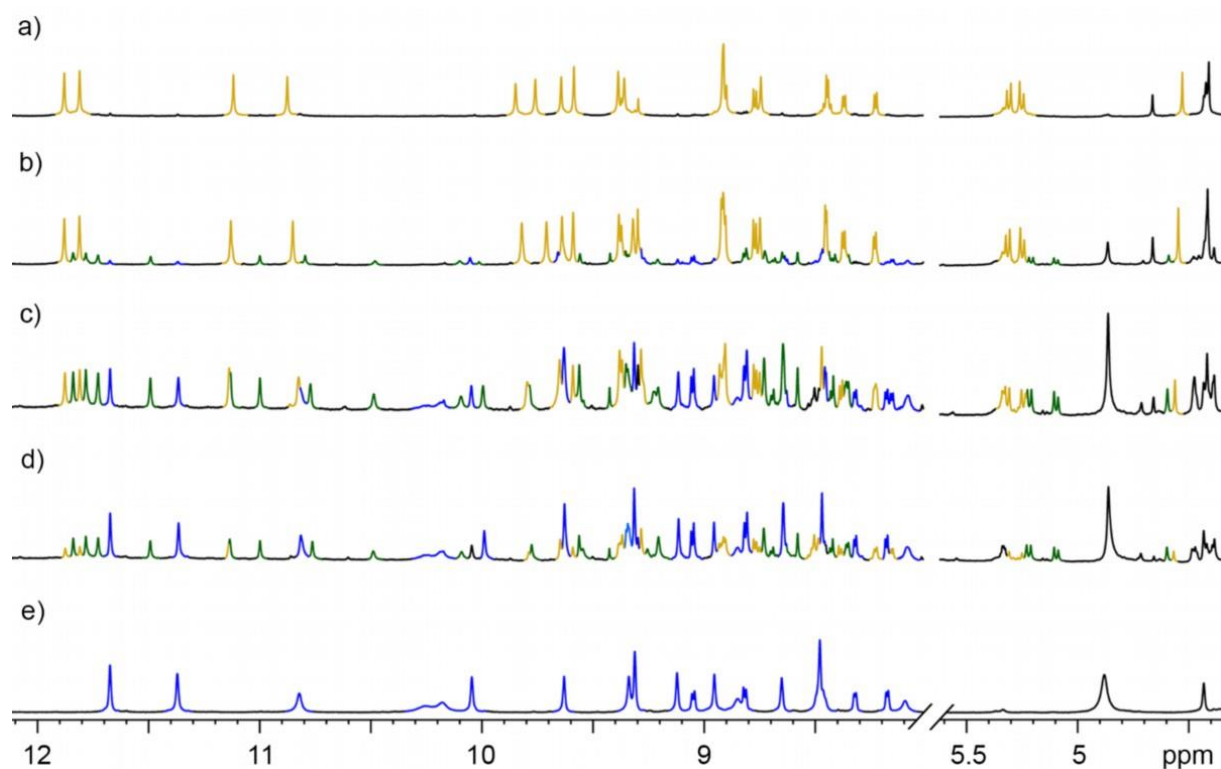


Figure S53. Part of the 700 MHz ^1H NMR spectra of **11a** at 298K (a) in CDCl_3 ; and after (b) 3 h; (c) 10 h; (e) 20 h; and (e) 48 h of heating in CDCl_3 at 333 K. The signals corresponding to the **11a** and **11** are showing in olive and blue respectively. In addition, signals correspond to monoadduct, **11a-tw-asm**, are shown in green.

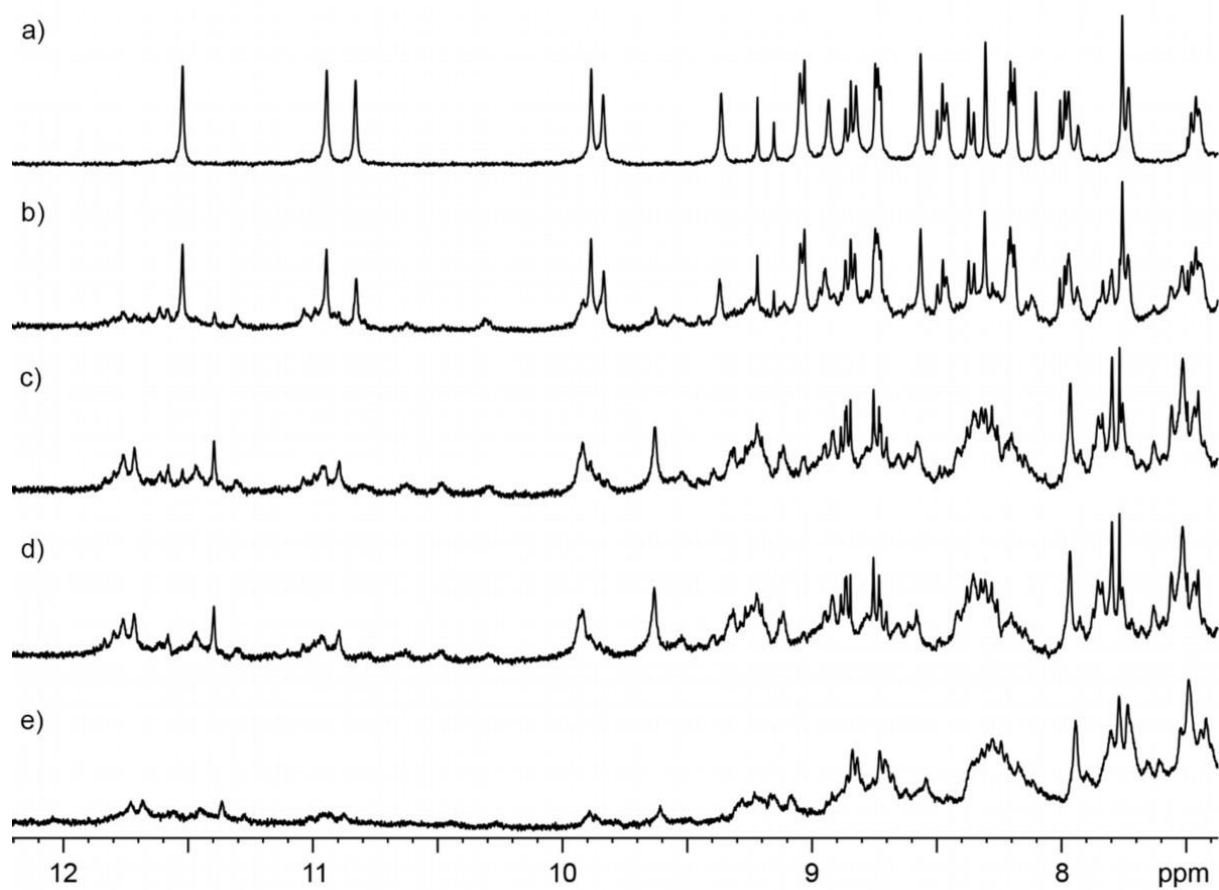


Figure S54. Part of the 700 MHz ¹H NMR spectra at 298K of **12** (1 mM in CDCl₃) under photoirradiation after (c) 0 min; (d) 30 min; (e) 2 h; (f) 3 h; and (g) 5 h. The signals corresponding to the photoproducts become significantly broad which makes difficult to monitor photoirradiation process.

