Supporting Information

Assessing Stabilization Through π - π Interactions in Aromatic Oligoamide β -Sheet Foldamers.

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Table of contents

1. Synthetic schemes	S2							
2. NMR experiments								
3. Solution structure determination	S5							
4. Crystallography	S9							
4.1. Methods for X-ray crystallography	S9							
4.2. Summary of X-Ray crystallographic data	S10							
5. Experimental section								
6. ¹ H NMR spectra of all relevant synthetic intermediates and title compounds								
7. References	S38							



Preparation of compounds 1 and 2

Reagents and conditions: a) 3 equiv. Boc₂O, THF, +70 °C, 8 days, 39%. b) H₂, 10% Pd/C, AcOEt, rt, 16h, 99% for **8**, 96% for **10**. c) 0.5 equiv. 1,5-difluoro-2,4-dinitrobenzene, DMSO, 4h, rt then 0.5 equiv. DIEA, DMSO, rt, 2 days, 86% for **1**, 72% for **2**. d) 0.9 equiv. 4-(ethoxycarbonyl)-2,5-diisobutoxybenzoic acid chloride, 1 equiv. DIEA, CHCl₃, 16h, rt, 71%.

Preparation of quinoline and quinoline dimers derivatives



Reagents and conditions: a) 1 equiv. dimethyl acetylenedicarboxylate, MeOH, rt, 3h then reflux, 1h, 90%. b) Ph₂O, 240-250 °C, 5 min, 91%. c) 1.5 equiv. K₂CO₃, DMF, 120 °C, 2h then 1.5 equiv. 1-iodo-2-methylpropane, 100 °C, 4h, 29%. d) 3 equiv. H₂SO₃, MeOH, rt, 16h, 49%. e) 3.5 equiv. Boc₂O, THF, +70 °C, 24h, 96%. f) 5 equiv. KOH, dioxane/H₂O 2.5:1, rt, 16h, 99%. g) 2 equiv. PyBOP, 3 equiv. DIEA, CHCl₃, +45 °C, 16h, 71%. h) 20% TFA/CH₂Cl₂, rt, 4h, 98%.

Preparation of compounds 3 and 4



Reagents and conditions: a) 5 equiv. KOH, dioxane/H₂O 7:1, rt, 24h, 96%. b) i. 10 equiv. $(COCl)_2$, CHCl₃, rt, 2h ii. 2 equiv. compound **15**, 2 equiv. NEt₃, CHCl₃, rt, 16h, 23%. c) 1.8 equiv. compound **19**, 8 eq. PyBOP, 11 eq. DIEA, +45 °C, CHCl₃, 24h, 16%.

2. NMR experiments



Figure S1. Excerpt of the 2D ROESY plot on 400MHz ($\tau_m = 300 \text{ ms}$) at +25 °C in CDCl₃ for nonamer **4**. Details on interstrand correlations.

a)



Figure S2. Parts of the 400 MHz ¹H NMR spectra in CDCl₃ of nonamer **4** a) at 313 K; b) at 298 K; c) at 273 K; d) at 248 K; and e) at 223 K.

Multi-dimentionnal NMR experiences (HSQC, HMBC, TOCSY et ROESY) allow complete structural assignment of three oligomers.¹

• Experimental conditions:

NMR spectra were recorded on 3 different NMR spectrometers: (1) an Avance II NMR spectrometer (Bruker Biospin) with a vertical 7.05T narrow-bore/ultrashield magnet operating at 300 MHz for ¹H observation and 75 MHz for ¹³C observation by means of a 5-mm direct BBO H/X probe with Z gradient capabilities; (2) a DPX-400 NMR spectrometer (Bruker Biospin) with a vertical 9.4T narrow-bore/ultrashield magnet operating at 400 MHz for ¹H observation by means of a 5-mm direct QNP ¹H/¹³C/³¹P/¹⁹F probe with gradient capabilities; (3) an Avance III NMR spectrometer (Bruker Biospin) with a vertical 16.45T narrow-bore/ultrashield magnet operating at 700 MHz for ¹H observation by means of a 5-mm TXI ¹H/¹³C/¹⁵N probe with Z gradient capabilities. Chemical shifts are reported in parts per million (ppm, δ) relative to the ¹H residual signal of the deuterated solvent used. ¹H NMR splitting patterns with observed first-order coupling are designated as singlet (s), doublet (d), triplet (t), or quartet (q). Coupling constants (*J*) are reported in hertz. Samples were not degassed. Data processing was performed with Topspin 2.0 software.

TOCSY. Total Correlation Spectroscopy (TOCSY) experiments were recorded at 400 MHz or at 700 MHz. They were used to identify protons of oligomer that belong to the same whole spin system regardless of the exact topology with the following acquisition parameters: the acquisition was performed with $2048(t_2) \times 256(t_1)$ data points, relaxation delay of 2 s, and 32 scans per increment. Processing was done after a sine-bell multiplication in both dimensions and Fourier transformation in 1K x 1K real points.

HSQC. Heteronuclear Single-Quantum Correlation spectroscopy (HSQC) experiments were recorded at 400 MHz or at 700 MHz. They were used to observe correlations between nuclei of two different types which are separated by one bond with the following acquisition parameters: the acquisition was performed with $2048(t_2) \times 256(t_1)$ data points, relaxation delay of 2 s, and 64 scans per increment. Processing was done after a sine-bell multiplication in both dimensions and Fourier transformation in 1K x 1K real points.

