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## Polar solvent effects on tartaric acid binding by aromatic oligoamide foldamer capsules†

Nagula Chandramouli,<sup>a,b</sup> Mohammed Farrag El-Behairy,<sup>a,b,c</sup> Guillaume Lautrette,<sup>a,b</sup> Yann Ferrand<sup>\*a,b</sup> and Ivan Huc<sup>\*a,b</sup>

Aromatic oligoamide sequences able to fold into single helical capsules were functionalized with two types of side chains to make them soluble in various solvents such as chloroform, methanol or water and their propensity to recognize tartaric acid was evaluated. The binding affinities to tartaric acid and binding thermodynamics in different media were investigated by variable temperature <sup>1</sup>H NMR and ITC experiments, the two methods giving consistent results. We show that tartaric acid binding mainly rests on enthalpically favourable polar interactions that were found to be sufficiently strong to be effective in the presence of a polar aprotic solvent (DMSO) and even in pure methanol. Binding in water was very weak. The stronger binding interactions were found to be more susceptible to the effect of competitive solvents and compensated by unfavourable entropic effects. Thus, the best host in a less polar medium eventually was found to be the worst host in protic solvents. An interesting case of entropically driven binding was evidenced in methanol.

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### Introduction

The design of synthetic hosts that can recognize polar guests in polar media and in particular in water remains very challenging.<sup>1</sup> This contrasts with many successful examples of artificial containers able to bind apolar guests in water as, for example, molecular capsules such as cavitand-based (hemi)-carcerands,<sup>2</sup> cryptophanes,<sup>3</sup> capsules assembled through electrostatic interactions,<sup>4</sup> self-assembled metallo-organic cages,<sup>5</sup> cyclodextrins<sup>6</sup> or cyclophanes.<sup>7</sup> While the binding of non-polar molecules in water is generally favoured by a strong hydrophobic component, the binding of polar molecules is influenced by solvent effects in ways that are complex and difficult to orchestrate.<sup>1,8,9</sup> Water is very effective at interacting with polar or charged groups through hydrogen bonds and, more often than not, the enthalpic term associated with the desolvation of hosts and guests is not well compensated by host-guest interactions, resulting in weaker binding in this solvent. However, entropic components arising from the

release of bound solvent molecules upon desolvation sometimes play a favourable role.

In recent years, a novel class of synthetic receptors have emerged from artificial folded molecular strands, *i.e.* foldamers.<sup>10</sup> Foldamers are inspired from biopolymers and exploit folding as a means to create cavities to accommodate guest molecules and control the spatial organization of recognition groups towards them.<sup>11</sup> Because their chemical nature differs from that of peptides and nucleotides, foldamers may be expected to show properties and functions beyond the reach of their natural counterparts. In particular, aromatic oligoamides feature a combination of easy synthetic access, highly stable folded conformations in a wide range of solvents,<sup>12</sup> and a high predictability of their folded structures using simple computational means.<sup>13</sup>

A number of synthetic receptors have been produced based on the aromatic amide backbone.<sup>14</sup> We have been interested in an original capsule design as a novel class of helical containers, able to completely surround their guests and isolate them from the external medium. These containers consist of a helix whose diameter is narrower at each extremity and wider in the centre, creating a cavity (Fig. 1).<sup>15</sup> In these capsules, amide protons and heteroaromatic nitrogen atoms converge towards the cavity resulting in a polar environment suitable to bind to polar guests in apolar medium through multiple hydrogen bonds.<sup>16</sup> For example, we showed that sequence **1** was able to encapsulate D/L tartaric acid **5** with high affinity, selectivity and diastereoselectivity in weakly polar organic solvents.<sup>16c</sup> The lack of any gap in the helical capsule wall implies that guest

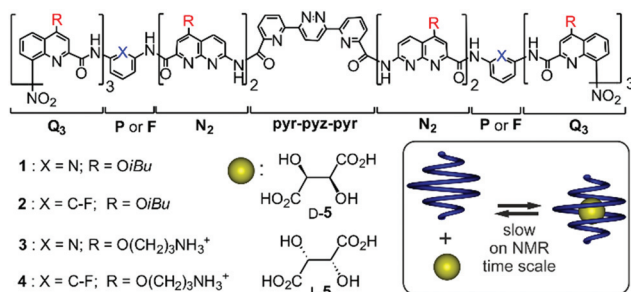
<sup>a</sup>Univ. Bordeaux, CBMN (UMR 5248), Institut Européen de Chimie Biologique, 2 rue Robert Escaipit, 33607 Pessac, France. E-mail: i.huc@iecb.u-bordeaux.fr,

y.ferrand@iecb.u-bordeaux.fr

<sup>b</sup>CNRS, CBMN (UMR 5248), Pessac, France

<sup>c</sup>Medicinal and Pharmaceutical Chemistry Department, Pharmaceutical and Drug Industries Research Division, National Research Centre, 12622 Dokki, Giza, Egypt

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**Fig. 1** (top) Formula of capsules **1**, **2**, **3** and **4**. R groups diverging from the helix and acting as solubilizing groups in either organic solvents or water are shown in red. Varying atoms pointing towards the centre of the cavity are denoted in blue; (bottom right) schematic representation of the encapsulation of a guest molecule (yellow sphere: D/L tartaric acid **5**) into a helically folded capsule (blue).

complexation and release proceed *via* partial unfolding of the helix that temporarily opens up a passage.<sup>17</sup> This mechanism was shown to be relatively slow due to the high stability of the receptor structure.