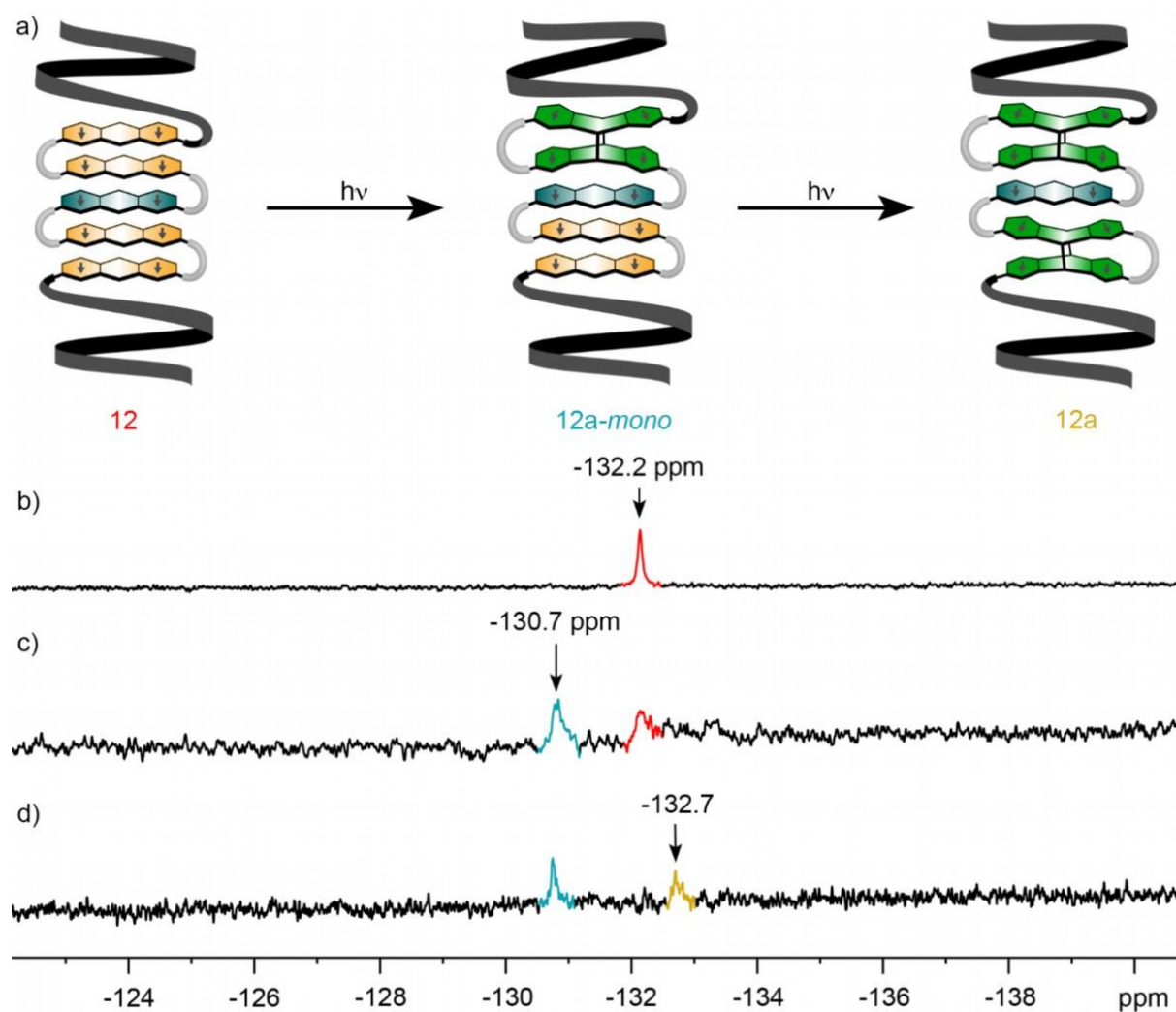


Figure S55. (a) Schematic illustration of stepwise photoadduct formation of **12**. It can undergo stepwise photoadduct formation to final photoproduct **12a** through intermediate monoadducts **12-mono**. Part of the 376 MHz ^{19}F NMR spectra at 298K of **12** (1 mM in CDCl_3) under photoirradiation after (b) 0 min; (c) 1 h; and (d) 5 h. The signals corresponding to the starting compound, **12**, have been shown in red and signals corresponding to photoproduct, **12a-mono** and **12a** are shown in cyan and olive respectively.