HMBC. Heteronuclear Multiple Bond Correlation spectroscopy (HMBC) experiments were recorded at 400 MHz or 700 MHz. They were used to detect heteronuclear correlations over longer ranges of about 2–4 bonds with the following acquisition parameters: the acquisition was performed with $2048(t_2) \times 256(t_1)$ data points, relaxation delay of 2 s, and 64 scans per increment. Processing was done after a sine-bell multiplication in both dimensions and Fourier transformation in 1K x 1K real points.

ROESY. Rotating-frame Overhauser Spectroscopy (ROESY) experiments were recorded at 400 or 700 MHz and were used to observe dipolar interactions between protons with the following acquisition parameters: the acquisition was performed with $2048(t_2) \times 256(t_1)$ data points, in States-TPPI mode with CW-spinlock for mixing, relaxation delay of 1.5 s, and 90 scans per increment, mixing time of 300 ms. Processing was done after a sine-bell multiplication in both dimensions and Fourier transformation in 1K x 1K real points.

• <u>Nonamer, 4 :</u>



Table S1 : ¹H and ¹³C chemical shifts for nonamer **4** in CDCl₃, 400 MHz, +25°C.

Proton	¹ H Chemical Shift (ppm)	¹³ C Chemical Shift (ppm)
Hext	9.31	129.11
H _{int}	6.08	96.85
NHA	9.45	
H1	6.69	110.26
H2	6.69	118.85
H3	8.50	121.51
OiBu αH1 CH ₂	3.85	75.92
OiBu aH1 CH	2.52-2.12	28.16
OiBu aH1 CH3	1.10	19.42
NHCOB	10.23	
H4	7.51	116.17
OiBu aH4 CH2	3.89	76.61
OiBu aH4 CH	2.52-2.12	27.64
OiBu aH4 CH3	1.27	19.66
H5	8.04	117.28
OiBu aH5 CH2	4.15	77.25
OiBu aH5 CH	2.52-2.12	28.13
OiBu aH5 CH3	1.09	19.55
NHCOC	10.68	
OMe	3.83	55.59
H6	6.37	105.93
H7	9.45	110.75
H8	7.81	97.30
OiBu aH8 CH2	4.22	75.45
OiBu aH8 CH	2.52-2.12	28.47
OiBu aH8 CH3	1.27	19.66
NHCOD	10.83	
OMe	4.16	56.54
H9	7.34	107.54
H10	8.95	108.82
H11	7.08	99.81
OiBu aH11 CH2	3.97	74.80
OiBu aH11 CH	2.52-2.12	28.38
OiBu aH11 CH3	1.27	19.71
COOMe	4.09	53.03

HSQC correlation HMBC correlation	Hext	Hint	NHA	HI	H2	H3	OiBu aH1 CH2	OiBu aH1 CH	OiBu aH1 CH3	NHCOB	H4	OiBu aH4 CH2	OiBu aH4 CH	OiBu aH4 CH3	HS	OiBu aH5 CH2	OiBu aH5 CH	OiBu aH5 CH3	NHCOC	OMe	9H	LH7	H8	OiBu aH8 CH2	OiBu aH8 CH	OiBu aH8 CH3	NHCO D	OMe	H9	H10	HII	OiBu aH11 CH2	OiBu aH11 CH	OiBu aH11 CH3	COOMe
	9.31	6.08	9.45	69.9	69.9	8.50	3.85	2.52-2.12	1.10	10.23	7.51	3.89	2.52-2.12	1.27	8.04	4.15	2.52-2.12	1.09	10.68	3.83	6.37	9.45	7.81	4.22	2.52-2.12	1.27	10.83	4.16	7.34	8.95	7.08	3.97	2.52-2.12	1.27	4.09
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162.67					-	-																						┞						\vdash	
162.39																																	-		
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145.46																																			
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Table S2: HSQC and HMBC correlation for nonamer 4 in CDCl₃, 400 MHz, +25°C.

TOCSY correlation ROESY correlation TOCSY + ROES	on on Y correlation	Hext	Hint	NHA	IH	H2	H3	OiBu aH1 CH2	OiBu aH1 CH	OiBu aH1 CH3	NHCOB	H4	OiBu aH4 CH2	OiBu aH4 CH	OiBu aH4 CH3	H5	OiBu aH5 CHb	OiBu aH5 CH	OiBu aH5 CH3	NHCOc	OMe	9H	H7	H8	OiBu aH8 CHe	OiBu aH8 CH	OiBu aH8 CH ₈	NHCOD	OMe	6H	H10	TH	OiBu aH11 CHz	OiBu aH11 CH	OiBu aH11 CH3	COOMe
		9.31	6.08	9.45	69.9	69.9	8.50	3.85	2.52-2.12	1.10	10.23	7.51	3.89	2.52-2.12	1.27	8.04	4.15	2.52-2.12	1.09	10.68	3.83	6.37	9.45	7.81	4.22	2.52-2.12	1.27	10.83	4.16	7.34	8.95	7.08	3.97	2.52-2.12	1.27	4.09
Hext	9.31																																			
Hint	6.08																																			
NH _A	9.45																																			
H1	6.69																																			
H2	6.69																																			
H3	8.50																																			
OiBu aH1 CH2	3.85																																			
OiBu aH1 CH	2.52-2.12																																			
OiBu aH1 CH3	1.10																																			
NHCOB	10.23																																			
H4	7.51																																			
OiBu a H4 CH ₂	3.89																																			
OiBu aH4 CH	2.52-2.12					1	1																													
OiBu aH4 CH3	1.27					1	1																													
H5	8.04					1	1																													
OiBu aH5 CH ₂	4.15					1	1																													
OiBu aH5 CH	2.52-2.12																																			
OiBu aH5 CH3	1.09					1	1																													
NHCO _C	10.68																																			
OMe	3.83																																			
H6	6.37																																			
H7	9.45																																			
H8	7.81																																			
OiBu αH8 CH ₂	4.22																																			
OiBu aH8 CH	2.52-2.12																																			
OiBu aH8 CH3	1.27																																			
NHCOn	10.83					1	1																													
OMe	4.16					1	1																													
H9	7.34	1	1		1				1																											
H10	8.95	1	1		1				1																											
H11	7.08		1	1	1	1	1	1	1					1		1		1			1															
OiBu aH11 CH ₂	3.97			1		1	1							1		1		1			1															
OiBu aH11 CH	2.52-2.12					1	1					1		1		1																				
OiBu aH11 CH3	1.27																																			
COOMe	4.09																																			