While binding is high in solvents such as CDCl<sub>3</sub>: [D<sub>6</sub>]-DMSO 90 : 10 (vol/vol), we had no information about the extent to which it would withstand the presence of protic solvents that compete for hydrogen bond formation between the host and guest. To answer this question, and to assess whether the current binding modes could be exploited for applications in water, we first had to face the challenge to synthesize analogues of **1** that would be soluble in methanol and water in the millimolar range. Henceforth, we report the successful synthesis of the first water soluble aromatic oligoamide helical foldamer capsules and the investigation of tartaric acid recognition thermodynamics in the presence of DMSO, methanol and water. We find that binding decreases in protic solvents but does not completely vanish in some cases, with *K*<sub>a</sub> values as high as 2000 L mol<sup>-1</sup> in methanol and 20 L mol<sup>-1</sup> in water. Entropy appears to play a favourable role in maintaining this binding level.

## Results and discussion

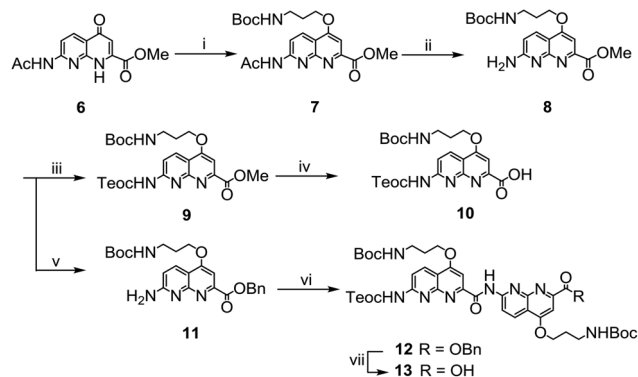
### Design and synthesis

In early foldamer capsule designs,<sup>16</sup> quinoline trimers (Q<sub>3</sub>) at each extremity of the sequences were introduced as end-caps of a helix cavity and also to prevent dimerization into double helices. Pyridine monomers (P) and naphthyridine dimers (N<sub>2</sub>) were then included in the sequences because their weaker curvature was expected to generate a cavity and because of their capacity to form hydrogen bonds, in particular, with carboxylic acids.<sup>16c,18</sup> The linear pyr-pyz-pyr central monomer was added so as to enlarge the capsule inner volume.<sup>16c</sup> This led to the design of **1**, which was shown to bind to tartaric acid with very high affinity and diastereoselectivity in organic solvents such as CDCl<sub>3</sub> containing 10% [D<sub>6</sub>]-DMSO.<sup>16c</sup> In analogue **2** (Fig. 1),

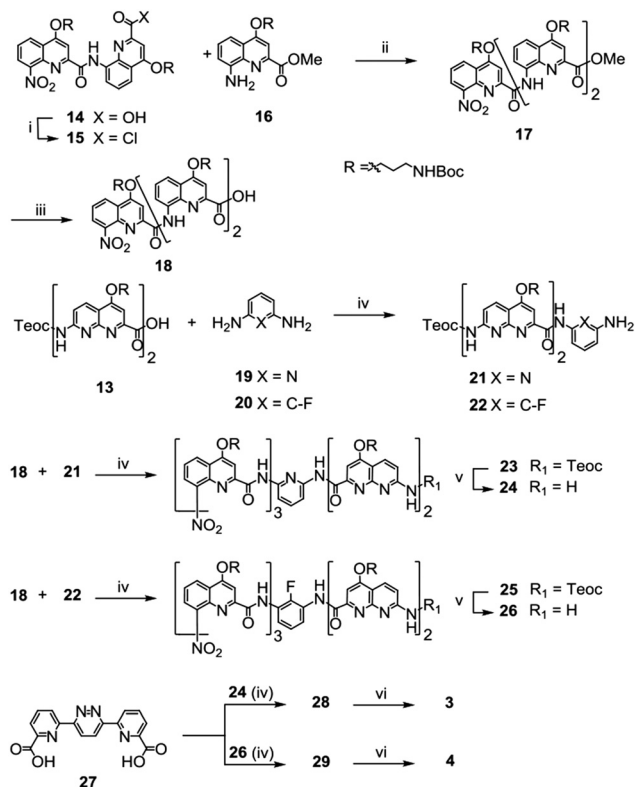
the two pyridine units of **1** have been replaced by two fluoro-benzene monomers. This sequence was produced in the context of iterative modifications of **1** aiming at improving its binding properties for other guests than tartaric acid in CDCl<sub>3</sub>. The synthesis of **2** and its binding properties for these other guests will be reported elsewhere.