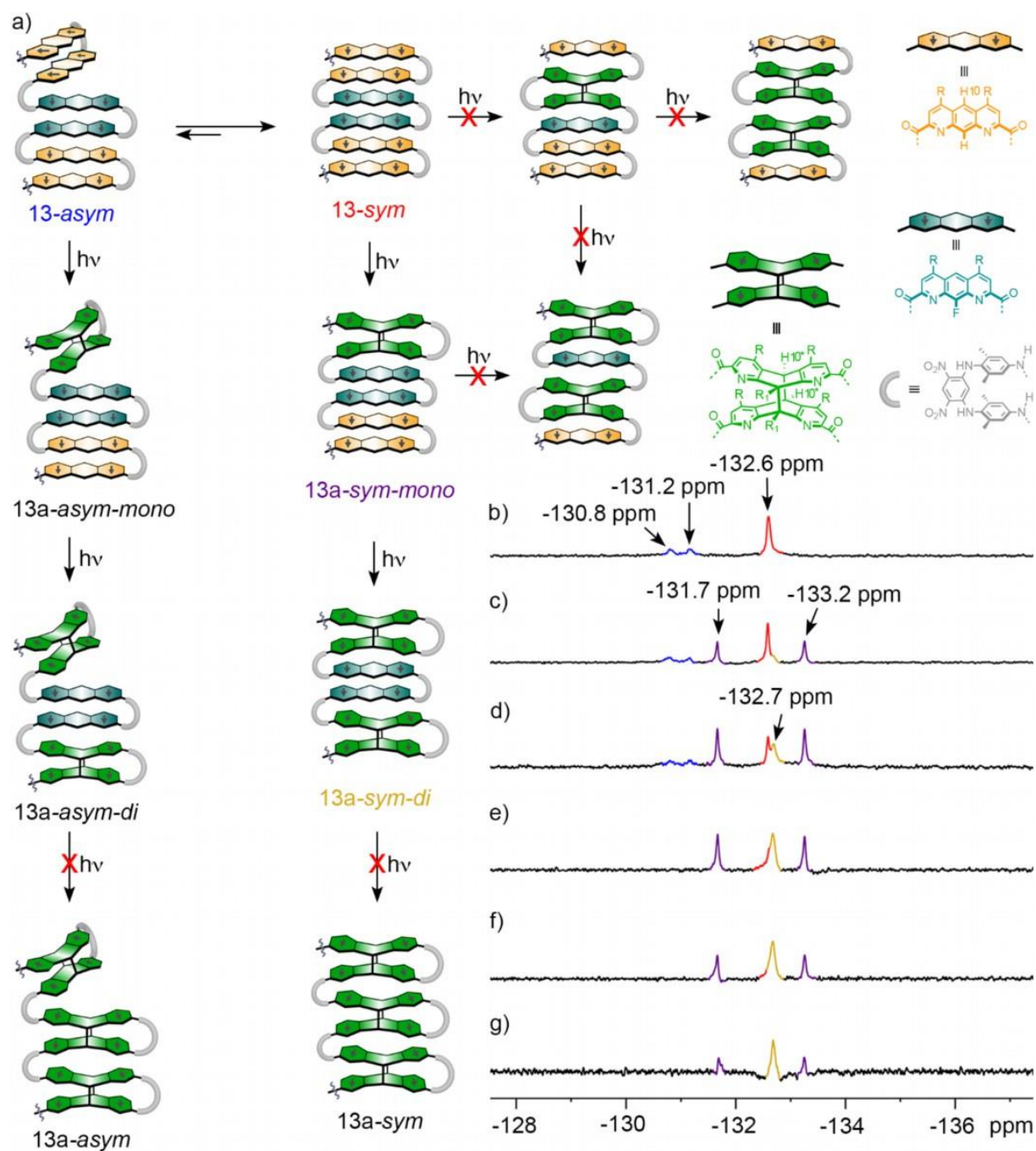


Figure S56. a) Schematic illustration of stepwise photoadduct formation of **13**. Initially two conformers, **13-asy** and **13-sym**, are present in an equilibrium at room temperature. Both have equal possibility to undergo stepwise photoadduct formation to final photoproduct **13a-asy** and **13a-sym** respectively. However, other uncontrolled photoadduct formation can be ruled out based on the design principle. In the illustration, helices are omitted for clarity. Chemical formula of each abbreviation given in the top right corner. Part of the 376 MHz ^{19}F NMR spectra of **13** (1 mm in acetone- d_6) at 298 K under photoirradiation after b) 0 min; c) 1 h; d) 2 h; e) 4 h; f) 6 h; and g) 12 h. Signals correspond to **13a-asy** and **13a-sym** are shown in blue and red respectively. Signals assigned to photoproducts **13a-sym-mono** and **13a-sym-di** are highlighted in purple and olive respectively. Note that final desired photoproducts **13a-asy** and **13a-sym** were not formed due to the low reactivity among two central A^{F} units.

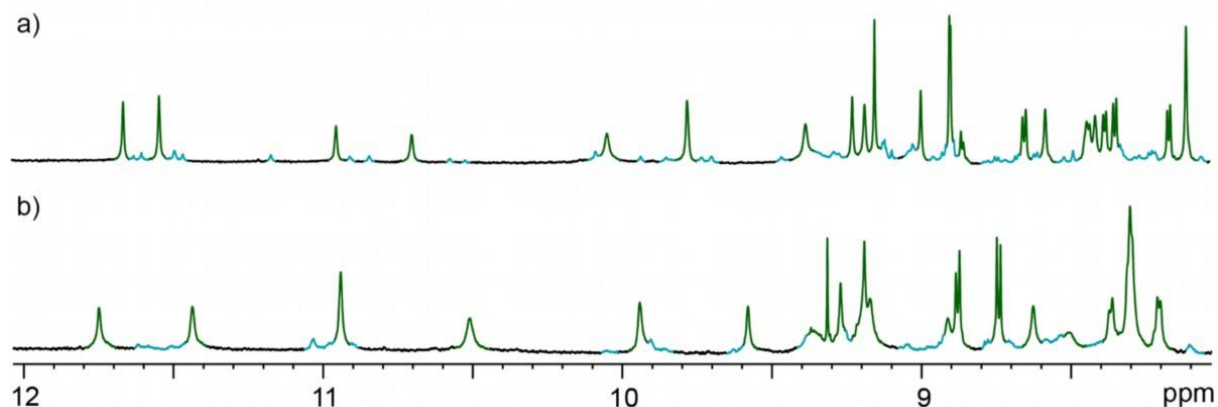


Figure S57. Part of the 700 MHz ^1H NMR spectra at 298K of **14a** (obtained from photoirradiation of **14** in acetone- d_6) in (a) acetone- d_6 ; and (c) CDCl_3 . The signals corresponding to conformers **14a-sym** and **14a-asm** have been shown in green and cyan respectively.

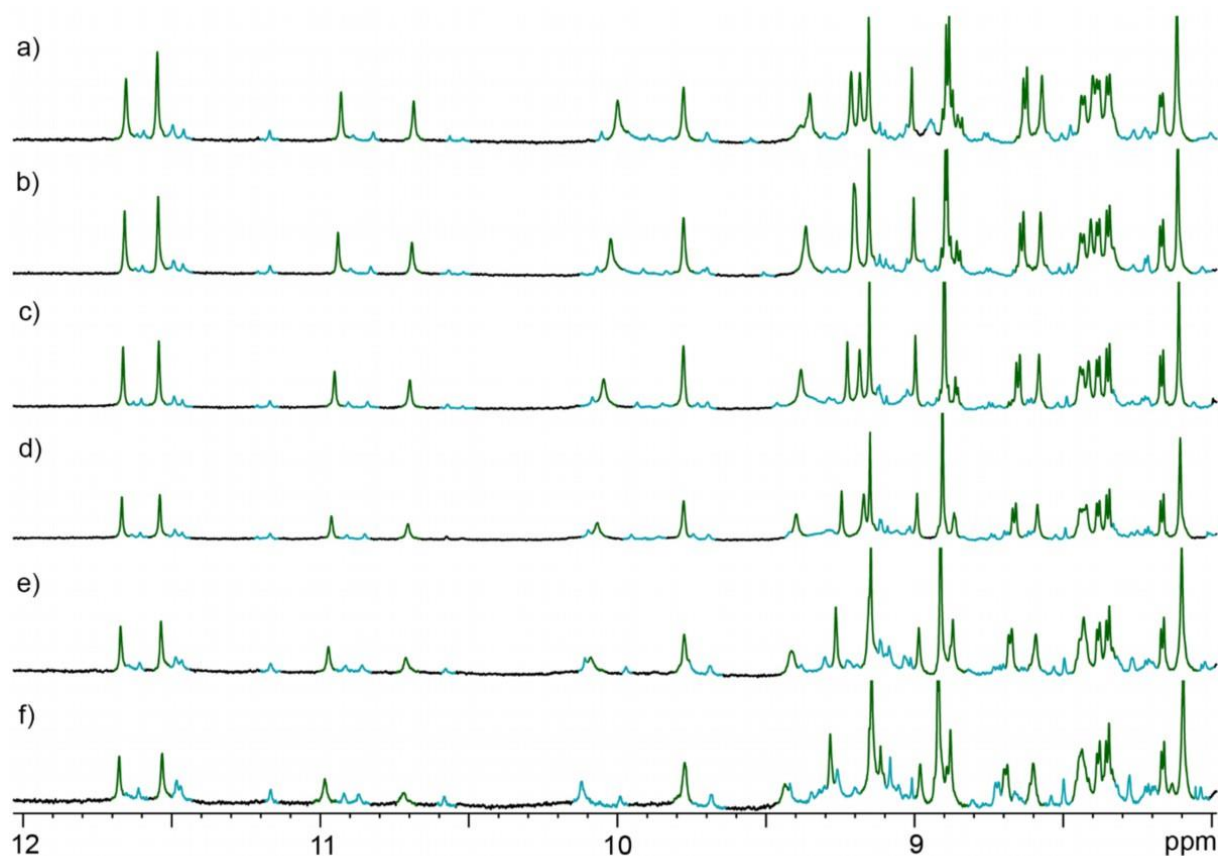


Figure S58. Part of the variable temperature ^1H NMR spectra (700 MHz, acetone- d_6) of **14a** at different temperature. a) 318K; b) 308K; c) 298K; d) 288K; e) 278K; and f) 258K. The signals corresponding to conformers **14a-sym** and **14a-asm** have been shown in green and cyan respectively. After photoproduct formation conformers have negligible influence of temperature variation.