Table S3: TOCSY and ROESY correlation for nonamer 13 in CDCl₃, 400 MHz, +25°C.

4.1 Methods for X-ray crystallography

X-ray analyses were carried out at the IECB X-ray facility (UMS 3033 CNRS) on a R-Axis Rapid Rigaku MSC with a Cu K α rotating anode and an image plate as detector at the copper K $_{\alpha}$ wavelength. Both crystals were collected at 213K and mounted on cryo-loops after quick soaking on Paratone—N oil from Hampton research before to be flash-frozen. The data were processed using the CrystalClear© suite². Both structures were solved by direct methods using SHELXD or and refined using Shelxl2013³ in the integrated WinGX system.⁴ The positions of the H atoms were deduced from coordinates of the non-H atoms and confirmed by Fourier synthesis. The non-H atoms were mostly refined with anisotropic temperature parameters. Due to the modest quality of the data (low resolution and then low ratio parameters/data) some atoms on both structures were kept with isotropic U factors. H atoms were included for structure factor calculations but not refined. The BYPASS SQUEEZE procedure was tested to take into account the electron density in the potential disordered solvent area for the crystal structure of both compounds but no improvement was observed.⁵ Finally, the non-modified structure factors were used to push the refinement as far as possible. Data statistics are shown in table S4.

4.2 Summary of X-Ray crystallographic data

Name	Pentamer, 2	Nonamer, 4							
Formula	C61.25 H74.5 N6 O16	C ₁₂₁ H ₁₄₁ Cl ₃ N ₁₄ O ₂₈							
М	1150.76	2345.82							
Crystal system	P-1	P -1							
Space group	Triclinic	Triclinic							
a/Å	10.604(2)	14.837(6)							
b/Å	23.472(5)	18.674(8)							
c/Å	27.553(6)	25.801(11)							
α/ο	77.89(3)	76.16(3)							
β/ο	83.86(3)	89.61(3)							
γ/ο	80.29(3)	72.57(3)							
U/Å ³	6591(2)	6606(5)							
Т /К	213(2)	213(2)							
Z	2	2							
ρ/g cm ⁻¹	1.160	1.179							
Size (mm)	0.1x0.05x0.01	0.2 x 0.05 x 0.05							
λ/ Å	1.54178	1.54178							
μ/mm ⁻¹	0.695	1.230							
Unique data	20949	13278							
Parameters / Restraints	59/1341	7 / 1376							
Final R indices [I>2sigma(I)]	R1 = 0.22, wR2 = 0.4951	R1 = 0.1772, wR2 = 0.4005							
Goodness of fit	1.498	1.128							

Table S4: X-ray crystallographic data.

5. Experimental section

General. All reactions were carried out under a dry nitrogen atmosphere. Commercial reagents were purchased from Sigma-Aldrich, TCI Chemicals or Alfa-Aesar and were used without further purification unless otherwise specified. Tetrahydrofurane (THF) and dichloromethane (DCM) were dried over alumina columns; chloroform, triethylamine (Et_3N) and diisopropylethylamine (DIEA) were distilled over calcium hydride (CaH₂) prior to use. Reactions were monitored by thin layer chromatography (TLC) on Merck silica gel 60-F254 plates and observed under UV light. Column chromatography purifications were carried out on Merck GEDURAN Si60 (40-63 µm). Circular chromatography purifications were carried out on Chromatotron® with silica gel, Merck grade 7749, TLC grade with binder and fluorescent indicator. NMR spectra were recorded on Bruker 300 Avance II, Bruker 400 DPX or Bruker Avance III 700 NB US. Chemical shifts are expressed in parts per million (ppm, δ) using residual solvent protons as internal standards (chloroform: δ 7.26 ppm; DMSO: δ 2.50 ppm). Coupling constants are expressed in Hertz. ESI and MALDI mass spectra were obtained on a Waters LCT Premier and a Bruker Reflex III spectrometers respectively, from the Mass Spectrometry Laboratory at the European Institute of Chemistry and Biology (UMS 3033 - IECB), Pessac, France and a Voyager DE-STR mass spectrometer from AB Sciex, Les Ulis, France. Melting points were obtained on a Büchi B-540 at the European Institute of Chemistry and Biology (UMS 3033 - IECB), Pessac, France.



Hairpin turn trimer, 1. To compound **7** (0.46 g, 1.64 mmol) dissolved in DMSO (15 mL) was added 1,5-difluoro-2,4-dinitrobenzene (0.17 g, 0.82 mmol). After stirring for 4h at room temperature, distilled DIEA (143 μ L, 0.82 mmol) was added. Then the reaction mixture was allowed to proceed at room temperature for 48h. After addition of water (30 mL), the mixture was filtered. The precipitate was solubilized in AcOEt and washed with H₂O. After evaporation, the residue was purified by chromatography (SiO₂) eluting with CH₂Cl₂/toluene (5:5, vol/vol) to obtain the product **1** as a yellowish solid (0.5 g, 86% yield).