To explore the extent to which molecular recognition using aromatic oligoamide capsules could be efficient in protic organic solvents or water, we considered new capsules **3** and **4** that carry water solubilising ornithine-like side chains instead of the lipophilic isobutoxy side chains of **1** and **2**. For this purpose, we prepared both the previously described 8-amino-2-quinoline-carboxylic acid monomer having a Boc protected 3-aminopropoxy side chain<sup>19</sup> and a new 7-amino-4-(3-*tert*-butoxycarbonylamino)propoxy)-1,8-naphthyridine-2-carboxylic acid monomer (Scheme 1). Their preparation first involved the introduction of a Boc protected aminopropoxy side chain on naphthyridone **6** *via* a Mitsunobu reaction (65%). Fortunately, the acetamido group of **7** could be methanolized by using a stoichiometric amount of H<sub>2</sub>SO<sub>4</sub> (0.5 equiv.) in the presence of the Boc group (66%). This reaction presumably proceeds *via* the protonation of the slightly more basic position 8 of naphthyridine which increases the electrophilicity of the acetyl group and its reactivity towards methanol while conditions remain moderately acidic, thus preventing Boc cleavage. Then, amine **8** was protected with a Teoc group (2-(trimethylsilyl) ethyl carbamate) using 4-nitrophenyl-2-(trimethylsilyl)ethyl carbonate (90%). The methyl ester of **9** was quantitatively saponified to give naphthyridine acid **10** (99%). Amino-naphthyridine benzyl ester **11** was obtained by transesterification of the methyl ester of **8** using a large excess of benzyl alcohol and base (80%).

As described earlier for organic soluble counterpart **1**,<sup>16c</sup> a hexameric hemicapsule O<sub>2</sub>N-Q<sub>3</sub>PN<sub>2</sub>-Teoc **23** bearing Boc protected aminopropoxy side chains was synthesized in a convergent manner *via* the coupling of a O<sub>2</sub>N-Q<sub>3</sub>-CO<sub>2</sub>H segment with



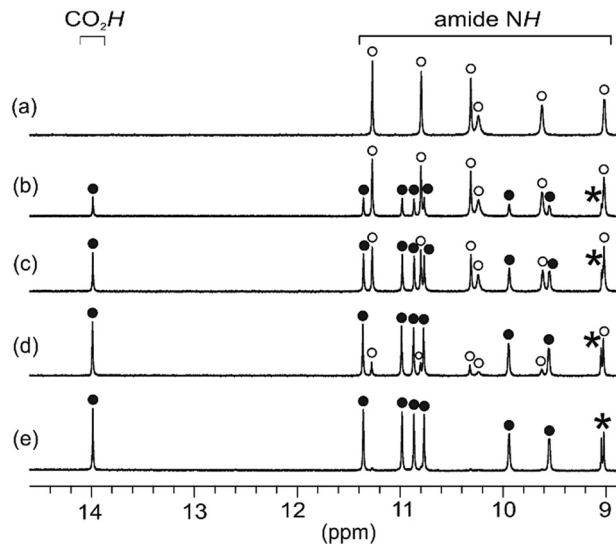
**Scheme 1** Synthesis of naphthyridine derivatives bearing protected ornithine side chains. (i) PPh<sub>3</sub>, diisopropylazodicarboxylate, *N*-(*tert*-butoxycarbonyl)-3-aminopropanol (65%); (ii) H<sub>2</sub>SO<sub>4</sub>, MeOH (66%); (iii) 4-nitrophenyl 2-(trimethylsilyl)ethyl carbonate,  $\Delta$  (90%); (iv) NaOH (99%); (v) BnOH, Et<sub>3</sub>N (80%); (vi) **10**, PyBOP, DIEA (86%); (vii) Pd/C, H<sub>2</sub> (98%).



the amino group of  $\text{H}_2\text{N-PN}_2\text{-Teoc}$  using PyBOP (Scheme 2). A second hemicapsule  $\text{O}_2\text{N-Q}_3\text{FN}_2\text{-Teoc}$  **25** was obtained using the same strategy. After removing the Teoc groups from **23** and **25** using TBAF, each amine-functionalized hexamer of **24** and **26** was reacted separately with diacid **27** using PyBOP as a coupling agent to yield Boc protected capsules **28** and **29**, respectively (Scheme 2). Side-chain deprotection was carried out with TFA in dichloromethane and the resulting water soluble oligomers **3** and **4** were purified by reversed-phase HPLC (C18) prior to physicochemical studies.

### Recognition in the presence of a polar aprotic solvent

To compare with the binding properties of **1** for *D/L* tartaric acid **5** in  $\text{CDCl}_3$ : $[\text{D}_6]$ -DMSO 90:10 (vol/vol), the efficacy of capsule **2** was assessed by  $^1\text{H}$  NMR titrations in the same medium. As previously observed for the encapsulation of *D/L*-**5** in **1**,<sup>16c</sup> the addition of the guest to capsule **2** led to the emergence of a new set of sharp signals indicating slow exchange on the NMR time scale between empty host **2** and host-guest complex **2**  $\supset$  *D/L*-**5** (Fig. 2). At low field (14 ppm), one can distinguish the sharp resonance of the hydrogen-bonded carboxylic acid protons of the guest. Through the integration of the signals of free **2** and **2**  $\supset$  *D/L*-**5** an affinity constant ( $K_a$ ) was calculated to be  $2.9 \times 10^4 \text{ L mol}^{-1}$  at 298 K in this solvent. This is considerably higher than the value previously observed



**Fig. 2** Excerpts of the 400 MHz  $^1\text{H}$  NMR spectra of capsule **2** (1 mM) at 298 K in the presence of: (a) 0 equiv., (b) 0.25 equiv., (c) 0.5 equiv., (d) 1 equiv., and (e) 2 equiv. of *D/L*-**5** in a mixture of  $\text{CDCl}_3$ : $[\text{D}_6]$ -DMSO (90:10 vol/vol). The signals of the empty host and of the host-guest complex are denoted in empty and black circles, respectively. The carboxylic acid resonance of the bound guest can be observed at low field (14 ppm). The asterisks (\*) denote aromatic resonances.