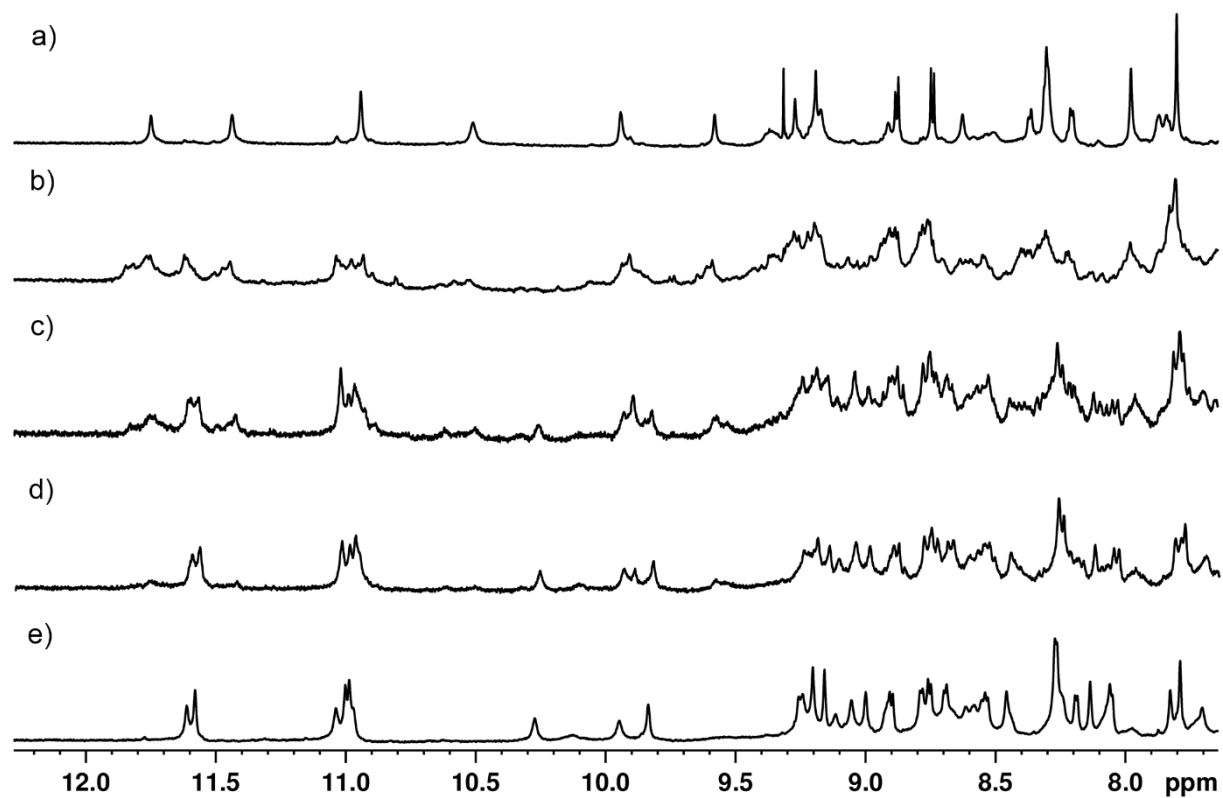


Figure S59. Part of the 400 MHz ¹H NMR spectra of **14a** at 298K (a) in CDCl₃; and after (b) 10 h; (c) 30 h; and (d) 60 h of heating in CDCl₃ at 333 K; e) Reference spectrum of **14**. The signals of the intermediate compounds are broad and correspond to photoproducts **14a-sym-mono**, **14a-sym-di**.

6. X-ray crystallography

Table S1. Crystal data and structure refinement for **8**.

<i>Identification code</i>	8
<i>Empirical formula</i>	C ₂₅₉ H ₂₅₇ Cl ₉ N ₄₈ O ₄₄
<i>Formula weight</i>	5065.16
<i>Temperature/K</i>	130
<i>Crystal system</i>	monoclinic
<i>Space group</i>	P2 ₁ /n
<i>a/Å</i>	24.5746(6)
<i>b/Å</i>	49.5101(11)
<i>c/Å</i>	27.7011(6)
<i>α/°</i>	90
<i>β/°</i>	102.621(2)
<i>γ/°</i>	90
<i>Volume/Å³</i>	32889.3(13)
<i>Z</i>	4
<i>ρ_{calc}/cm³</i>	1.023
<i>μ/mm⁻¹</i>	1.232
<i>F(000)</i>	10608.0
<i>Crystal size/mm³</i>	0.05 × 0.05 × 0.05
<i>Radiation</i>	CuKα (λ = 1.54184)
<i>2θ range for data collection/°</i>	3.57 to 111.084
<i>Reflections collected</i>	135800
<i>Independent reflections</i>	41120 [R _{int} = 0.0594, R _{sigma} = 0.0530]
<i>Data/restraints/parameters</i>	41120/111/3281
<i>Goodness-of-fit on F²</i>	1.396
<i>Final R indexes [I >= 2σ (I)]</i>	R ₁ = 0.1378, wR ₂ = 0.3880
<i>Final R indexes [all data]</i>	R ₁ = 0.1857, wR ₂ = 0.4189
<i>CCDC #</i>	2144662

Table S2. Crystal data and structure refinement for **10**

<i>Identification code</i>	10
<i>Empirical formula</i>	C _{292.5} H ₂₇₅ Cl ₁₂ N ₅₆ O ₅₄
<i>Formula weight</i>	5864.07
<i>Temperature/K</i>	130
<i>Crystal system</i>	triclinic
<i>Space group</i>	P-1
<i>a/Å</i>	24.1819(8)
<i>b/Å</i>	28.1154(8)
<i>c/Å</i>	41.6885(7)
<i>α/°</i>	103.493(2)
<i>β/°</i>	91.075(2)
<i>γ/°</i>	114.626(2)
<i>Volume/Å³</i>	24837.6(12)
<i>Z</i>	2
<i>ρ_{calc}/cm³</i>	0.784
<i>μ/mm⁻¹</i>	1.026
<i>F(000)</i>	6116.0
<i>Crystal size/mm³</i>	0.02 × 0.02 × 0.02
<i>Radiation</i>	CuKα (λ = 1.54178)
<i>2θ range for data collection/°</i>	3.586 to 95.304
<i>Reflections collected</i>	80745
<i>Independent reflections</i>	41817 [R _{int} = 0.0667, R _{sigma} = 0.0900]
<i>Data/restraints/parameters</i>	41817/17375/3693
<i>Goodness-of-fit on F²</i>	1.843
<i>Final R indexes [I >= 2σ (I)]</i>	R ₁ = 0.1995, wR ₂ = 0.5081
<i>Final R indexes [all data]</i>	R ₁ = 0.2369, wR ₂ = 0.5345
<i>CCDC #</i>	2144663

Table S3. Crystal data and structure refinement for **13-sym**.