¹H NMR (CDCl₃, 300 MHz): δ 9.70 (s, 2H), 9.32 (s, 1H), 8.04 (d, *J* = 8.5 Hz, 2H), 6.99 (s, 2H), 6.72 (dd, *J* = 8.6, 2.2 Hz, 2H), 6.68 (s, 1H), 6.60 (d, *J* = 2.2 Hz, 2H), 3.56 (d, *J* = 6.6 Hz, 4H), 2.09 (dt, *J* = 13.3, 6.7 Hz, 2H), 1.54 (s, 18H), 1.01 (d, *J* = 6.7 Hz, 12H). ¹³C NMR (CDCl₃, 75 MHz): δ 152.68, 147.78, 147.36, 131.36, 129.61, 127.20, 125.27, 118.74, 117.17, 108.28, 94.64, 80.75, 75.34, 28.49, 28.19, 19.35. HRMS (ES⁺): m/z calcd for C₃₆H₄₉N₆O₁₀ [M+H]⁺ 725.35047 found 725.35020, mp 194-195 °C.



Hairpin turn pentamer, 2. To compound **9** (0.480 g, 0.96 mmol) dissolved in DMSO (10 mL) was added 1,5-difluoro-2,4-dinitrobenzene (0.098 g, 0.48 mmol). After stirring for 4h at room temperature, distilled DIEA (80 μ L, 0.48 mmol) was added. Then the reaction mixture was allowed to proceed at room temperature for 48h. After addition of water (20 mL), the precipitate was filtered off, solubilized in AcOEt and washed with H₂O. After drying with MgSO₄ and evaporation, the residue was purified by chromatography (SiO₂) eluting with Cyclohexane/Ethyl acetate (9:1, vol/vol) to obtain the product **2** as a yellowish solid (0.404 g, 72% yield).

¹H NMR (CDCl₃, 300 MHz): δ 10.18 (s, 2H), 9.89 (s, 2H), 9.38 (s, 1H), 8.58 (d, J = 8.6 Hz, 2H), 7.77 (s, 2H), 7.40 (s, 2H), 6.93 (s, 1H), 6.85 (dd, J = 8.7, 2.1 Hz, 2H), 6.74 (d, J = 2.2 Hz, 2H), 4.39 (q, J = 7.1 Hz, 4H), 3.90 (d, J = 7.0 Hz, 4H), 3.81 (d, J = 6.4 Hz, 4H), 3.66 (d, J = 6.9 Hz, 4H), 2.18 – 1.88 (m, 6H), 1.40 (t, J = 7.2 Hz, 6H), 1.03 (d, J = 6.7 Hz, 12H), 0.94 (d, J = 6.7 Hz, 12H), 0.88 (d, J = 6.7 Hz, 12H). ¹³C NMR (CDCl₃, 75 MHz) : δ 166.35, 162.63, 152.77, 149.89, 149.12, 146.86, 133.03, 129.82, 126.87, 126.45, 125.51, 124.31, 122.03, 117.01, 116.63, 116.39, 107.98, 94.65, 75.87, 75.67, 61.55, 30.30, 29.84, 28.53, 28.03, 27.88, 27.03, 19.32, 19.24, 19.21, 14.45. HRMS (ES⁻): *m/z* calcd for C₆₂H₇₉N₆O₁₆ [M-H]⁻ 1163.5553 Found 1163.5559, mp 95-97 °C.



Hairpin turn heptamer, 3. Compound **19** (0.029 g, 0.026 mmol) was suspended in anhydrous CHCl₃ (0.5 mL). Oxalyl chloride (20 μ L, 0.26 mmol) was added and the reaction was allowed to stir at room temperature for 2h. The solvent and excess reagents were removed under vacuum and the residue was dried under vacuum for at least 2h to yield acid chloride as a white solid. To a solution of compound **14** (0.017 g, 0.057 mmol) and distilled NEt₃ (8 μ L, 0.057 mmol) in anhydrous CHCl₃ (1 mL) was added dropwise a solution of the freshly prepared diacid chloride in anhydrous CHCl₃ (1 mL) via a syringe. The reaction was allowed to proceed at room temperature for 16h. The solution was washed with H₂O, dried over Na₂SO₄, filtered, and then concentrated. The residue was purified by chromatography (SiO₂) eluting with DCM to obtain the product **3** as a yellowish solid (0.010 g, 23% yield).

¹H NMR (CDCl₃, 300 MHz): δ 10.71 (s, 2H), 10.20 (s, 2H), 9.82 (s, 2H), 9.37 (s, 1H), 9.36 (s, 2H), 8.55 (d, *J* = 8.6 Hz, 2H), 7.99 (s, 2H), 7.81 (s, 2H), 7.46 (s, 2H), 7.45 (s, 2H), 6.84 (d, *J* = 8.8 Hz, 2H), 6.80 (s, 1H), 6.75 (s, 2H), 4.06 (s, 6H), 4.06-4.04 (m, 8H), 3.99 (s, 6H), 3.98 (d, *J* = 8.7 Hz, 4H), 3.73 (d, *J* = 6.9 Hz, 4H), 2.34 – 2.19 (m, 4H), 1.14 (d, *J* = 6.7 Hz, 12H), 1.05 (d, *J* = 6.6 Hz, 12H), 0.99 (d, *J* = 6.6 Hz, 12H), 0.95 (d, *J* = 6.6 Hz, 12H). ¹³C NMR (CDCl₃, 75 MHz): δ 162.60, 162.42, 152.01, 150.95, 150.79, 149.39, 147.13, 133.17, 129.81, 129.79, 129.75, 129.64, 126.89, 126.49, 125.49, 125.38, 122.20, 117.72, 117.44, 116.86, 116.52, 110.93, 108.53, 100.56, 95.44, 75.72, 75.36, 56.29, 53.35, 28.26, 28.12, 28.05, 19.53, 19.46, 19.35, 19.31. MS(MALDI-TOF): *m/z* calcd for C₉₀H₁₀₉N₁₀O₂₂ [M+H]⁺ 1681.769 Found 1681.76, mp 247-249 °C.