for **1** ( $5.3 \times 10^3 \text{ L mol}^{-1}$ ).<sup>16c</sup> The double **P**  $\rightarrow$  **F** mutation thus led to an increase of  $K_a$  by almost 6 fold, even though **P** units do not appear to be directly involved in tartaric acid binding in the structure of **1**  $\supset$  **5**.<sup>16c</sup> Compared to **P** units, **F** units feature an exocyclic fluorine atom pointing towards the capsule cavity that fills a small void of the structure of **1**  $\supset$  **5**. Presumably, the polarity of fluorine atoms also comes into play in a favourable way, similar to what was observed in the binding of monosaccharides.<sup>16d</sup>

However, when the proportion of  $[\text{D}_6]$ -DMSO was increased to 20% (vol/vol), association of **5** to **1** and **2** dropped to similar levels ( $K_a$  of 750 and 910  $\text{L mol}^{-1}$ , respectively, see Table 1). The addition of a polar aprotic solvent thus has a more pronounced effect on binding within **2**.

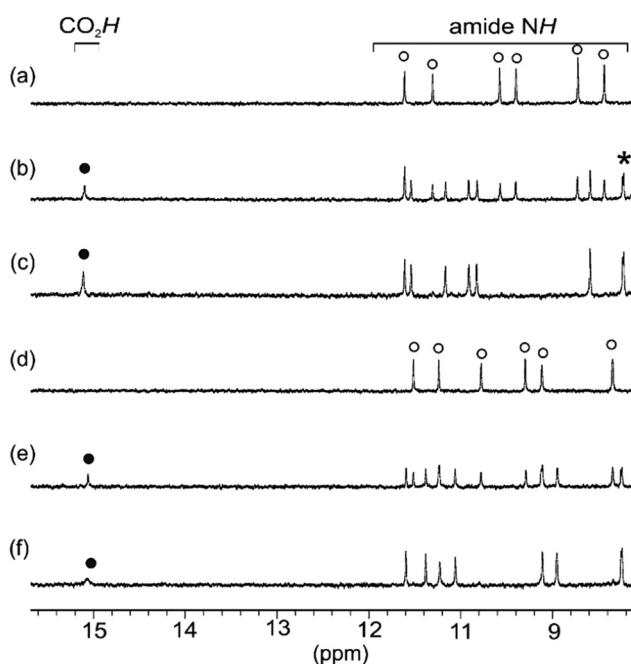
### The effect of methanol on binding

Intrigued by these results, we set to explore the behaviour of capsules **3** and **4**, the analogues of **1** and **2**, respectively, bearing charged side chains, and the extent to which slight modifications (**P** vs. **F**) of the sequence may alter or favour the binding of a guest in protic organic solvents and water. We first assessed the affinity of receptors **3** and **4** for tartaric acid in  $\text{CD}_3\text{OH}$ , a solvent that strongly competes with hydrogen bond formation between the host and guest. In this medium, both **3** and **4** exhibit good solubility. The absence of non-specific aggregation is supported by sharp  $^1\text{H}$  NMR signals. Upon increasing the concentration of **5** in a  $\text{CD}_3\text{OH}$  solution of **3** or **4**, free host resonances were again replaced by a new single set of sharp signals (Fig. 3), consistent with slow

**Table 1** Association constants ( $K_a$ ) and thermodynamic parameters for 1 : 1 host–guest complexes for caspules **1**, **2**, **3** and **4** with *D/L* tartaric acid **5** in organic solvents and water at 298 K, as measured by  $^1\text{H}$  NMR and ITC

Complex/solvent mixture	$K_a$ [ $\text{M}^{-1}$ ]	$\Delta H$ [ $\text{kJ mol}^{-1}$ ]	$T\Delta S$ [ $\text{kJ mol}^{-1}$ ]	$\Delta G$ [ $\text{kJ mol}^{-1}$ ]
5C1/ $\text{CDCl}_3$ : DMSO (9 : 1)	5300 <sup>a</sup>	n.d.	n.d.	n.d.
5C2/ $\text{CDCl}_3$ : DMSO (9 : 1)	29 000 <sup>a</sup>	n.d.	n.d.	n.d.
5C1/ $\text{CDCl}_3$ : DMSO (8 : 2)	750 <sup>a</sup>	-16.7 <sup>a</sup>	-0.5 <sup>a</sup>	-16.3 <sup>a</sup>
5C2/ $\text{CDCl}_3$ : DMSO (8 : 2)	910 <sup>a</sup>	-25.2 <sup>a</sup>	-8.6 <sup>a</sup>	-16.6 <sup>a</sup>
5C3/MeOH	2000 <sup>a</sup>	-7.3 <sup>a</sup>	11.4 <sup>a</sup>	-18.8 <sup>a</sup>
	2660 <sup>b</sup>	-9.7 <sup>b</sup>	9.9 <sup>b</sup>	-19.5 <sup>b</sup>
5C3/ $\text{H}_2\text{O}$ : $\text{D}_2\text{O}$ (9 : 1)	20 <sup>a</sup>	n.d. <sup>c</sup>	n.d. <sup>c</sup>	n.d. <sup>c</sup>
5C4/MeOH	200 <sup>a</sup>	n.d.	n.d.	n.d.
	280 <sup>b</sup>	-15.9 <sup>b</sup>	-2.0 <sup>b</sup>	-13.9 <sup>b</sup>
5C4/ $\text{H}_2\text{O}$ : $\text{D}_2\text{O}$ (9 : 1)	<1 <sup>a</sup>	n.d. <sup>c</sup>	n.d. <sup>c</sup>	n.d. <sup>c</sup>

<sup>a</sup> Measured by  $^1\text{H}$  NMR. <sup>b</sup> Measured by ITC. <sup>c</sup> Affinity is too low for accurate determination of the thermodynamic parameters; n.d.: not determined.