<i>Identification code</i>	13-sym
<i>Empirical formula</i>	C ₃₅₂ H ₂₇₅ F ₂ N ₇₂ O ₆₈
<i>Formula weight</i>	6639.42
<i>Temperature/K</i>	130
<i>Crystal system</i>	orthorhombic
<i>Space group</i>	Pbca
<i>a/Å</i>	45.2959(12)
<i>b/Å</i>	40.8492(8)
<i>c/Å</i>	61.6329(12)
<i>α/°</i>	90
<i>β/°</i>	90
<i>γ/°</i>	90
<i>Volume/Å³</i>	114039(4)
<i>Z</i>	8
<i>ρ_{calc}/cm³</i>	0.773
<i>μ/mm⁻¹</i>	0.463
<i>F(000)</i>	27624.0
<i>Crystal size/mm³</i>	0.05 × 0.05 × 0.05
<i>Radiation</i>	CuKα (λ = 1.54178)
<i>2θ range for data collection/°</i>	4.088 to 89.078
<i>Reflections collected</i>	345597
<i>Independent reflections</i>	44023 [R _{int} = 0.1821, R _{sigma} = 0.0816]
<i>Data/restraints/parameters</i>	44023/29266/4339
<i>Goodness-of-fit on F²</i>	1.767
<i>Final R indexes [I >= 2σ (I)]</i>	R ₁ = 0.2454, wR ₂ = 0.5257
<i>Final R indexes [all data]</i>	R ₁ = 0.2877, wR ₂ = 0.5581
<i>CCDC #</i>	2144666

Table S4. Crystal data and structure refinement for **14-*asym***

<i>Identification code</i>	14-<i>asym</i>
<i>Empirical formula</i>	C ₄₁₁ H ₄₃₈ N ₇₂ O ₈₁
<i>Formula weight</i>	7682.31
<i>Temperature/K</i>	120
<i>Crystal system</i>	triclinic
<i>Space group</i>	P-1
<i>a/Å</i>	21.130(4)
<i>b/Å</i>	34.790(7)
<i>c/Å</i>	39.590(8)
<i>α/°</i>	69.20(3)
<i>β/°</i>	77.70(3)
<i>γ/°</i>	86.10(3)
<i>Volume/Å³</i>	26581(11)
<i>Z</i>	2
<i>ρ_{calc}/cm³</i>	0.960
<i>μ/mm⁻¹</i>	0.358
<i>F(000)</i>	8112.0
<i>Crystal size/mm³</i>	0.05 × 0.05 × 0.05
<i>Radiation</i>	Ga (λ = 1.340)
<i>2θ range for data collection/°</i>	4.066 to 85.182
<i>Reflections collected</i>	157328
<i>Independent reflections</i>	54334 [R _{int} = 0.1338, R _{sigma} = 0.1068]
<i>Data/restraints/parameters</i>	54334/4510/4942
<i>Goodness-of-fit on F²</i>	1.202
<i>Final R indexes [I >= 2σ(I)]</i>	R ₁ = 0.1266, wR ₂ = 0.3449
<i>Final R indexes [all data]</i>	R ₁ = 0.1886, wR ₂ = 0.3828
<i>CCDC #</i>	2144667

8. Molecular mechanics

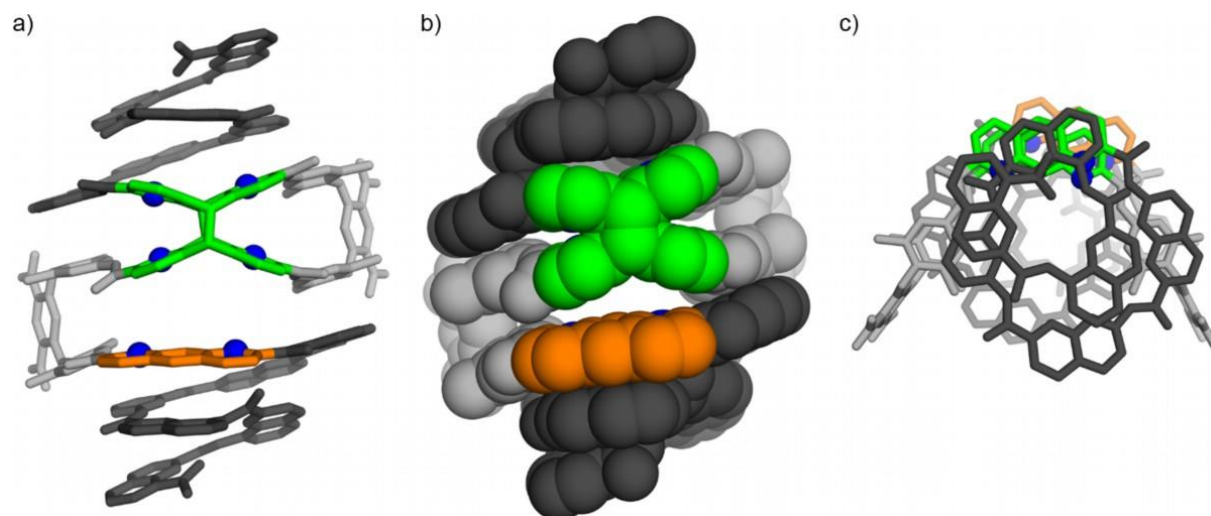


Figure S60. (a) Back view of the energy-minimized molecular model of **8a** using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b) and (c) Back and top view of the same in space filling and tube representation, respectively. (The structures are shown with color-coded monomers as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

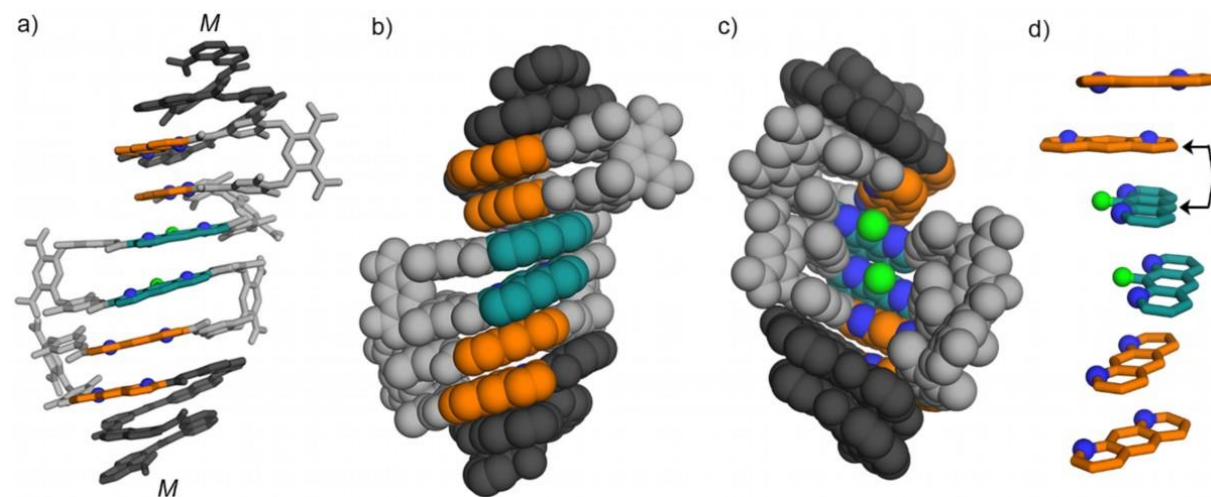


Figure S61. (a) Back view of the energy-minimized molecular model of **13-asm** using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b) and (c) Back and front view of the same in space filling representation, respectively. (d) Orientation of the diazaanthracene units. Arrows indicate the only pair of antiparallel diazaanthracene units. The structures are shown with color-coded monomers as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