Hairpin turn nonamer, 4. Compound **19** (0.100 g, 0.09 mmol) and compound **18** (0.095 g, 0.16 mmol) were dissolved in CHCl₃ (1 mL), then distilled DIEA (172 μ L, 0.99 mmol) and PyBOP (0.343 g, 0.66 mmol) were added. The reaction mixture was allowed to reach +45 °C and stirred for 24h. Then the solution was washed with H₂O and evaporated. The residue was precipitated in methanol and purified by flash chromatography (SiO₂) eluting with CH₂Cl₂/MeOH (95:5, vol/vol), and circular chromatography (SiO₂) eluting with CH₂Cl₂ to obtain product **4** as a yellowish solid (0.030 g, 16% yield).

¹H NMR (CDCl₃, 300 MHz): δ 10.83 (s, 2H), 10.68 (s, 2H), 10.23 (s, 2H), 9.45 (s, 4H), 9.31 (s, 1H), 8.95 (s, 2H), 8.50 (d, J = 9.1 Hz, 2H), 8.04 (s, 2H), 7.81 (s, 2H), 7.51 (s, 2H), 7.34 (s, 2H), 7.08 (s, 2H), 6.69 (m, 4H), 6.37 (s, 2H), 6.08 (s, 1H), 4.22 (d, J = 6.7 Hz, 4H), 4.16 (s, 6H), 4.15 (d, J = 9.1 Hz, 4H), 4.09 (s, 6H), 3.97 (d, J = 6.6 Hz, 4H), 3.89 (d, J = 6.9 Hz, 4H), 3.85 (d, J = 7.1 Hz, 4H), 3.83 (s, 6H), 2.52 – 2.12 (m, 10H), 1.27 (m, 36H), 1.10 (s br, 12H), 1.09 (s br, 12H).¹³C NMR (CDCl₃, 75 MHz): δ 166.45, 162.67, 162.59, 162.20, 161.96, 161.57, 151.59, 151.23, 150.59, 150.19, 149.36, 148.17, 147.08, 146.11, 145.46, 132.73, 129.43, 129.11, 128.73, 127.79, 126.26, 125.23, 124.82, 121.51, 118.85, 117.28, 117.16, 117.09, 116.17, 110.75, 110.26, 108.82, 107.54, 105.93, 99.81, 97.30, 96.85, 77.25, 76.61, 75.92, 75.45, 74.80, 56.54, 55.59, 53.03, 28.47, 28.38, 28.16, 28.13, 27.64, 19.71, 19.66, 19.55, 19.42. MS(MALDI-TOF): m/z calcd for C₁₂₀H₁₄₁N₁₄O₂₈ [M+H]⁺ 2226.00 Found 2225.96, degradation at 326-327 °C.



tert-butyl 2-isobutoxy-4-nitrophenylcarbamate, **7.** To 2-isobutoxy-4-nitrobenzenamine 6^6 (7.688 g, 36.6 mmol) dissolved in THF (150 mL) was added di-*tert*-butyl dicarbonate (24 g, 110 mmol). After heating for 8 days at +70 °C, the reaction mixture was evaporated. The residue was purified by chromatography (SiO₂) eluting with toluene, and precipitated in MeOH to obtain the product **7** as a yellowish solid (4.444 g, 39% yield).

¹H NMR (CDCl₃, 300 MHz): δ 8.26 (d, *J* = 9.0 Hz, 1H), 7.89 (dd, *J* = 8.9, 1.8 Hz, 1H), 7.70 (d, *J* = 1.7 Hz, 1H), 7.29 (s, 1H), 3.89 (d, *J* = 6.6 Hz, 2H), 2.20 (dt, *J* = 13.2, 6.6 Hz, 1H), 1.55 (s, 9H), 1.08 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz) : δ 152.16, 146.37, 142.26, 134.87, 117.80, 116.51, 106.18, 81.89, 75.77, 28.38, 28.18, 19.40. HRMS (ES⁻): m/z calcd for C₁₅H₂₁N₂O₅ [M-H]⁻ 309.14450 Found 309.14529, mp 98-99 °C.



tert-butyl 4-amino-2-isobutoxyphenylcarbamate, 8. To compound 7 (4.444 g, 14 mmol) dissolved in AcOEt (100 mL) was added 10% Pd/C (0.44 g). The reaction mixture was stirred for 16h under hydrogen at atmospheric pressure. After filtration through celite and concentration, the product 8 was obtained as purple oil (4 g, 99% yield) and used without further purification. ¹H NMR (CDCl₃, 300 MHz): δ 7.73 (s, 1H), 6.70 (s, 1H), 6.29-6.23 (m, 2H), 3.71 (d, *J* = 6.6 Hz, 2H), 3.49 (br, 2H), 2.12 (dt, *J* = 13.3, 6.7 Hz, 1H), 1.51 (s, 9H), 1.03 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz): δ 153.04, 148.55, 142.56, 120.08, 119.13, 106.72, 99.56, 79.45, 74.54, 28.18, 27.92, 19.04. HRMS (ES⁺): m/z calcd for C₁₅H₂₅N₂O₃ [M+H]⁺ 281.1860 Found 281.1873.