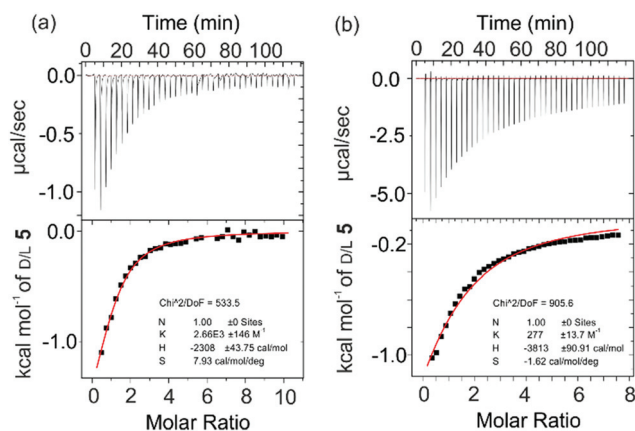
**Fig. 3** Excerpts of the 700 MHz  $^1\text{H}$  NMR spectra of capsule **3** (0.25 mM) at 298 K in the presence of: (a) 0 equiv., (b) 4.5 equiv., and (c) 30 equiv. of *D/L*-**5** in  $\text{CD}_3\text{OH}$ . Under the same conditions,  $^1\text{H}$  NMR spectra of capsule **4** in the presence of: (d) 0 equiv., (e) 15 equiv., and (f) 135 equiv. of *D/L*-**5**. The signals of the empty host and of the host–guest complex are denoted with empty and black circles, respectively. The carboxylic acid resonance of the bound guest can be observed at low field (~15 ppm, Fig. 3) though the signals were slightly broader than those for similar complexes in chloroform–DMSO mixtures.

exchange on the NMR time scale and showing that a single diastereomeric complex is formed (de > 99%) in protic solvents as well. The NMR spectra indicated that the guest carboxylic acid groups were strongly hydrogen-bonded (resonance above 15 ppm, Fig. 3) though the signals were slightly broader than those for similar complexes in chloroform–DMSO mixtures.

The association constant of capsule **3** to *D/L*-**5** in  $\text{CD}_3\text{OH}$  was calculated to be  $2000 \text{ L mol}^{-1}$  from the proportions of

bound vs. unbound species, as measured by integration of the corresponding amide NMR signals (Table 1). As could be expected, the  $K_a$  values decrease as the solvent polarity increases. Nevertheless, the decrease remained modest as  $K_a$  was lowered by a factor of only ~2.7 compared to the value measured for **1** in  $\text{CDCl}_3$  :  $[\text{D}_6]$ -DMSO 90 : 10 (vol/vol). For caspules **1** and **3**, methanol thus appears to be less detrimental to binding than DMSO, and the effect of neither polar solvents is dramatic. In contrast, the affinity of **4** towards **5** in methanol was shown to collapse to  $200 \text{ L mol}^{-1}$ , *i.e.* two orders of magnitude lower than the affinity of **2** in  $\text{CDCl}_3$  :  $[\text{D}_6]$ -DMSO 90 : 10 (vol/vol). This confirms that the effect of polar solvents on binding by **2** and **4** is higher.

In addition to NMR measurements, association constants were also determined by Isothermal Titration Calorimetry (ITC, Fig. 4). With both methods, the data gave excellent fits to a 1 : 1 binding model and the calculated  $K_a$  values were consist-



**Fig. 4** Raw data (top) and integrated heat pulse data (bottom) of ITC binding studies on: (a) capsule **3** and (b) capsule **4** with *D/L*-**5** in methanol at 298 K. Both analyses constitute a differential binding curve that may be fitted to a 1 : 1 binding model (red line) to give thermodynamic parameters.

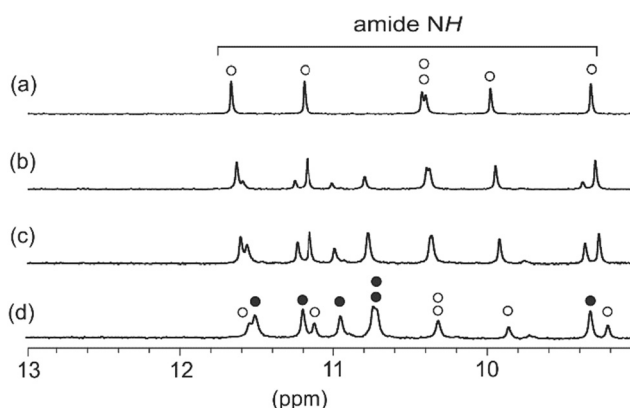
ent with the NMR data (Table 1). The differential effect of polar protic and aprotic solvents on **1** and **3** on the one hand, and **2** and **4** on the other hand, is remarkable as the only difference between the two pairs concerns the features located within the host cavities. Guest solvation/desolvation being identical, one must infer that **2** and **4** possess a more polar interior that interacts more favourably with polar molecules, that is, with tartaric acid, and also with polar solvents.