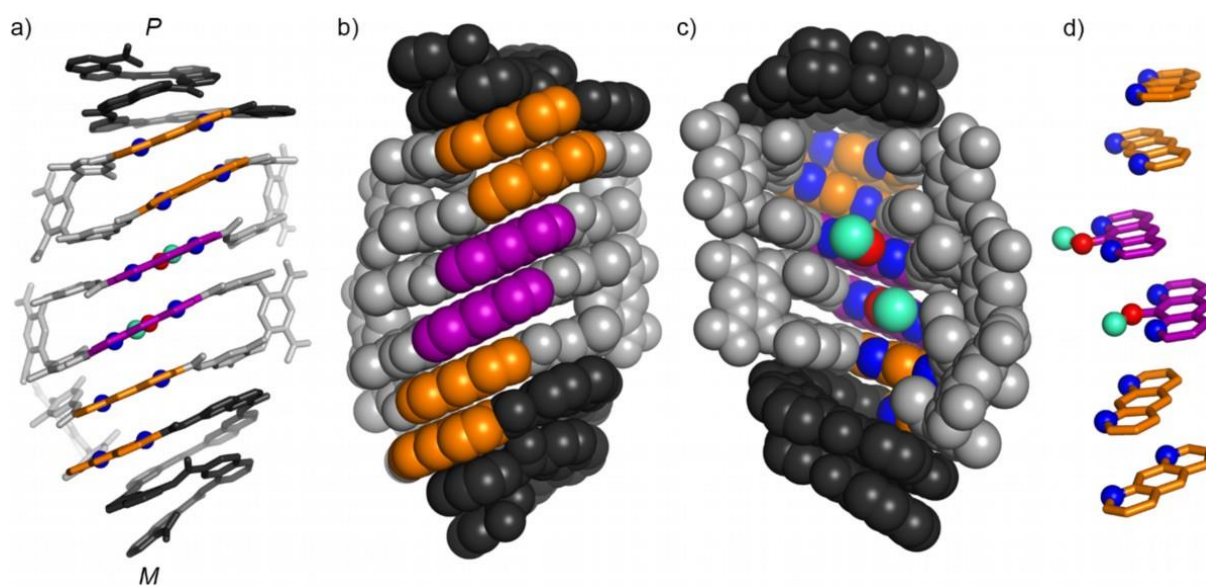


Figure S62. (a) Back view of the energy-minimized molecular model of **14-sym** using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b) and (c) Back and front view of the same in space filling representation, respectively. (d) Parallel orientation of the diazaanthracene units. The structures are shown with color-coded monomers as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

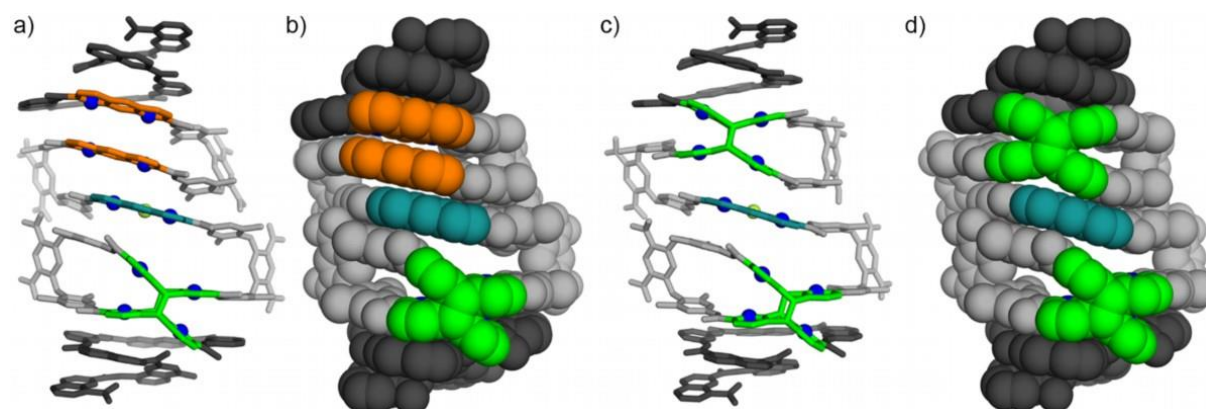


Figure S63. (a) and (b) Back view of the energy-minimized molecular model of mono (**12a-mono**) and di-photoadduct (**12a**) products of **12**, respectively, using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b) and (d) Back view of the same in space filling representation, respectively. The structures are shown with color-coded representation as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