Dimer-NO₂ alkoxyl-terephtalate, **9**. 4-(ethoxycarbonyl)-2,5-diisobutoxybenzoic acid (4.390 g, 13 mmol) was suspended in anhydrous CHCl₃ (5 mL). Oxalyl chloride (5.6 mL, 65 mmol) was added and the reaction was allowed to stir at room temperature for 2h. The solvent and excess reagents were removed under vacuum and the residue was dried under vacuum for at least 2h to yield acid chloride as a white solid. To a solution of 2-isobutoxy-4-nitrobenzenamine **6**⁶ (3 g, 14.3 mmol) and distilled DIEA (2 mL, 14.3 mmol) in anhydrous CHCl₃ (13 mL) was added dropwise a solution of the freshly prepared diacid chloride in anhydrous CHCl₃ (2 mL) via a syringe. The reaction was allowed to proceed at room temperature for 16h. The solution was washed with H₂O, dried over Na₂SO₄, filtered, and then concentrated. The residue was purified by chromatography (SiO₂) eluting with DCM to obtain the product **9** as a yellowish solid (4.882 g, 71% yield).

¹H NMR (CDCl₃, 300 MHz): δ 10.44 (s, 1H), 8.77 (d, *J* = 9.0 Hz, 1H), 7.95 (dd, *J* = 9.0, 2.2 Hz, 1H), 7.78 (d, *J* = 2.5 Hz, 1H), 7.77 (s, 1H), 7.46 (s, 1H), 4.40 (q, *J* = 7.1 Hz, 2H), 3.99 (d, *J* = 7.0 Hz, 2H), 3.95 (d, *J* = 7.0 Hz, 2H), 3.85 (d, *J* = 6.4 Hz, 2H), 1.41 (t, *J* = 7.1 Hz, 3H), 1.05 (d, *J* = 6.7 Hz, 12H), 0.98 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz): δ 166.30, 163.38, 152.86, 150.01, 147.83, 143.46, 134.42, 126.05, 125.00, 119.97, 117.58, 117.28, 116.62, 106.70, 76.09, 75.93, 61.64, 31.05, 28.54, 28.09, 28.02, 19.29, 14.45. HRMS (ES⁺): m/z calcd for C₂₈H₃₉N₂O₈ [M+H]⁺



Dimer-NH₂ **alkoxyl-terephtalate, 10**. To compound **9** (6.557 g, 12 mmol) dissolved in AcOEt (130 mL) was added 10% Pd/C (0.66 g). The reaction mixture was stirred for 16h under hydrogen at atmospheric pressure. After filtration through celite and concentration, the product **10** was obtained as yellowish solid (5.95 g, 96% yield) and used without further purification.

¹H NMR (CDCl₃, 300 MHz): δ 9.94 (s, 1H), 8.22 (d, J = 8.5 Hz, 1H), 7.83 (s, 1H), 7.43 (s, 1H), 6.33 (dd, J = 8.5, 2.4 Hz, 1H), 6.29 (d, J = 2.3 Hz, 1H), 4.39 (q, J = 7.1 Hz, 2H), 3.93 (d, J = 6.9 Hz, 2H), 3.85 (d, J = 6.5 Hz, 2H), 3.76 (d, J = 6.9 Hz, 2H), 3.61 (s, 2H), 2.26 – 2.03 (m, 3H), 1.40 (t, J = 7.1 Hz, 3H), 1.04 (d, J = 6.7 Hz, 6H), 0.98 (d, J = 6.7 Hz, 12H). ¹³C NMR (CDCl3, 75 MHz): δ 166.38, 161.95, 152.72, 149.87, 149.83, 143.85, 127.17, 123.51, 122.97, 119.50, 116.84, 116.56, 106.92, 99.75, 77.11, 75.73, 75.13, 61.30, 28.41, 28.08, 27.98, 19.24, 19.21, 19.19, 14.34. HRMS (ES⁻): m/z calcd for C₂₈H₃₉N₂O₆ [M-H]⁻ 499.2798 Found 499.2705, mp 113-114 °C.



Dimethyl 2-(4-acetamido-3-methoxyphenylamino)fumarate, 12. In a 500 mL round-bottom flask, dimethyl acetylenedicarboxylate (15.8 mL, 0.12 mol) and N-(4-amino-2-methoxyphenyl)acetamide **11** (21.3 g, 0.12 mol) were dissolved in 65 mL of methanol. The reaction mixture was stirred at room temperature for 3h then heated at reflux for 1h. After cooling, the product started to precipitate. The product **12** was purified by precipitation in a minimum volume of cooled methanol and obtained with a yield of 90%.

¹H NMR (DMSO, 300 MHz): δ 9.62 (s, 1H), 9.12 (s, 1H), 7.80 (d, J = 8.5 Hz, 1H), 6.76 (d, J = 2.3 Hz, 1H), 6.43 (dd, J = 8.6, 2.3 Hz, 1H), 5.25 (s, 1H), 3.81 (s, 3H), 3.70 (s, 3H), 3.68 (s, 3H), 2.07 (s, 3H). ¹³C NMR (DMSO, 75 MHz) : δ 168.52, 168.33, 164.53, 150.21, 147.60, 136.59, 123.71, 122.57, 111.51, 104.21, 92.40, 55.65, 53.04, 51.05, 23.72. HRMS (ES⁺): *m/z* calcd for C₁₅H₁₈N₂O₆Na [M+Na]⁺ 345.1047 Found 345.1069, mp 138-139°C.