### Binding in water

Guest encapsulation within **3** and **4** was also investigated by  $^1\text{H}$  NMR titrations in water. Varying the concentration of *D/L*-**5** in a solution of receptor **3** dissolved in water led to the appearance of a new set of signals corresponding to the formation of complex **3**  $\supset$  *D/L*-**5** (Fig. 5) with complete diastereoselectivity (*de* > 99%). Yet binding was weak and saturation was not reached even in the presence of 250 mM of **5**. Furthermore, no signals could be observed at 14 ppm for the carboxylic acid group of the bound guest, possibly because of the exchange with the free guest in large excess. Integration of the  $^1\text{H}$  NMR signals of the free *vs.* bound capsules allowed us to determine  $K_a$  of around  $20\text{ L mol}^{-1}$ , about two orders of magnitude lower than that in methanol. A similar experiment conducted with receptor **4** only revealed a trace amount of the **4**  $\supset$  *D/L*-**5** complex ( $K_a < 1\text{ L mol}^{-1}$ ), again corresponding to a drop of at least two orders of magnitude with respect to methanol. Water was thus found to be highly detrimental to binding.

### Thermodynamics of the encapsulation process

To deepen our understanding of the encapsulation, we investigated the thermodynamic parameters of tartaric acid binding by capsules **1** and **2** in  $\text{CDCl}_3$  :  $[\text{D}_6]$ -DMSO 80 : 20 (vol/vol). With this proportion of DMSO, the  $K_a$  values fall in a range that can be very accurately measured using  $^1\text{H}$  NMR. The temperature was increased from 278 K to 328 K, leading to progressive



**Fig. 5** Excerpts of the 300 MHz  $^1\text{H}$  NMR spectra of capsule **3** (1 mM) at 298 K in the presence of: (a) 0 equiv., (b) 10 equiv., (c) 40 equiv., and (d) 250 equiv. of *D/L*-**5** in a  $\text{H}_2\text{O} : \text{D}_2\text{O}$  (90 : 10 vol/vol) mixture. The signals of the empty host and of the host–guest complex are denoted in empty and black circles, respectively.

host–guest complex dissociation. The linear van't Hoff plots (Fig. S1 and S2†) showed that encapsulation of **5** in both **1** and **2** is enthalpy driven (strong negative  $\Delta H$  values, Table 1). Nevertheless, similar  $K_a$  values in fact hide distinct thermodynamic parameters. The enthalpic term was found to be more favourable in the case of **2** ( $\Delta H = -25.2$ , *vs.*  $-16.7\text{ kJ mol}^{-1}$  for **1**), revealing stronger, presumably polar, interactions with tartaric acid. This is consistent, with the very high binding observed upon decreasing the proportion of DMSO down to 10%, and also with the overall higher effects of polar solvents on binding to **2** as opposed to **1**. As the cavity of **2** is more polar, not only tartaric acid binding but also competitive solvent binding is more efficient. Not surprisingly, the large enthalpic term of binding by **2** is compensated by an unfavourable entropic term ( $-T\Delta S = 8.6\text{ kJ mol}^{-1}$  at 298 K). In contrast, the binding entropy by sequence **1** is very small. The enthalpy–entropy compensation observed for **2** is typical<sup>18,20</sup> and reflects the fact that for all favourable interactions to be established upon binding tartaric acid in the cavity of **2**, the complex geometry has to be more restricted than upon binding to **1**.

We also investigated the thermodynamics of tartaric acid encapsulation by capsules **3** and **4** in methanol. In this solvent, the binding constants were sufficiently high to use ITC. In the case of **3**, the thermodynamic parameters extracted from ITC were corroborated by a van't Hoff plot of variable temperature NMR data (Table 1, Fig. S3†). Similar to what was observed in chloroform–DMSO for capsules **1** and **2**, binding in methanol proceeded with favourable enthalpy, this term being more favourable in the case of **4** ( $\Delta H = -15.9\text{ kJ mol}^{-1}$ ) than that for **3** ( $-9.7\text{ kJ mol}^{-1}$ ). However, both terms were significantly smaller than those in chloroform–DMSO mixtures, reflecting the better ability of methanol to solvate both the host and guest and the overall weaker binding in this solvent. The values of entropic terms in methanol for **3** and **4** differed significantly from those observed in chloroform–DMSO. Indeed, the binding of tartaric acid to capsule **3** was found to have a strongly favourable entropic term ( $-T\Delta S = -9.9\text{ kJ mol}^{-1}$  at 298 K). This effect appeared to be characteristic of the solvent and not of a change in the binding itself, and was assigned to the favorable release of encapsulated methanol molecules from the helical capsule upon binding tartaric acid. Encapsulation by **3** in methanol is thus both enthalpically and entropically driven. The favorable entropic solvent effects also operated for capsule **4** but, since the binding entropy was inherently less favorable, as explained above for **2**, the outcome was a weakly unfavourable entropic effect ( $-T\Delta S = 2.0\text{ kJ mol}^{-1}$  at 298 K).

Similar experiments using ITC or NMR could not be conducted in water due to the low affinity of both receptors **3** and **4** in this medium. By extrapolation of the results observed in methanol, one may expect the enthalpic term to further decrease due to competitive binding of both hosts and guests by the solvent. If any solvent-derived favourable entropic effect occurs in water, as for methanol, it is too weak to compensate for the weak enthalpy.