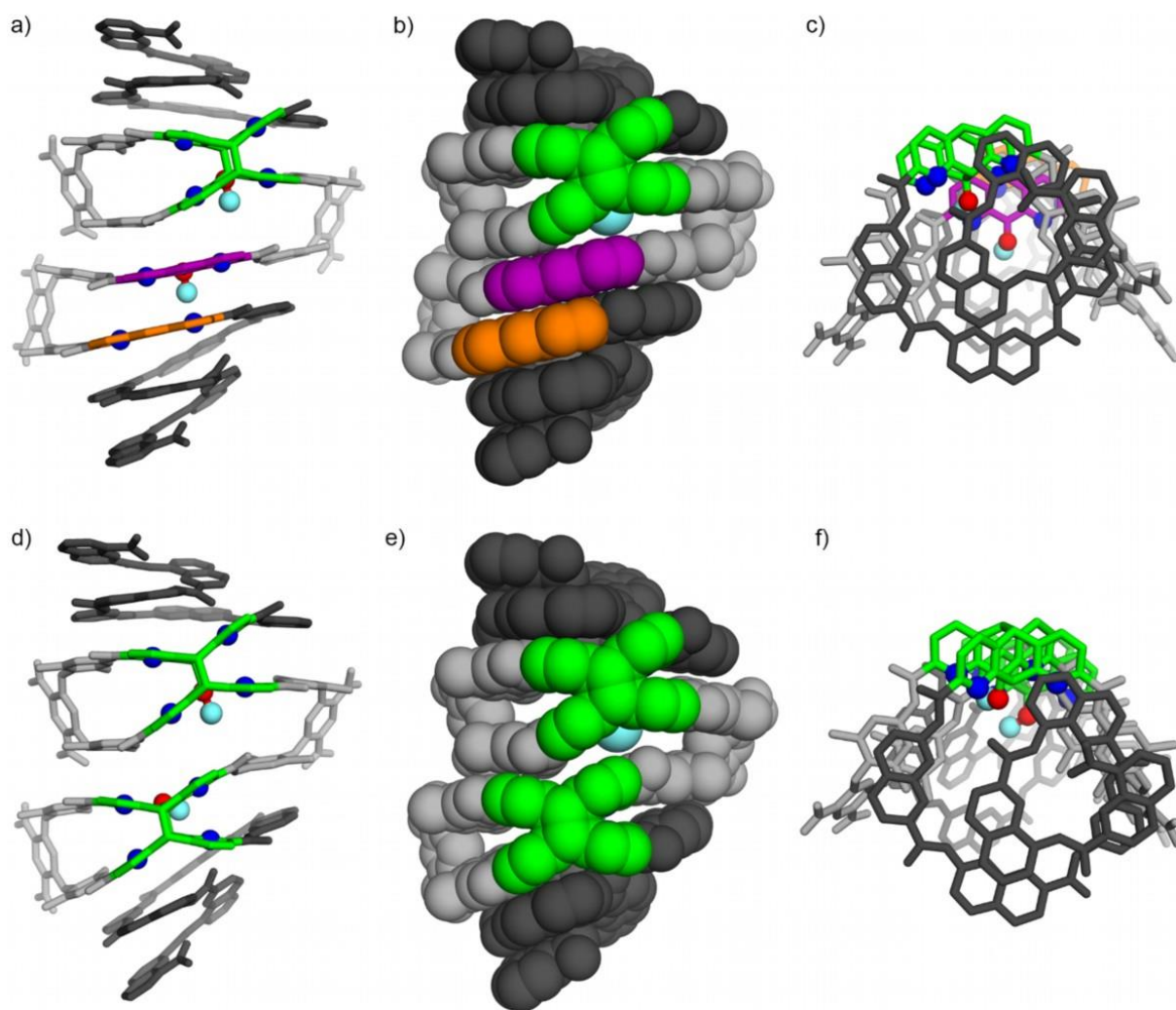


Figure S64. (a) and (d) Back view of the energy-minimized molecular model of mono (**10a-asy**) and di-photoadduct (**10a-sym**) products of **10-sym**, respectively, using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b) and (e) Back view of the same in space filling representation, respectively. (c) and (f) Represents the respective top views. The structures are shown with color-coded representation as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

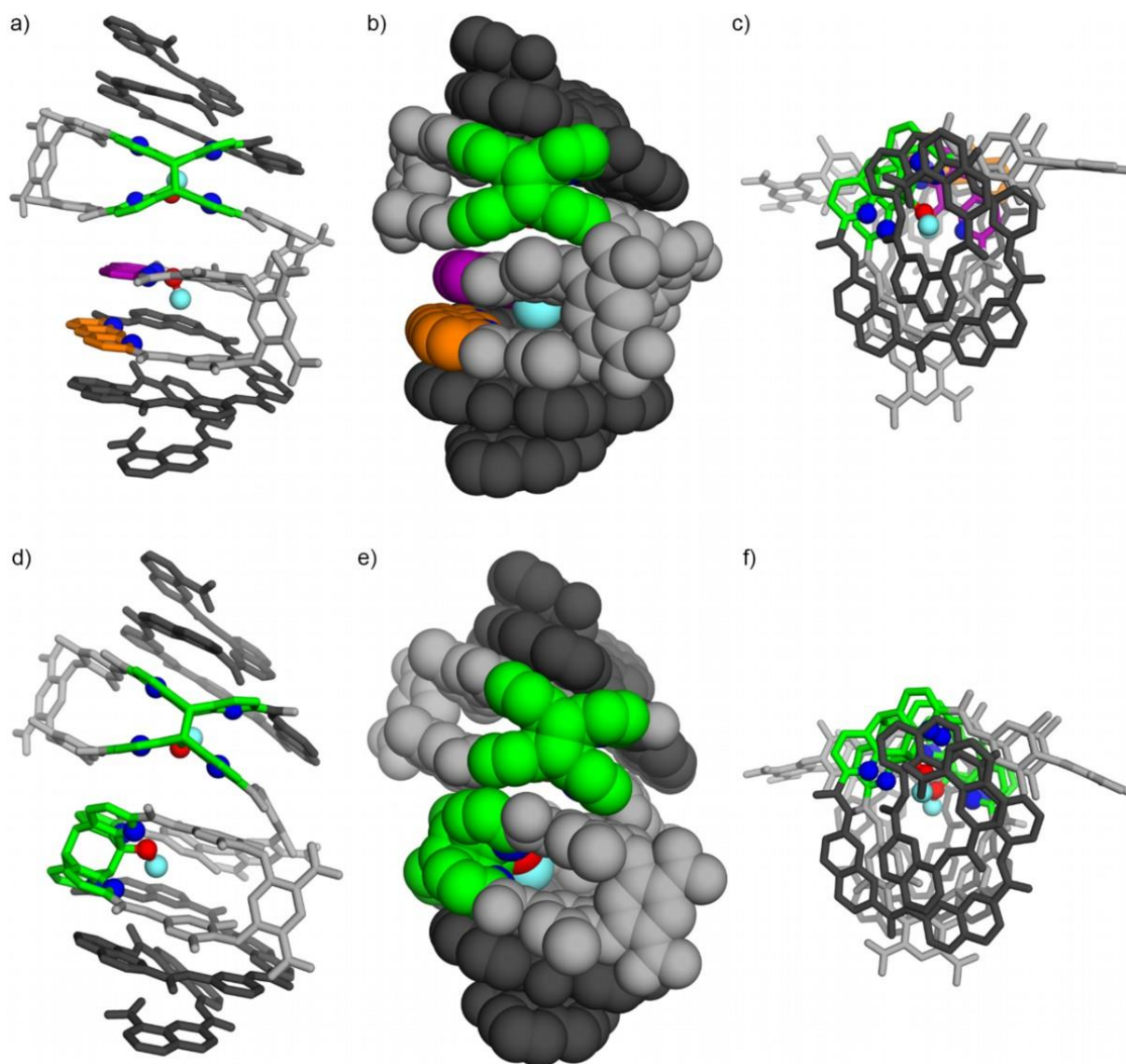


Figure S65. (a) and (d) Back view of the energy-minimized molecular model of mono (**10a-*tw-asy***) and di-photoadduct (**10a-*tw***) products of **10-*tw***, respectively, using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b) and (e) Back view of the same in space filling representation, respectively. (c) and (f) Represents the respective top views. The structures are shown with color-coded representation as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