Quinolinone monomer, 13. In a 500 mL round-bottom flask, compound **12** (5 g, 15 mmol) was dissolved in 80 mL of diphenyl ether and heated with stirring at 240-250 °C for 5 minutes. After cooling, the mixture was poured into 100 mL of light petroleum ether to complete the precipitation. 5.04 g of solid was filtered off with a yield of 91% and used as product **13** without any further purification.

¹H NMR (DMSO, 300 MHz): δ 11.87 (s, 1H), 9.31 (s, 1H), 8.65 (s, 1H), 7.48 (s, 1H), 6.54 (s, 1H), 3.95 (s, 3H), 3.92 (s, 3H), 2.12 (s, 3H). ¹³C NMR (DMSO, 75 MHz) : δ 176.53, 168.66, 162.74, 153.71, 137.65, 136.60, 125.75, 120.18, 116.07, 109.78, 99.31, 56.00, 53.44, 23.95. HRMS (ES⁺): m/z calcd for C₁₄H₁₅N₂O₅ [M+H]⁺ 291.0975 Found 291.0985 and C₁₄H₁₄N₂O₅Na [M+Na]⁺ 313.0975 Found 313.0806, mp 293-294 °C.



Quinoline-NHAc monomer, 14. In a 500 mL round-bottom flask, compound **13** (5 g, 17 mmol) and potassium carbonate (3.6 g, 26 mmol) were dissolved in 50 mL of DMF and heated with stirring at +120 °C for 2h. The reaction mixture was cooled to +100 °C and 1-iodo-2-methylpropane (3 mL, 26 mmol) was added. The reaction mixture was stirred for 4h at +100 °C. After cooling, the mixture was filtered. The filtrate was extracted by CHCl₃ (3x100 mL), washed with H₂O and evaporated. Both precipitate and residue from extraction were recrystallized in hot methanol and compound **14** was obtained (1.7 g, 29%).

¹H NMR (CDCl₃, 300 MHz): δ 9.19 (s, 1H), 8.05 (s, 1H), 7.54 (s, 1H), 7.47 (s, 1H), 4.05 (s, 3H), 4.02 (s, 3H), 4.02 (d, *J* = 6.4 Hz, 2H), 2.27 (s, 3H), 1.14 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz) : δ 168.60, 166.57, 162.46, 151.11, 148.07, 146.41, 129.21, 117.91, 110.03, 107.84, 100.47, 75.13, 56.35, 28.31, 25.18, 19.42. HRMS (ES⁺): *m/z* calcd for C₁₈H₂₃N₂O₅ [M+H]⁺ 347.1601 Found 347.1611, mp 206-208 °C.



Quinoline-NH₂ monomer, 15. In a 100 mL round-bottom flask, compound **14** (1.7 g, 5 mmol) was dissolved in 75 mL of methanol and stirred until solubilization. Sulfuric acid (0.8 mL, 15 mmol) was added and the reaction mixture was stirred at room temperature for 16h. A saturated solution of sodium bicarbonate was added slowly to stop the reaction and precipitation occured. Solid was filtrered off and washed with H₂O. After drying, the product **15** was obtained (0.7 g, 49%) and used without any further purification.

¹H NMR (CDCl₃, 300 MHz): δ 7.47 (s, 1H), 7.43 (s, 1H), 7.29 (s, 1H), 4.36 (s, 2H), 4.04 (s, 3H), 3.99 (d, *J* = 6.5 Hz, 2H), 3.99 (s, 3H), 2.30 – 2.18 (m, 1H), 1.12 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz) : δ 166.90, 160.45, 151.69, 145.25, 144.66, 138.69, 118.76, 108.05, 101.14, 100.16, 74.83, 56.03, 31.09, 28.38, 19.46. HRMS (ES⁺): m/z calcd for C₁₆H₂₁N₂O₄ [M+H]⁺ 305.1496 Found 305.1518, mp 234-236 °C.



Quinoline-NHBoc monomer, 16. Compound **15** (0.346 g, 1.1 mmol) and di-tert-butyl dicarbonate (0.744 g, 3.8 mmol) were dissolved in THF (4 mL). The mixture reaction was heated for 24h at +70 $^{\circ}$ C then evaporated. The product **16** was precipitated in Et₂O and purified by chromatography (SiO₂) eluting with Cyclohexane/Ethyl Acetate (7:3, vol/vol) to obtain as a solid (0.441 g, 96% yield).

¹H NMR (CDCl₃, 300 MHz): δ 8.81 (s, 1H), 7.56 (s, 1H), 7.52 (s, 1H), 7.41 (s, 1H), 4.09 (s, 3H), 4.06 (d, *J* = 6.8 Hz, 2H), 4.05 (s, 3H), 2.35-2.26 (m, 1H), 1.61 (s, 9H), 1.17 (d, *J* = 6.7 Hz, 6H). HRMS (ES⁺): m/z calcd for C₂₁H₂₉N₂O₆ [M+H]⁺ 405.2018 Found 405.2020.



Quinoline-COOH monomer, 17. Compound **16** (2.7 g, 6.7 mmol) and potassium hydroxyde (1.9 g, 33 mmol) were dissolved in a mixture dioxane/H₂O (2.5:1). The reaction mixture was allowed to proceed at room temperature for 16h. The dioxane was evaporated and the solution acidified by a

5% citric acid solution. Compound **17** was filtered off, washed with H_2O , dried and used without any further purification (2.5 g, 99%).