## Conclusions

In conclusion, we have shown that tartaric acid binding by two related foldamer sequences is strong enough to be observed in polar and in protic media such as methanol, or chloroform–DMSO mixtures. Binding of this polar guest in water was considerably weaker. Sequences **2** and **4** show stronger interactions with tartaric acid than sequences **1** and **3**, as expressed by a more negative binding enthalpy. These stronger, presumably more polar, interactions lead to higher binding when decreasing the solvent polarity. However, the same interactions are more susceptible to competitive binding by polar solvents, eventually leading to lower affinity in protic medium. To achieve binding in water, one may thus not rely only on polar groups and hydrogen bonding. In a classical manner, stronger interactions were found to be partly compensated by unfavourable entropic effects.

An interesting case of solvent-induced entropically driven binding in methanol was observed. One may expect that such favourable solvent effects become stronger for larger capsule designs whose cavity would release multiple solvent molecules upon guest binding. Currently efforts are being made in this direction in our laboratories and will be reported in due course.

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## Notes and references

- (a) E. A. Kataev and C. Müller, *Tetrahedron*, 2014, **70**, 137; (b) A. Dalla Cort, G. Forte and L. Schiaffino, *J. Org. Chem.*, 2011, **76**, 7569; (c) A. P. Davis, *Nature*, 2010, **464**, 169; (d) S. Kubik, *Chem. Soc. Rev.*, 2008, **38**, 585; (e) G. V. Oshovsky, D. N. Reinhoudt and W. Verboom, *Angew. Chem., Int. Ed.*, 2007, **46**, 2366; (f) E. A. Meyer, R. K. Castellano and F. Diederich, *Angew. Chem., Int. Ed.*, 2003, **42**, 1210.
- (a) S. M. Liu, H. Y. Gan, A. T. Hermann, S. W. Rick and B. C. Gibb, *Nat. Chem.*, 2010, **2**, 847; (b) C. L. D. Gibb and B. C. Gibb, *J. Am. Chem. Soc.*, 2004, **126**, 11408; (c) L. S. Kaanumalle, C. L. D. Gibb, B. C. Gibb and V. Ramamurthy, *J. Am. Chem. Soc.*, 2005, **127**, 3674; (d) L. Trembleau and J. Rebek Jr., *Science*, 2003, **301**, 1219.
- (a) G. Huber, T. Brotin, L. Dubois, H. Desvaux, J. P. Dutasta and P. Berthault, *J. Am. Chem. Soc.*, 2006, **128**, 6239; (b) C. Hilty, T. J. Lowery, D. E. Wemmer and A. Pines, *Angew. Chem., Int. Ed.*, 2006, **45**, 70; (c) E. L. Piatnitski, R. A. Flowers and K. Deshayes, *Chem. – Eur. J.*, 2000, **6**, 999.
- (a) F. Corbellini, L. Di Costanzo, M. Crego-Calama, S. Geremia and D. N. Reinhoudt, *J. Am. Chem. Soc.*, 2003, **125**, 9946; (b) F. Corbellini, R. Fiammengo, P. Timmerman, M. Crego-Calama, K. Versluis, A. J. R. Heck, I. Luyten and D. N. Reinhoudt, *J. Am. Chem. Soc.*, 2002, **124**, 6569.
- (a) A. V. Davis and K. N. Raymond, *J. Am. Chem. Soc.*, 2005, **127**, 7912; (b) C. Sgarlata, J. S. Mugridge, M. D. Pluth, B. E. F. Tiedemann, V. Zito, G. Arena and K. N. Raymond, *J. Am. Chem. Soc.*, 2010, **132**, 1005; (c) M. Yoshizawa, J. Nakagawa, K. Kumazawa, M. Nagao, M. Kawano, T. Ozeki and M. Fujita, *Angew. Chem., Int. Ed.*, 2005, **44**, 1810; (d) T. Sawada, M. Yoshizawa, S. Sato and M. Fujita, *Nat. Chem.*, 2009, **1**, 53; (e) J. L. Bolliger, T. K. Ronson, M. Ogawa and J. R. Nitschke, *J. Am. Chem. Soc.*, 2014, **136**, 14545; (f) P. Mal, D. Schultz, K. Beyeh, K. Rissanen and J. R. Nitschke, *Angew. Chem., Int. Ed.*, 2008, **47**, 8297; (g) M. Yamashina, Y. Sei, M. Akita and M. Yoshizawa, *Nat. Commun.*, 2014, **5**, 4662.
- M. Rekharsky and Y. Inoue, *J. Am. Chem. Soc.*, 2000, **122**, 10949.
- (a) A. McCurdy, L. Jimenez, D. A. Stauffer and D. A. Dougherty, *J. Am. Chem. Soc.*, 1992, **114**, 10314; (b) S. B. Ferguson, E. M. Sanford, E. M. Seward and F. Diederich, *J. Am. Chem. Soc.*, 1991, **113**, 5410.
- K. N. Houk, A. G. Leach, S. P. Kim and X. Y. Zhang, *Angew. Chem., Int. Ed.*, 2003, **42**, 4872.
- E. Klein, Y. Ferrand, N. P. Barwell and A. P. Davis, *Angew. Chem., Int. Ed.*, 2008, **47**, 2693.
- (a) S. H. Gellman, *Acc. Chem. Res.*, 1998, **31**, 173; (b) G. Guichard and I. Huc, *Chem. Commun.*, 2011, **47**, 5933.
- (a) D.-W. Zhang, X. Zhao, J.-L. Hou and Z.-T. Li, *Chem. Rev.*, 2012, **112**, 5271; (b) I. Saraogi and A. D. Hamilton, *Chem. Soc. Rev.*, 2009, **38**, 1726; (c) I. Huc, *Eur. J. Org. Chem.*, 2004, **17**; (d) T. Nishinaga, A. Tanatani, K. Oh and J. S. Moore, *J. Am. Chem. Soc.*, 2002, **124**, 5934; (e) A. Tanatani, T. S. Hughes and J. S. Moore, *Angew. Chem., Int. Ed.*, 2002, **41**, 325; (f) K.-J. Chang, B.-N. Kang, M.-H. Lee and K.-S. Jeong, *J. Am. Chem. Soc.*, 2005, **127**, 12214; (g) M. Waki, H. Abe and M. Inouye, *Angew. Chem., Int. Ed.*, 2007, **46**, 3059; (h) H. Abe, H. Machiguchi, S. Matsumoto and M. Inouye, *J. Org. Chem.*, 2008, **73**, 4650; (i) Y. Hua and A. H. Flood, *J. Am. Chem. Soc.*, 2010, **132**, 12838; (j) Y. Wang, F. Bie and H. Jiang, *Org. Lett.*, 2010, **12**, 3630.
- T. Qi, V. Maurizot, H. Noguchi, T. Charoenraks, B. Kauffmann, M. Takafuji, H. Ihara and I. Huc, *Chem. Commun.*, 2012, **48**, 6337.
- (a) A. M. Abramyan, Z. Liu and V. Pophristic, *Chem. Commun.*, 2016, **52**, 669; (b) Z. Liu, A. M. Abramyan and V. Pophristic, *New J. Chem.*, 2015, **39**, 3229.
- (a) B. Gong, H. Zeng, J. Zhu, L. Yuan, Y. Han, S. Cheng, M. Furukawa, R. D. Parra, A. Y. Kovalevsky, J. L. Mills, E. Skrzypczak-Jankun, S. Martinovic, R. D. Smith, C. Zheng, T. Szyperski and X. Cheng Zeng, *Proc. Natl. Acad.*