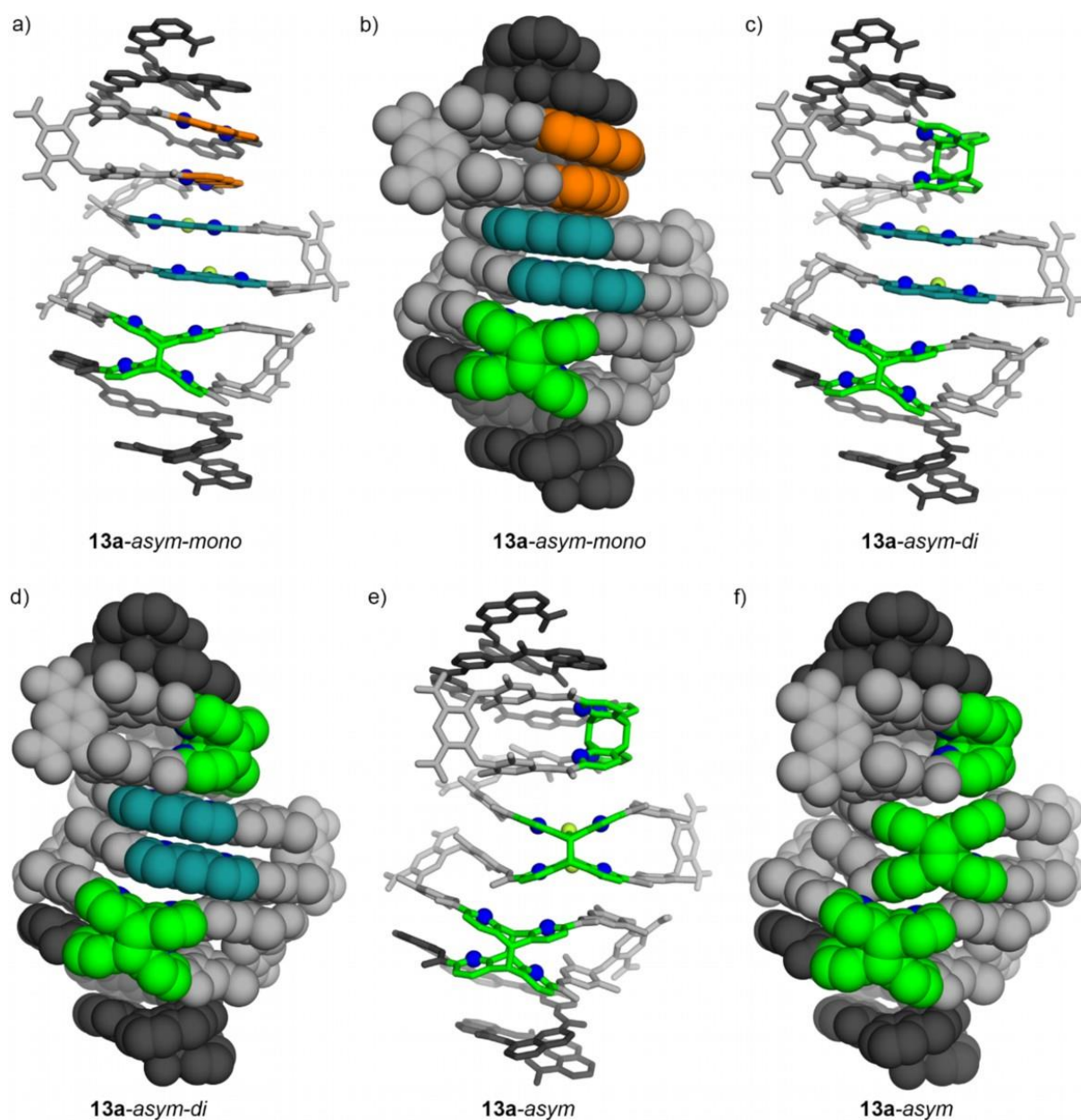


Figure S66. Stepwise photoadduct formation in **13-asm**. (a), (c) and (e) Back view of the energy-minimized molecular model of *mono-* (**13a-asm-mono**), *di-* (**13a-asm-di**) and *tri-* (**13a-asm**) photoadduct products of **13-asm**, respectively, using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b), (d) and (f) Back view of the same in space filling representation, respectively. The structures are shown with color-coded representation as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

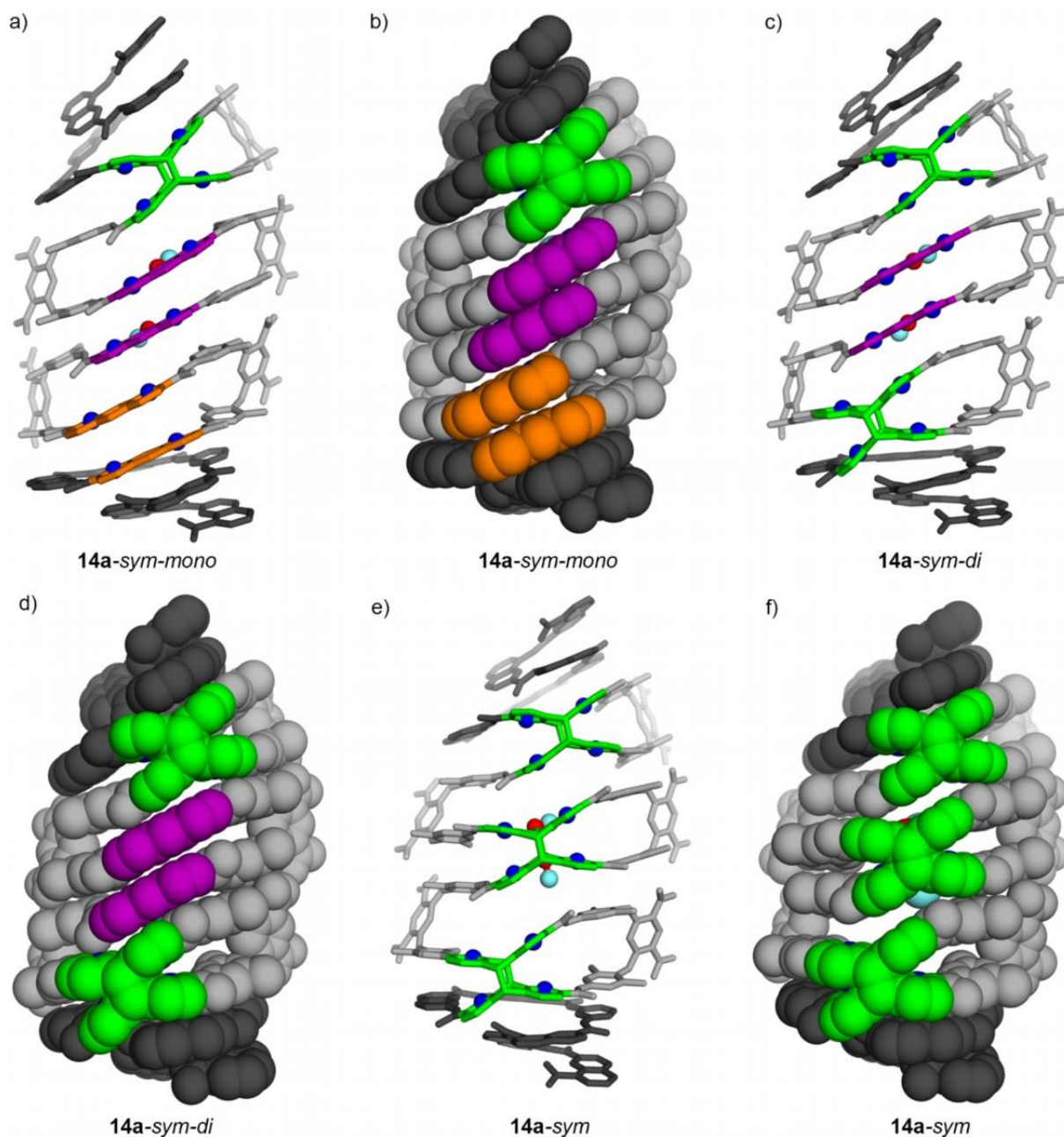


Figure S67. Stepwise photoadduct formation in **14-sym**. (a), (c) and (e) Back view of the energy-minimized molecular model of *mono-* (**14a-sym-mono**), *di-* (**14a-sym-di**) and *tri-* (**14a-sym**) photoadduct products of **14-sym**, respectively, using the Merck Molecular Force Field static (MMFFs) shown in tube representation. (b), (d) and (f) Back view of the same in space filling representation, respectively. The structures are shown with color-coded representation as defined earlier. Blue balls depicting endocyclic nitrogen atoms. Hydrogen atoms and side chains are omitted for clarity.

9. References

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10. NMR Spectra