¹H NMR (CDCl₃, 300 MHz): δ 8.89 (s, 1H), 8.40 (s, 1H), 7.73 (s, 1H), 7.46 (s, 1H), 4.26 (s, 3H), 4.20 (d, *J* = 6.8 Hz, 2H), 2.42-2.33 (m, 1H), 1.60 (s, 9H), 1.16 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz) : δ 166.85, 161.91, 153.72, 152.19, 149.52, 137.20, 130.83, 116.51, 107.43, 101.52, 100.32, 81.57, 57.52, 28.37, 28.04, 19.22. HRMS (ES⁺): m/z calcd for C₂₀H₂₇N₂O₆ [M+H]⁺ 391.1864 Found 391.18778, mp 207-208 °C.



Quinoline dimer, 18. Compound **17** (0.107 g, 0.27 mmol) and compound **15** (0.076 g, 0.25 mmol) were dissolved in CHCl₃ (1 mL), then distilled DIEA (130 μ L, 0.75 mmol) and PyBOP (0.260 g, 0.5 mmol) were added. The reaction mixture was allowed to reach +45 °C and stirred for 16h. Then the solution was washed with H₂O and evaporated. The residue was purified by precipitation in methanol and the quinoline dimer **18** was obtained as a solid (0.120 g, 71% yield).

¹H NMR (CDCl₃, 300 MHz): δ 11.07 (s, 1H), 9.44 (s, 1H), 8.83 (s, 1H), 7.71 (s, 1H), 7.66 (s, 1H), 7.52 (s, 1H), 7.40 (s, 1H), 7.36 (s, 1H), 4.17 (s, 3H), 4.11 (d, *J* = 10.1 Hz, 4H), 4.10 (s, 3H), 4.08 (s, 3H), 2.42 – 2.30 (m, 2H), 1.59 (s, 9H), 1.19 (d, *J* = 6.7 Hz, 6H), 1.16 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz) : δ 163.48, 162.88, 152.54, 151.41, 150.01, 145.09, 129.81, 129.63, 118.12, 110.11, 108.29, 107.94, 107.06, 100.57, 98.10, 81.17, 75.39, 56.67, 56.28, 53.21, 29.87, 28.56, 28.40, 28.33, 19.59, 19.51. HRMS (ES⁺): m/z calcd for C₃₆H₄₅N₄O₉ [M+H]⁺ 699.3175 Found 677.3184, mp 242-243 °C.



Quinoline-NH₂ **dimer, 19**. Compound **18** (0.109 g, 0.16 mmol) was dissolved in a solution 20% TFA/CH₂Cl₂ (5 mL) and stirred for 4h at room temperature. Then toluene (10 mL) was added to the

solution and evaporated. The residue was dissolved in CH_2Cl_2 (20 mL), and washed with saturated NaHCO₃, dried over Na₂SO₄, filtered and then concentrated to give compound **19** (0.091 g, 98% yield) that was used without further purification.

¹H NMR (CDCl₃, 300 MHz): δ 11.09 (s, 1H), 9.44 (s, 1H), 7.66 (s, 1H), 7.65 (s, 1H), 7.52 (s, 1H), 7.35 (s, 1H), 7.34 (s, 1H), 4.17 (s, 3H), 4.08-4.08 (m, 8H), 4.06 (d, *J* = 3.4 Hz, 2H), 2.41-2.23 (m, 2H), 1.18 (d, *J* = 6.7 Hz, 6H), 1.13 (d, *J* = 6.7 Hz, 6H). ¹³C NMR (CDCl₃, 75 MHz): δ 166.50, 163.66, 162.47, 161.00, 152.34, 151.74, 147.58, 146.42, 143.52, 138.30, 129.62, 118.54, 117.97, 109.63, 107.69, 106.98, 101.57, 100.37, 97.57, 75.20, 74.94, 56.55, 55.92, 53.23, 28.32, 28.26, 19.54, 19.40. HRMS (ES⁺): m/z calcd for C₃₁H₃₇N₄O₇ [M+H]⁺ 577.2674 Found 577.2674, mp 265-266 °C.



Hairpin turn pentamer-COOH, 20. Compound **2** (0.319 g, 0.27 mmol) and potassium hydroxyde (0.077 g, 1.37 mmol) were dissolved in a mixture dioxane/H₂O (7:1). The reaction mixture was allowed to proceed at room temperature for 24h. The dioxane was evaporated and the solution acidified to pH=1 by a HCl 1N solution. Compound **20** was filtered off, washed with H₂O, dried and used without any further purification (0.292 g, 96%).

¹H NMR (CDCl₃, 300 MHz): δ 11.09 (s, 2H), 10.19 (s, 2H), 9.89 (s, 2H), 9.38 (s, 1H), 8.53 (d, J = 8.6 Hz, 2H), 7.88 (s, 2H), 7.81 (s, 2H), 6.89 (s, 1H), 6.86 (dd, J = 8.7, 2.2 Hz, 2H), 6.75 (d, J = 2.2 Hz, 2H), 4.04 (d, J = 6.5 Hz, 4H), 3.98 (d, J = 7.1 Hz, 4H), 3.72 (d, J = 7.0 Hz, 4H), 2.27 – 1.99 (m, 6H), 1.07 (d, J = 6.7 Hz, 12H), 0.96 (d, J = 6.7 Hz, 12H), 0.92 (d, J = 6.7 Hz, 12H). HRMS (ES⁺): m/z calcd for C₅₈H₇₃N₆O₁₆ [M+H]⁺ 1109.50776 Found 1109.50859, mp 260-261°C.

6. ¹H NMR spectra of all relevant synthetic intermediates and title compounds.



































¹ For methods, see C. Dolain, A. Grélard, M. Laguerre, H. Jiang, V. Maurizot, I. Huc. *Chem. Eur. J.* **2005**, *11*, 6135-6144.

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