- Sci. U. S. A.*, 2002, **99**, 11583; (b) J.-L. Hou, X.-B. Shao, G.-J. Chen, Y.-X. Zhou, X.-K. Jiang and Z.-T. Li, *J. Am. Chem. Soc.*, 2004, **126**, 12386; (c) C. Li, G.-T. Wang, H.-P. Yi, X.-K. Jiang, Z.-T. Li and R.-X. Wang, *Org. Lett.*, 2007, **9**, 1797; (d) Q. Gan, Y. Ferrand, C. Bao, B. Kauffmann, A. Grélard, H. Jiang and I. Huc, *Science*, 2011, **331**, 1172; (e) Y. Ferrand, Q. Gan, B. Kauffmann, H. Jiang and I. Huc, *Angew. Chem., Int. Ed.*, 2011, **50**, 7572; (f) L.-Y. You, S.-G. Chen, X. Zhao, Y. Liu, W.-X. Lan, Y. Zhang, H.-J. Lu, C.-Y. Cao and Z.-T. Li, *Angew. Chem., Int. Ed.*, 2012, **51**, 1657; (g) Q. Gan, Y. Ferrand, N. Chandramouli, B. Kauffmann, C. Aube, D. Dubreuil and I. Huc, *J. Am. Chem. Soc.*, 2012, **134**, 15656.
- 15 (a) J. Garric, J.-M. Léger and I. Huc, *Angew. Chem., Int. Ed.*, 2005, **44**, 1954; (b) Y. Hua, Y. Liu, C.-H. Chen and A. H. Flood, *J. Am. Chem. Soc.*, 2013, **135**, 14401.
- 16 (a) C. Bao, B. Kauffmann, Q. Gan, K. Srinivas, H. Jiang and I. Huc, *Angew. Chem., Int. Ed.*, 2008, **47**, 4153; (b) C. Bao, Q. Gan, B. Kauffmann, H. Jiang and I. Huc, *Chem. – Eur. J.*, 2009, **15**, 11530; (c) Y. Ferrand, A. M. Kendhale, B. Kauffmann, A. Grélard, C. Marie, V. Blot, M. Pipelier, D. Dubreuil and I. Huc, *J. Am. Chem. Soc.*, 2010, **132**, 7858; (d) N. Chandramouli, Y. Ferrand, G. Lautrette, B. Kauffmann, C. D. Mackereth, M. Laguerre, D. Dubreuil and I. Huc, *Nat. Chem.*, 2015, **7**, 334.
- 17 (a) Y. Ferrand, N. Chandramouli, A. M. Kendhale, C. Aube, B. Kauffmann, A. Grélard, M. Laguerre, D. Dubreuil and I. Huc, *J. Am. Chem. Soc.*, 2012, **134**, 11282; (b) A. M. Abramyan, Z. Liu and V. Pophristic, *Phys. Chem. Chem. Phys.*, 2014, **16**, 20406.
- 18 F. Garcia-Tellado, S. J. Geib, S. Goswami and A. D. Hamilton, *J. Am. Chem. Soc.*, 1991, **113**, 9265.
- 19 E. Gillies, F. Deiss, C. Staedel, J.-M. Schmitter and I. Huc, *Angew. Chem., Int. Ed.*, 2007, **46**, 4081.
- 20 C.-E. Chang and M. K. Gilson, *J. Am. Chem. Soc.*, 2004, **126**, 13156.